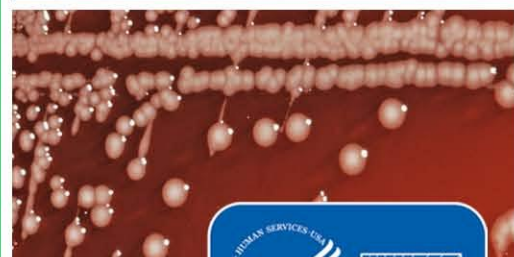
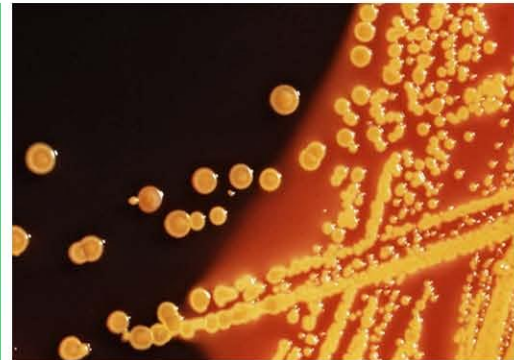


**N
A
R
M
S**

National Antimicrobial Resistance Monitoring System: Enteric Bacteria

2010

Human Isolates Final Report



National Center for Emerging and Zoonotic Infectious Diseases
Division of Foodborne, Waterborne, and Environmental Diseases



Table of Contents

List of Tables.....	2
List of Figures	5
List of Boxes	6
List of Abbreviations and Acronyms.....	7
NARMS Working Group	8
Introduction.....	11
What is New in the NARMS Report for 2010	12
Summary of NARMS 2010 Surveillance Data	13
Antimicrobial Resistance: 1996–2010	15
WHO Categorization of Antimicrobial Agents	21
Surveillance and Laboratory Testing Methods	22
Results	32
1. Non-typhoidal <i>Salmonella</i>	32
A. <i>Salmonella</i> ser. Enteritidis	35
B. <i>Salmonella</i> ser. Typhimurium	37
C. <i>Salmonella</i> ser. Newport	39
D. <i>Salmonella</i> ser. Heidelberg	41
E. <i>Salmonella</i> ser. I 4,[5],12:i:-	43
2. Typhoidal <i>Salmonella</i>	46
A. <i>Salmonella</i> ser. Typhi	46
B. <i>Salmonella</i> ser. Paratyphi A, Paratyphi B, and Paratyphi C	48
3. <i>Shigella</i>	50
4. <i>Escherichia coli</i> O157.....	57
5. <i>Campylobacter</i>	59
6. <i>Vibrio</i> species other than <i>V.cholerae</i>	63
References	66
NARMS Publications in 2010.....	67
Appendix A.....	68
Appendix B.....	73

Suggested Citation: CDC. National Antimicrobial Resistance Monitoring System for Enteric Bacteria (NARMS): Human Isolates Final Report, 2010. Atlanta, Georgia: U.S. Department of Health and Human Services, CDC, 2012.

Information Available Online: Previous reports and additional information about NARMS are posted on the CDC NARMS website: <http://www.cdc.gov/narms>

Disclaimer: Commercial products are mentioned for identification only and do not represent endorsement by the Centers for Disease Control and Prevention or the U. S. Department of Health and Human Services.

List of Tables

Table 1.	WHO categorization of antimicrobials of critical importance to human medicine.....	21
Table 2.	Population size and number of isolates received and tested, NARMS, 2010	23
Table 3.	Antimicrobial agents used for susceptibility testing for <i>Salmonella</i> , <i>Shigella</i> , and <i>Escherichia coli</i> O157 isolates, NARMS, 2010	25
Table 4.	Antimicrobial agents used for susceptibility testing of <i>Campylobacter</i> isolates, NARMS, 1997–2010.....	27
Table 5.	Antimicrobial agents used for susceptibility testing of <i>Vibrio</i> species other than <i>V. cholerae</i> isolates, NARMS, 2009	28
Table 6.	Number and percentage of isolates with resistance to at least ACSSuT, ACSSuTAuCx, nalidixic acid, and ceftriaxone among the 20 most common non-typhoidal <i>Salmonella</i> serotypes isolated in NARMS, 2010	32
Table 7.	Minimum inhibitory concentrations (MICs) and resistance of non-typhoidal <i>Salmonella</i> isolates to antimicrobial agents, 2010 (N=2474).....	33
Table 8.	Percentage and number of non-typhoidal <i>Salmonella</i> isolates resistant to antimicrobial agents, 2001–2010	34
Table 9.	Resistance patterns of non-typhoidal <i>Salmonella</i> isolates, 2001–2010	34
Table 10.	Minimum inhibitory concentrations (MICs) and resistance of <i>Salmonella</i> ser. Enteritidis isolates to antimicrobial agents, 2010 (N=522).....	35
Table 11.	Percentage and number of <i>Salmonella</i> ser. Enteritidis isolates resistant to antimicrobial agents, 2001–2010	36
Table 12.	Resistance patterns of <i>Salmonella</i> ser. Enteritidis isolates, 2001–2010.....	37
Table 13.	Minimum inhibitory concentrations (MICs) and resistance of <i>Salmonella</i> ser. Typhimurium isolates to antimicrobial agents, 2010 (N=366)	37
Table 14.	Percentage and number of <i>Salmonella</i> ser. Typhimurium isolates resistant to antimicrobial agents, 2001–2010	38
Table 15.	Resistance patterns of <i>Salmonella</i> ser. Typhimurium isolates, 2001–2010	39
Table 16.	Minimum inhibitory concentrations (MICs) and resistance of <i>Salmonella</i> ser. Newport isolates to antimicrobial agents, 2010 (N=305).....	39
Table 17.	Percentage and number of <i>Salmonella</i> ser. Newport isolates resistant to antimicrobial agents, 2001–2010.....	40
Table 18.	Resistance patterns of <i>Salmonella</i> ser. Newport isolates, 2001–2010.....	41
Table 19.	Minimum inhibitory concentrations (MICs) and resistance of <i>Salmonella</i> ser. Heidelberg isolates to antimicrobial agents, 2010 (N=62)	41
Table 20.	Percentage and number of <i>Salmonella</i> ser. Heidelberg isolates resistant to antimicrobial agents, 2001–2010	42
Table 21.	Resistance patterns of <i>Salmonella</i> ser. Heidelberg isolates, 2001–2010.....	43
Table 22.	Minimum inhibitory concentrations (MICs) and resistance of <i>Salmonella</i> ser. I 4,[5],12:i:- isolates to antimicrobial agents, 2010 (N=77)	43
Table 23.	Percentage and number of <i>Salmonella</i> ser. I 4,[5],12:i:- isolates resistant to antimicrobial agents, 2001–2010	44
Table 24.	Resistance patterns of <i>Salmonella</i> ser. I 4,[5],12:i:- isolates, 2001–2010	45
Table 25.	Minimum inhibitory concentrations (MICs) and resistance of <i>Salmonella</i> ser. Typhi isolates to antimicrobial agents, 2010 (N=444)	46

Table 26. Percentage and number of <i>Salmonella</i> ser. Typhi isolates resistant to antimicrobial agents, 2001–2010.....	47
Table 27. Resistance patterns of <i>Salmonella</i> ser. Typhi isolates, 2001–2010.....	47
Table 28. Frequency of <i>Salmonella</i> ser. Paratyphi A, Paratyphi B, and Paratyphi C, 2010.....	48
Table 29. Minimum inhibitory concentrations (MICs) and resistance of <i>Salmonella</i> ser. Paratyphi A, Paratyphi B, and Paratyphi C isolates to antimicrobial agents, 2010 (N=146).....	48
Table 30. Percentage and number of <i>Salmonella</i> ser. Paratyphi A, Paratyphi B, and Paratyphi C isolates resistant to antimicrobial agents, 2001–2010.....	49
Table 31. Resistance patterns of <i>Salmonella</i> ser. Paratyphi A, Paratyphi B, and Paratyphi C isolates, 2001–2010.....	49
Table 32. Frequency of <i>Shigella</i> species, 2010.....	50
Table 33. Minimum inhibitory concentrations (MICs) and resistance of <i>Shigella</i> isolates to antimicrobial agents, 2010 (N=407).....	50
Table 34. Percentage and number of <i>Shigella</i> isolates resistant to antimicrobial agents, 2001–2010.....	51
Table 35. Resistance patterns of <i>Shigella</i> isolates, 2001–2010.....	52
Table 36. Minimum inhibitory concentrations (MICs) and resistance of <i>Shigella sonnei</i> isolates to antimicrobial agents, 2010 (N=333).....	52
Table 37. Percentage and number of <i>Shigella sonnei</i> isolates resistant to antimicrobial agents, 2001–2010.....	53
Table 38. Resistance patterns of <i>Shigella sonnei</i> isolates, 2001–2010.....	54
Table 39. Minimum inhibitory concentrations and resistance of <i>Shigella flexneri</i> isolates to antimicrobial agents, 2010 (N=60).....	54
Table 40. Percentage and number of <i>Shigella flexneri</i> isolates resistant to antimicrobial agents, 2001–2010.....	55
Table 41. Resistance patterns of <i>Shigella flexneri</i> isolates, 2001–2010.....	56
Table 42. Minimum inhibitory concentrations (MICs) and resistance of <i>Escherichia coli</i> O157 isolates to antimicrobial agents, 2010 (N=167).....	57
Table 43. Percentage and number of <i>Escherichia coli</i> O157 isolates resistant to antimicrobial agents, 2001–2010.....	58
Table 44. Resistance patterns of <i>Escherichia coli</i> O157 isolates, 2001–2010.....	58
Table 45. Frequency of <i>Campylobacter</i> species, 2010.....	59
Table 46. Minimum inhibition concentrations (MICs) and resistance of <i>Campylobacter</i> isolates to antimicrobial agents, 2010 (N=1310).....	59
Table 47. Percentage and number of <i>Campylobacter</i> isolates resistant to antimicrobial agents, 2001–2010.....	60
Table 48. Resistance patterns of <i>Campylobacter</i> isolates, 2001–2010.....	60
Table 49. Minimum inhibitory concentrations (MICs) and resistance of <i>Campylobacter jejuni</i> isolates to antimicrobial agents, 2010 (N=1158).....	61
Table 50. Percentage and number of <i>Campylobacter jejuni</i> isolates resistant to antimicrobial agents, 2001–2010.....	61
Table 51. Minimum inhibitory concentrations (MICs) and resistance of <i>Campylobacter coli</i> isolates to antimicrobial agents, 2010 (N=115).....	62
Table 52. Percentage and number of <i>Campylobacter coli</i> isolates resistant to antimicrobial agents, 2001–2010.....	62
Table 53. Frequency of <i>Vibrio</i> species other than <i>V. cholerae</i> , 2009.....	63

Table 54. Minimum inhibition concentrations (MICs) and resistance of isolates of <i>Vibrio</i> species other than <i>V. cholerae</i> to antimicrobial agents, 2009 (N=275)	63
Table 55. Percentage and number of isolates of <i>Vibrio</i> species other than <i>V. cholerae</i>, by ampicillin MIC interpretation, 2009	63
Appendix A, Table 1. Non-typhoidal <i>Salmonella</i> outbreaks caused by antimicrobial resistant isolates (N=18), 2004-2008	70
Appendix A, Table 2. Non-typhoidal <i>Salmonella</i> outbreaks caused by isolates with no resistance detected (N=85), 2004-2008	71
Appendix A, Table 3. Number and percent of outbreaks caused by antimicrobial resistant non-typhoidal <i>Salmonella</i>, by agent and food commodity group (N=18), 2004-2008	72
Appendix A, Table 4. Antimicrobial resistance patterns of non-typhoidal <i>Salmonella</i> outbreak isolates, by commodity group (N=103), 2004-2008	72
Appendix B, Table 1. Unlikely or discordant resistance phenotypes	73
Appendix B, Table 2. Uncommon resistance phenotypes	74

List of Figures

Figure 1.	Percentage of non-typhoidal <i>Salmonella</i> isolates resistant to nalidixic acid, by year, 1996–2010.....	15
Figure 2.	Percentage of non-typhoidal <i>Salmonella</i> isolates resistant to ceftriaxone, by year, 1996–2010.....	16
Figure 3.	Percentage of <i>Salmonella</i> ser. Enteritidis isolates resistant to nalidixic acid, by year, 1996–2010.....	16
Figure 4.	Percentage of <i>Salmonella</i> ser. Heidelberg isolates resistant to ceftriaxone, by year, 1996–2010.....	17
Figure 5.	Percentage of <i>Salmonella</i> ser. Typhimurium isolates resistant to at least ampicillin, chloramphenicol, streptomycin, sulfonamide, and tetracycline (ACSSuT), by year, 1996–2010.....	17
Figure 6.	Percentage of <i>Salmonella</i> ser. Newport isolates resistant to at least ampicillin, chloramphenicol, streptomycin, sulfonamide, tetracycline, amoxicillin-clavulanic acid, and ceftriaxone (ACSSuTAuCx), by year, 1996–2010	18
Figure 7.	Percentage of non-typhoidal <i>Salmonella</i> isolates resistant to 1 or more antimicrobial classes, by year, 1996–2010.....	18
Figure 8.	Percentage of non-typhoidal <i>Salmonella</i> isolates resistant to 3 or more antimicrobial classes, by year, 1996–2010.....	19
Figure 9.	Percentage of <i>Salmonella</i> ser. Typhi isolates resistant to nalidixic acid, by year, 1999–2010.....	19
Figure 10.	Percentage of <i>Campylobacter</i> isolates resistant to ciprofloxacin, by year, 1997–2010	20
Figure 11.	How to read a squashtogram	30
Figure 12.	Proportional chart, a categorical graph of a squashtogram.....	31
Figure 13.	Antimicrobial resistance pattern for non-typhoidal <i>Salmonella</i> , 2010	33
Figure 14.	Antimicrobial resistance pattern for <i>Salmonella</i> ser. Enteritidis, 2010	35
Figure 15.	Antimicrobial resistance pattern for <i>Salmonella</i> ser. Typhimurium, 2010.....	38
Figure 16.	Antimicrobial resistance pattern for <i>Salmonella</i> ser. Newport, 2010	40
Figure 17.	Antimicrobial resistance pattern for <i>Salmonella</i> ser. Heidelberg, 2010	42
Figure 18.	Antimicrobial resistance pattern for <i>Salmonella</i> ser. I 4,[5],12:i:-, 2010.....	44
Figure 19.	Antimicrobial resistance pattern for <i>Salmonella</i> ser. Typhi, 2010	46
Figure 20.	Antimicrobial resistance pattern for <i>Salmonella</i> ser. Paratyphi A, Paratyphi B, and Paratyphi C, 2010.....	48
Figure 21.	Antimicrobial resistance pattern for <i>Shigella</i> , 2010.....	51
Figure 22.	Antimicrobial resistance pattern for <i>Shigella sonnei</i> , 2010.....	53
Figure 23.	Antimicrobial resistance pattern for <i>Shigella flexneri</i> , 2010.....	55
Figure 24.	Antimicrobial resistance pattern for <i>Escherichia coli</i> O157, 2010	57
Figure 25.	Antimicrobial resistance pattern for <i>Campylobacter</i> , 2010	59
Figure 26.	Antimicrobial resistance pattern for <i>Campylobacter jejuni</i> , 2010.....	61
Figure 27.	Antimicrobial resistance pattern for <i>Campylobacter coli</i> , 2010	62
Figure 28.	Antibiotic resistance pattern for <i>Vibrio</i> species other than <i>V. cholerae</i> , 2009	63

List of Boxes

Box 1. Changes in antimicrobial resistance: 2010 vs. 2003–07.....	64
Box 2. Ciprofloxacin breakpoint changes for <i>Salmonella</i>	65

List of Abbreviations and Acronyms

ACSSuT	Resistance to ampicillin, chloramphenicol, streptomycin, sulfamethoxazole/sulfisoxazole, and tetracycline
ACSSuTAuCx	Resistance to ampicillin, chloramphenicol, streptomycin, sulfamethoxazole/sulfisoxazole, tetracycline, amoxicillin-clavulanic acid, and ceftriaxone
ACT/S	Resistance to at least ampicillin, chloramphenicol, and trimethoprim-sulfamethoxazole
ANT/S	Resistance to at least ampicillin, nalidixic acid and trimethoprim-sulfamethoxazole
AT/S	Resistance to at least ampicillin and trimethoprim-sulfamethoxazole
CDC	Centers for Disease Control and Prevention
CI	Confidence interval
CLSI	Clinical and Laboratory Standards Institute
EIP	Emerging Infections Program
ELC	Epidemiology and Laboratory Capacity
ESBL	Extended-spectrum beta-lactamase
FDA-CVM	Food and Drug Administration-Center for Veterinary Medicine
FoodNet	Foodborne Diseases Active Surveillance Network
MIC	Minimum inhibitory concentration
NARMS	National Antimicrobial Resistance Monitoring System for Enteric Bacteria
OR	Odds ratio
PHLIS	Public Health Laboratory Information System
USDA	United States Department of Agriculture
WHO	World Health Organization

NARMS Working Group

Centers for Disease Control and Prevention

Enteric Diseases Epidemiology Branch

Enteric Diseases Laboratory Branch

Division of Foodborne, Waterborne and Environmental Diseases

National Center for Emerging and Zoonotic Infectious Diseases

Jason Folster
Peter Gerner-Smidt
Julian Grass
Audrey Green
Patricia Griffin
Robert Michael Hoekstra
Rebecca Howie
Kevin Joyce
Maria Karlsson
Beth Karp
Amy Krueger
Andre McCullough
Felicita Medalla
Allison O'Donnell
Gary Pecic
Melissa Pitcher
Jared Reynolds
Regan Rickert
Robert Tauxe
Julia Taylor
Jean Whichard

U.S. Food and Drug Administration

Center for Veterinary Medicine

Heather Green
Claudine Kabera
Patrick McDermott
Emily Tong
Niketia Womack

Participating State and Local Health Departments

Alabama Department of Public Health

LaDonna Cranidiotis
Sherri Davidson
Sharon Massingale
Patricia Morrow
Joanna Roberson

Alaska Department of Health and Social Services

Shellie Smith
Catherine Xavier

Arizona Department of Health Services

Shoana Anderson
Aarikha D'Souza
Daniel Flood
Melissa Hoffman
Ken Komatsu
William Slanta
Victor Waddell

Arkansas Department of Health

Rossina Stefanova

California Department of Health Services

Wendy Cheung
Claudia Crandall
Samar Fontanoz
Paul Kimsey
Will Probert
Sam Shin
Duc Vugia

Colorado Department of Public Health and Environment

Alicia Cronquist
Laura Gillim-Ross
Joyce Knutsen
Hugh Maguire

Connecticut Department of Public Health

Diane Barden
Sharon Hurd
Aristea Kinney
Mona Mandour

Delaware Health and Social Services

Gaile McLaughlin
Bela Patel
Debra Rutledge

Florida Department of Health

Ronald Baker
Maria Calcaterra
Sonia Etheridge
Dian Sharma

Georgia Division of Public Health

Jim Benson
Elizabeth Franko
Tameka Hayes
Mary Hodel
Susan Lance
Bob Manning
Mahin Park
Lynett Poventud
Suzanne Segler
Stepy Thomas
Melissa Tobin-D'Angelo

Hawaii Department of Health

Rebecca Kanenaka
Norman O'Connor

Houston Health and Human Services Department

Raouf Arafat
Adebowale Awosika-Olumo
Gregory Dufour
Vern Juchau
Sudha Pottumarthy
Joan Rogers

Idaho Department of Health and Welfare

Colleen Greenwalt
Vivian Lockary
Raemi Nolevanko

Illinois Department of Public Health

Nancy Barstead
Bob Cox
Mark Dworkin
Juan Garcia
Rebecca Hambelton
Stephen Hendren
Steve Hopkins
Patrick Miller
Mohammad Nasir
Kiran Patel
Tricia Patterson
Guinevere Reserva
Bindu Shah
Andrea Stadsholt

Indiana State Department of Health

Brent Barrett
Amie May
John Radosevic

Iowa Department of Public Health, University Hygienic Laboratory

Mary DeMartino
Randy Groepper

Kansas Department of Health and Environment

Cheryl Banez-Ocfemia
Robert Flahart
Gail Hansen
Carissa Pursell
June Sexton
Kathleen Waters

Kentucky Department of Public Health

Robin Cotton
Karim George
William Grooms
Darrin Sevier
Jack Wiedo

Los Angeles County Department of Health Services

Michael Stephens
Sheena Chu
Sue Sabet
Laurene Mascola
Roshan Reporter
Joan Sturgeon

Louisiana Department of Health and Hospitals

Gary Balsamo
Erin Delaune
Wayne Dupree
Catrin Jones-Nazar
Lori Kravet
Steven Martin
Raoult Ratard
Theresa Sokol
Susanne Straif-Bourgeois

Maine Department of Human Services

Geoff Beckett
Kathleen Gensheimer
Jeff Randolph
Vicki Rea
Lori Webber
Donna Wrigley
Anthony Yartel

Maryland Department of Health and Mental Hygiene

David Blythe
Kirsten Larson
Celere Leonard
Amanda Palmer
Jafar Razeq
Pat Ryan

Massachusetts Department of Public Health

Catherine Brown
Alfred DeMaria
Robert Goldbaum
Emily Harvey
Patricia Kludt
Joseph Peppe
Tracy Stiles

Michigan Department of Community Health

Carrie Anglewicz
Frances Downes
Teri Lee Dyke
James Rudrik
William Schneider
Patricia Somsel

Minnesota Department of Health

John Besser
Billie Juni
Fe Leano
Stephanie Meyer
Kirk Smith
Charlotte Taylor
Theresa Weber

Mississippi Department of Health

Jannifer Anderson
Jane Campbell
Gloria Kendrick
Sheryl Hand
Cathie Hoover
Daphne Ware

Missouri Department of Health

David Byrd
Steve Gladbach
Jason Herstein
Harvey Marx
JoAnn Rudroff

Montana Department of Public Health and Human Services

Bonnie Barnard
Anne Weber
Susanne Zanto

Nebraska Health and Human Services and the Nebraska Public Health Laboratory

Amy Armbrust
Jude Dean
Paul Fey
Peter Iwen
Tom Safranek

Nevada Department of Health and Human Services

Vince Abitria
Patricia Armour
Stephanie Ernaga
Jaime Frank
Paul Hug
Bradford Lee
Susanne Quianzon
Lisa Southern
Stephanie Van Hooser

New Hampshire Department of Health and Human Services

Christine Adamski
Christine Bean
Elizabeth Daly
Wendy Lamothe
Nancy Taylor
Daniel Tullo

New Jersey Department of Health

Ruth Besco
Michelle Malavet
Sylvia Matiuck
Paul Seitz

New Mexico Department of Health

Lisa Butler
Cynthia Nicholson
Lisa Onischuk
Erica Pierce
Paul Torres

New York City Department of Health

Sharon Balter
Ludwin Chicaiza
Heather Hanson
Lillian Lee
Jennifer Rakeman
Vasudha Reddy

New York State Department of Health

Leanna Armstrong
Nellie Dumas
Tammy Quinlan
Dale Morse
Tim Root
Shelley Zansky

North Carolina Department of Health and Human Services

Denise Griffin
Debra Springer

North Dakota Department of Health

Lisa Elijah
Julie Wagendorf
Eric Hieb
Nicole Meier
Tracy Miller
Lisa Well

Ohio Department of Health

Rick Bokanyi
Tammy Bannerman
Jane Carmean
Larry King
Mary Kay Parrish
Susan Luning
Ellen Salehi

Oklahoma State Department of Health

Rebekah Berry
Mike Lytle
Mike McDermot

Oregon Department of Human Service

Debbie Berquist
Cathy Ciaffoni
Paul Cieslak
Dawn Daly
Emilio Debess
Julie Hatch
Beletsachew Shiferaw
Larry Stauffer
Janie Tierheimer
Robert Vega
Veronica Williams

Pennsylvania Department of Human Service

Wayne Chmielecki
Lisa Dettinger
Nkuchia Mikanatha
Stanley Reynolds
Carol Sandt
James Tait

Rhode Island Department of Health

Tara Cooper
Deanna Simmons
Cindy Vanner

South Carolina Department of Health and Environmental Control

Dana Giurgiutiu
Mamie Turner
Jennifer Meredith
Arthur Wozniak

South Dakota Department of Health

Christopher Carlson
Lon Kightlinger
Mike Smith
Yvette Thomas

Tennessee Department of Health

Parvin Arjmandi
Paula Bailey
John Dunn
Samir Hanna
Henrietta Hardin

Texas Department of State Health Services

Tamara Baldwin
Leslie Bullion
Elizabeth Delamater
Linda Gaul
Eldridge Hutcheson
Miriam Johnson
Susan Neil
Pushker Raj
Ana Valle

Utah Department of Health

Dan Andrews
Kim Christensen
Jana Coombs
Cindy Fisher
David Jackson
Barbara Jepson
Susan Mottice

Vermont Department of Health

Erica Berl
Valerie Cook
Eunice H. Froeliger
Christine LaBarre

Virginia Division of Consolidated Laboratory Services and Virginia Department of Health

Ellen Bassinger
Sherry Giese
Jody Lowman
Mary Mismas
Denise Toney

Washington Department of Health

Jennifer Breezee
Romesh Gautom
Donna Green
Brian Hiatt
Yolanda Houze
Kathryn MacDonald

West Virginia Department of Health and Human Resources

Danae Bixler
Christi Clark
Maria del Rosario
Loretta Haddy
Andrea Labik
Megan Young

Wisconsin Department of Health and Family Services

John Archer
Susann Ahrabi-Fard
Charles Brokopp
Jeffrey Davis
Rick Hefferman
Rachel Klos
Tim Monson
Dave Warshauer

Wyoming Department of Health

Richard Harris
John Harrison
Clay Van Houten
Tracy Murphy
Jim Walford

Introduction

The primary purpose of the National Antimicrobial Resistance Monitoring System (NARMS) at CDC is to monitor antimicrobial resistance among enteric bacteria isolated from humans. Other components of the interagency NARMS program include surveillance for resistance in enteric bacteria isolated from foods, conducted by the FDA-CVM

(<http://www.fda.gov/AnimalVeterinary/SafetyHealth/AntimicrobialResistance/NationalAntimicrobialResistanceMonitoringSystem/default.htm>), and resistance in enteric bacteria isolated from animals, conducted by the USDA Agricultural Research Service (http://www.ars.usda.gov/main/site_main.htm?modecode=66-12-05-08).

Many NARMS activities are conducted within the framework of CDC's Emerging Infections Program (EIP), Epidemiology and Laboratory Capacity (ELC) Program, and the Foodborne Diseases Active Surveillance Network (FoodNet). In addition to surveillance of resistance in enteric pathogens, the NARMS program at CDC also includes research into the mechanisms and public health impact of resistance, education efforts to promote prudent use of antimicrobial agents, and antimicrobial susceptibility testing of isolates that caused outbreaks.

Before NARMS was established, CDC monitored antimicrobial resistance in *Salmonella*, *Shigella*, and *Campylobacter* through periodic surveys of isolates from a panel of sentinel counties. NARMS at CDC began in 1996 with prospective monitoring of antimicrobial resistance among clinical non-Typhi *Salmonella* and *Escherichia coli* O157 isolates in 14 sites. In 1997, testing of clinical *Campylobacter* isolates was initiated in the five sites participating in FoodNet. Testing of clinical *Salmonella enterica* serotype Typhi and *Shigella* isolates was added in 1999. Since 2003, all 50 states have been forwarding a representative sample of non-Typhi *Salmonella*, *Salmonella* ser. Typhi, *Shigella*, and *E. coli* O157 isolates to NARMS for antimicrobial susceptibility testing, and 10 FoodNet states have been participating in *Campylobacter* surveillance. Since 2008, all 50 states have been forwarding every *Salmonella* Paratyphi A and C to NARMS for antimicrobial susceptibility testing.

This annual report includes CDC's surveillance data for 2010 for non-typhoidal *Salmonella*, typhoidal *Salmonella*, *Shigella*, *Campylobacter* and *E. coli* O157 isolates in addition to surveillance data for 2009 *Vibrio* species other than *V. cholerae*. Data for earlier years are presented in tables and graphs when appropriate. Antimicrobial classes defined by Clinical and Laboratory Standards Institute (CLSI) are used in data presentation and analysis. CLSI classes constitute major classifications of antimicrobial agents, e.g., aminoglycosides and cepheems.

This report also includes the World Health Organization's categorization of antimicrobials of critical importance to human medicine ([Table 1](#)). The table includes only antimicrobials that are tested in NARMS.

Additional NARMS data and more information about NARMS activities are available at <http://www.cdc.gov/narms>

Vibrio* Species other than *V. cholerae

For the first time, in this NARMS report we present antimicrobial susceptibility data for *Vibrio* species other than *V. cholerae* isolated from humans. We asked NARMS participating public health laboratories to submit all *Vibrio* except *V. cholerae* species for susceptibility testing at the NARMS laboratory at CDC. CDC determined MICs for 9 antimicrobial agents using Etest® (BioMérieux, Marcy L'Etoile, France). Here we present MIC distributions for isolates collected in 2009 and report resistance frequencies for agents that have CLSI-published interpretive criteria for *Vibrio* species other than *V. cholerae*.

Fluoroquinolone Breakpoint Changes for *Enterobacteriaceae*

CLSI is revising fluoroquinolone interpretive criteria for invasive *Salmonella* and other *Enterobacteriaceae*. Specifically, for invasive *Salmonella*, updated ciprofloxacin MIC ranges for susceptible (S), intermediate (I), and resistant (R) categories appeared in the January 2012 CLSI M100 supplement. In this report, we show S, I, and R frequencies for *Salmonella* (typhoidal and non-typhoidal) using both the outgoing and new breakpoints in [Box 2](#). The figures and tables in the results section are based on the pre-2012 breakpoints.

Susceptibility Data for Bacteria from Outbreaks

CDC has enhanced its approaches to attributing foodborne disease to specific foods and other sources of human infection. These changes include determining sources of antimicrobial-resistant infections. To support antimicrobial resistance attribution goals, CDC has requested that NARMS-participating state public health laboratories submit representative bacterial isolates from foodborne disease outbreaks for antimicrobial susceptibility testing. The scope and number of isolates requested over the years is described in the methods section of [Appendix A](#). For the first time, in this NARMS report we show antimicrobial susceptibility results for outbreaks of *Salmonella* infections for which a vehicle was implicated.

Population

In 2010, all 50 states participated in NARMS, representing the entire U.S. population of approximately 309 million persons ([Table 2](#)). Surveillance was conducted in all states for non-typhoidal *Salmonella*, typhoidal *Salmonella*, *Shigella*, *Escherichia coli* O157, and *Vibrio* species other than *V. cholerae*. For *Campylobacter*, surveillance was conducted in 10 states that comprise the Foodborne Diseases Active Surveillance Network (FoodNet), representing approximately 47 million persons (15.2% of the U.S. population).

Clinically Important Antimicrobial Resistance Patterns

In the United States, fluoroquinolones (e.g., ciprofloxacin) and third-generation cephalosporins (e.g., ceftriaxone) are commonly used to treat severe *Salmonella* infections, including *Salmonella* ser. Typhi, the organism that causes typhoid fever. In *Enterobacteriaceae*, resistance to nalidixic acid, an elementary quinolone, correlates with decreased susceptibility to ciprofloxacin (MIC ≥ 0.12 $\mu\text{g/mL}$) and possible fluoroquinolone treatment failure. A substantial proportion of *Enterobacteriaceae* isolates tested in 2010 demonstrated resistance to clinically important antimicrobial agents.

- 2.0% (49/2474) of non-typhoidal *Salmonella* isolates were resistant to nalidixic acid, including
 - 5.2% (27/522) of *Salmonella* ser. Enteritidis isolates
 - Enteritidis was the most common serotype among nalidixic acid-resistant non-typhoidal *Salmonella* isolates: 55.1% (27/49) of nalidixic acid-resistant isolates were serotype Enteritidis.
- 2.8% (70/2474) of non-typhoidal *Salmonella* isolates were resistant to ceftriaxone, including
 - 24.2% (15/62) of *Salmonella* ser. Heidelberg isolates
 - 7.2% (22/305) of *Salmonella* ser. Newport isolates
 - 4.9% (18/366) of *Salmonella* ser. Typhimurium isolates
 - Newport was the most common serotype among ceftriaxone-resistant non-typhoidal *Salmonella* isolates: 31.4% (22/70) of ceftriaxone-resistant isolates were serotype Newport.
- 69.1% (307/444) of *Salmonella* ser. Typhi isolates were resistant to nalidixic acid and 2.7% (12/444) were resistant to ciprofloxacin.
- 90.4% (132/146) of *Salmonella* ser. Paratyphi A, Paratyphi B, and Paratyphi C were resistant to nalidixic acid
- 4.4% (18/407) of *Shigella* isolates were resistant to nalidixic acid and 1.7% (7/407) were resistant to ciprofloxacin.

In *Campylobacter*, fluoroquinolones and macrolides (e.g., erythromycin) are important agents in the treatment of severe infections.

- 22.4% (294/1310) of *Campylobacter* isolates were resistant to ciprofloxacin, including
 - 21.8% (253/1158) of *Campylobacter jejuni* isolates
 - 31.3% (36/115) of *Campylobacter coli* isolates
- 1.5% (19/1310) of *Campylobacter* isolates were resistant to erythromycin, including
 - 1.2% (14/1158) *Campylobacter jejuni* isolates
 - 4.3% (5/115) of *Campylobacter coli* isolates

Multidrug Resistance

Multidrug resistance is described in NARMS as resistance to three or more antimicrobial classes and also by specific coresistant phenotypes. Antimicrobial classes of agents defined by the Clinical and Laboratory Standards Institute (CLSI) are used in this report ([Table 3](#), [Table 4](#), [Table 5](#)). For non-typhoidal *Salmonella*, an important multidrug-resistant phenotype includes resistance to at least ampicillin, chloramphenicol, streptomycin, sulfonamide (sulfamethoxazole/sulfisoxazole), and tetracycline (ACSSuT). The ACSSuT phenotype includes resistance to at least five CLSI classes. Another important phenotype includes resistance to at least ampicillin, chloramphenicol, streptomycin, sulfonamide, tetracycline, amoxicillin-clavulanic acid, and ceftriaxone (ACSSuTAuCx). The ACSSuTAuCx phenotype includes resistance to at least seven CLSI classes.

- 11.3% (279/2474) of non-typhoidal *Salmonella* isolates were resistant to two or more CLSI classes of antimicrobial agents, and 9.1% (225/2474) were resistant to three or more CLSI classes.
 - 33.9% (21/62) of *Salmonella* ser. Heidelberg isolates were resistant to three or more CLSI classes.

- 27.3% (100/366) of *Salmonella* ser. Typhimurium isolates were resistant to three or more CLSI classes.
- 22.1% (17/77) of *Salmonella* ser. I,4,[5],12:i:- isolates were resistant to three or more CLSI classes.
- 7.5% (23/305) of *Salmonella* ser. Newport isolates were resistant to three or more CLSI classes.
- 2.1% (11/522) of *Salmonella* ser. Enteritidis isolates were resistant to three or more CLSI classes.
- Of 225 non-typhoidal *Salmonella* resistant to three or more CLSI classes, 44.4% were *Salmonella* ser. Typhimurium.
- 4.3% (107/2474) of non-typhoidal *Salmonella* isolates were at least ACSSuT-resistant, including
 - 18.6% (68/366) of *Salmonella* ser. Typhimurium isolates, and
 - 7.2% (22/305) of *Salmonella* ser. Newport isolates.
- 1.3% (33/2474) of non-typhoidal *Salmonella* isolates were at least ACSSuTAuCx-resistant, including
 - 7.2% (22/305) of *Salmonella* ser. Newport isolates, and
 - 1.9% (7/366) of *Salmonella* ser. Typhimurium isolates.
- 13.7% (61/444) of *Salmonella* ser. Typhi isolates were resistant to three or more classes.
- 40.0% (163/407) of *Shigella* isolates were resistant to three or more classes.
- 3.6% (6/167) of *E. coli* O157 isolates were resistant to three or more classes.

Antimicrobial Resistance: 1996–2010

The following figures display resistance from 1996–2010 for non-typhoidal *Salmonella*, 1999–2010 for *Salmonella* ser. Typhi, and 1997–2010 for *Campylobacter*.

Figure 1. Percentage of non-typhoidal *Salmonella* isolates resistant to nalidixic acid, by year, 1996–2010

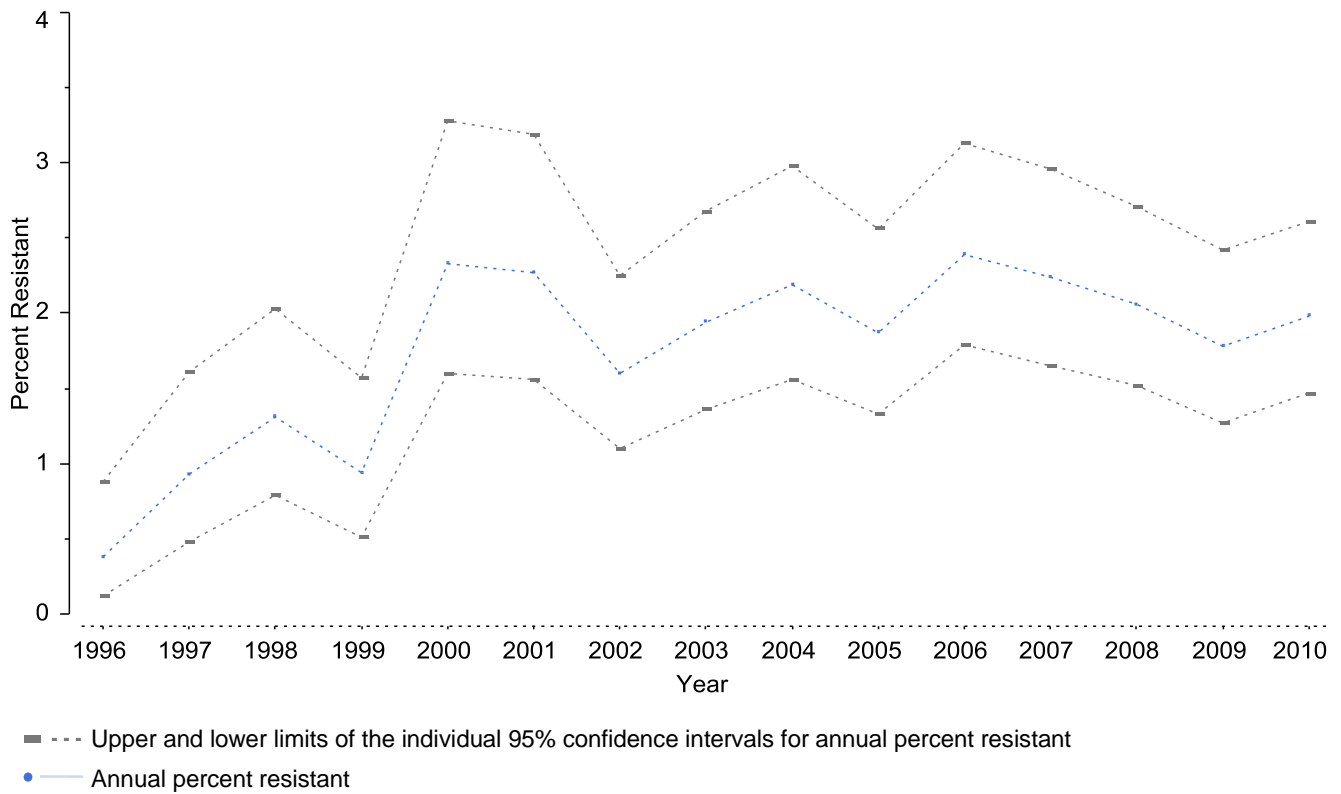


Figure 2. Percentage of *non-typhoidal Salmonella* isolates resistant to ceftriaxone, by year, 1996–2010

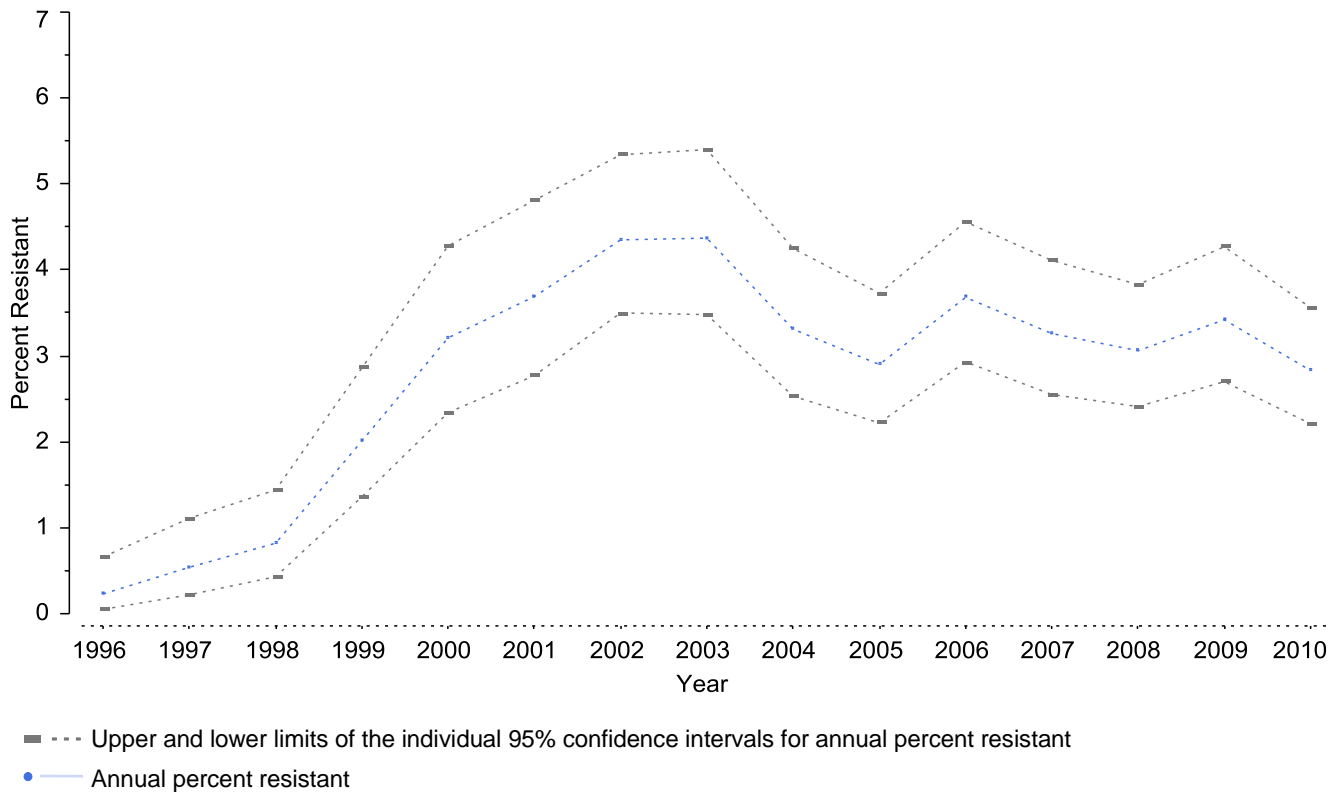


Figure 3. Percentage of *Salmonella ser. Enteritidis* isolates resistant to nalidixic acid, by year, 1996–2010

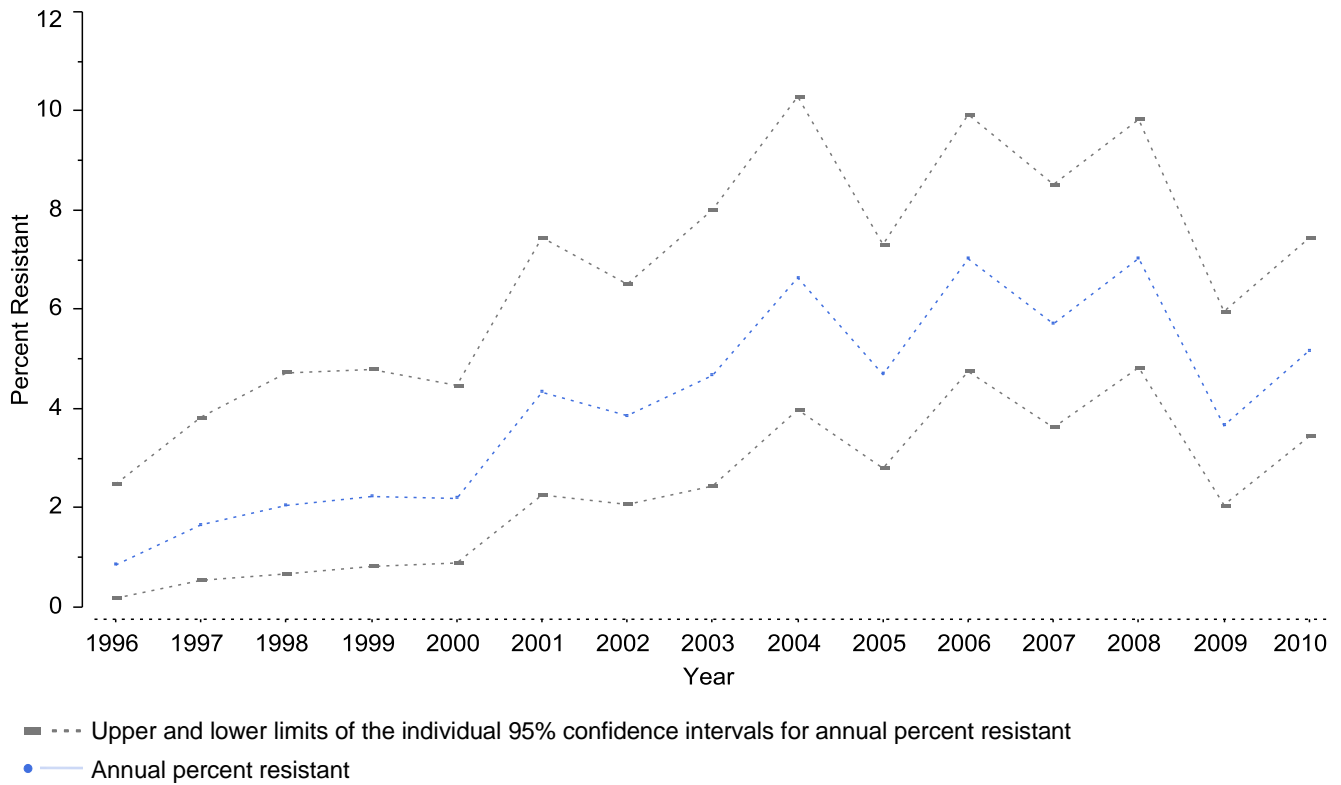


Figure 4. Percentage of *Salmonella ser. Heidelberg* isolates resistant to ceftriaxone, by year, 1996–2010

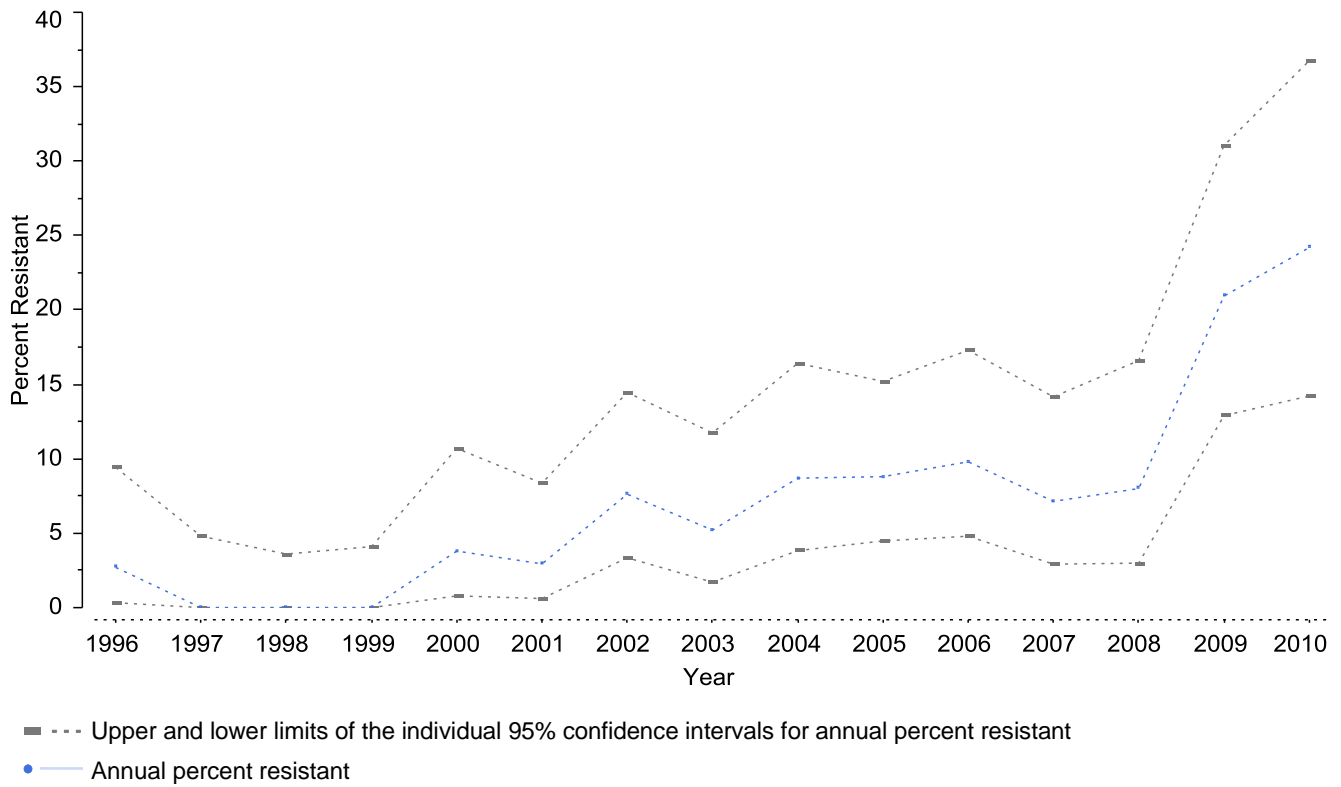


Figure 5. Percentage of *Salmonella ser. Typhimurium* isolates resistant to at least ampicillin, chloramphenicol, streptomycin, sulfonamide, and tetracycline (ACSSuT), by year, 1996–2010

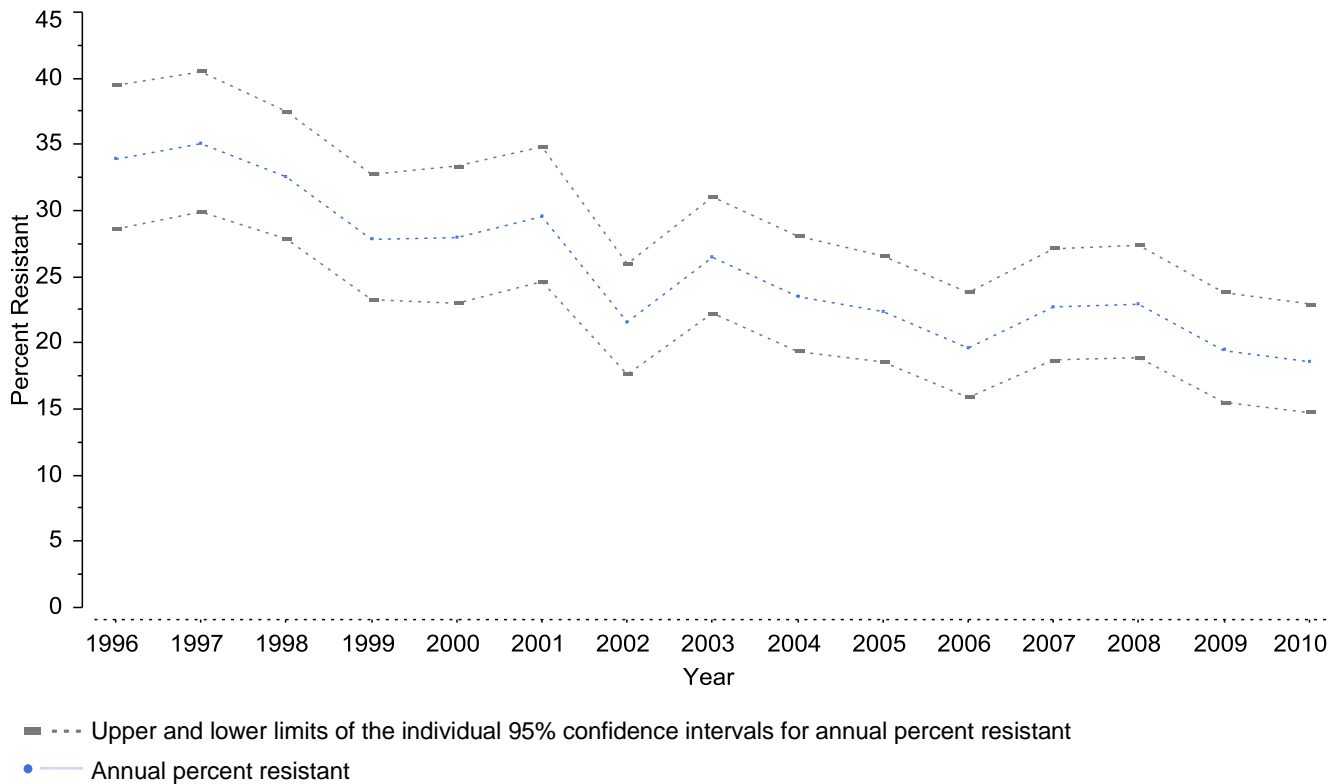


Figure 6. Percentage of *Salmonella ser. Newport* isolates resistant to at least ampicillin, chloramphenicol, streptomycin, sulfonamide, tetracycline, amoxicillin-clavulanic acid, and ceftriaxone (ACSSuTAuCx), by year, 1996–2010

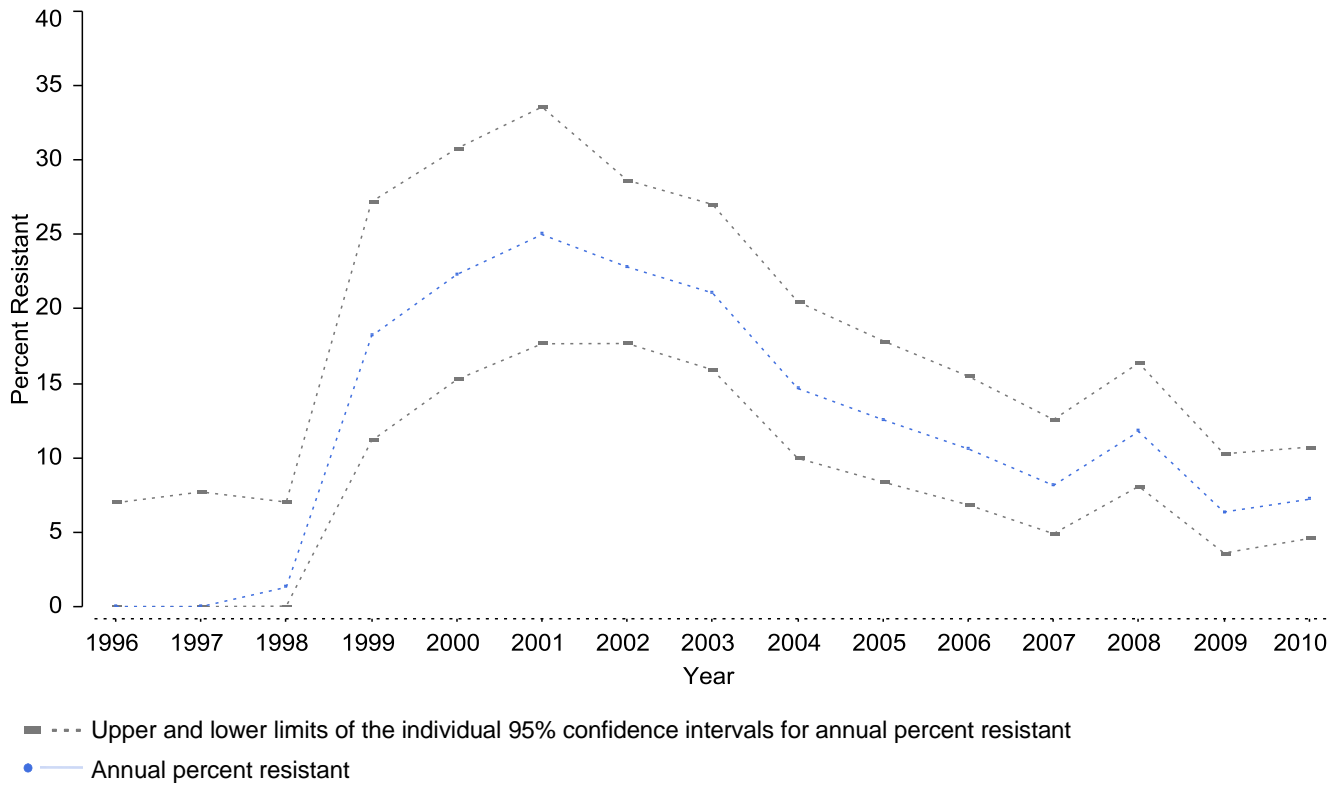


Figure 7. Percentage of non-typhoidal *Salmonella* isolates resistant to 1 or more antimicrobial classes, by year, 1996–2010

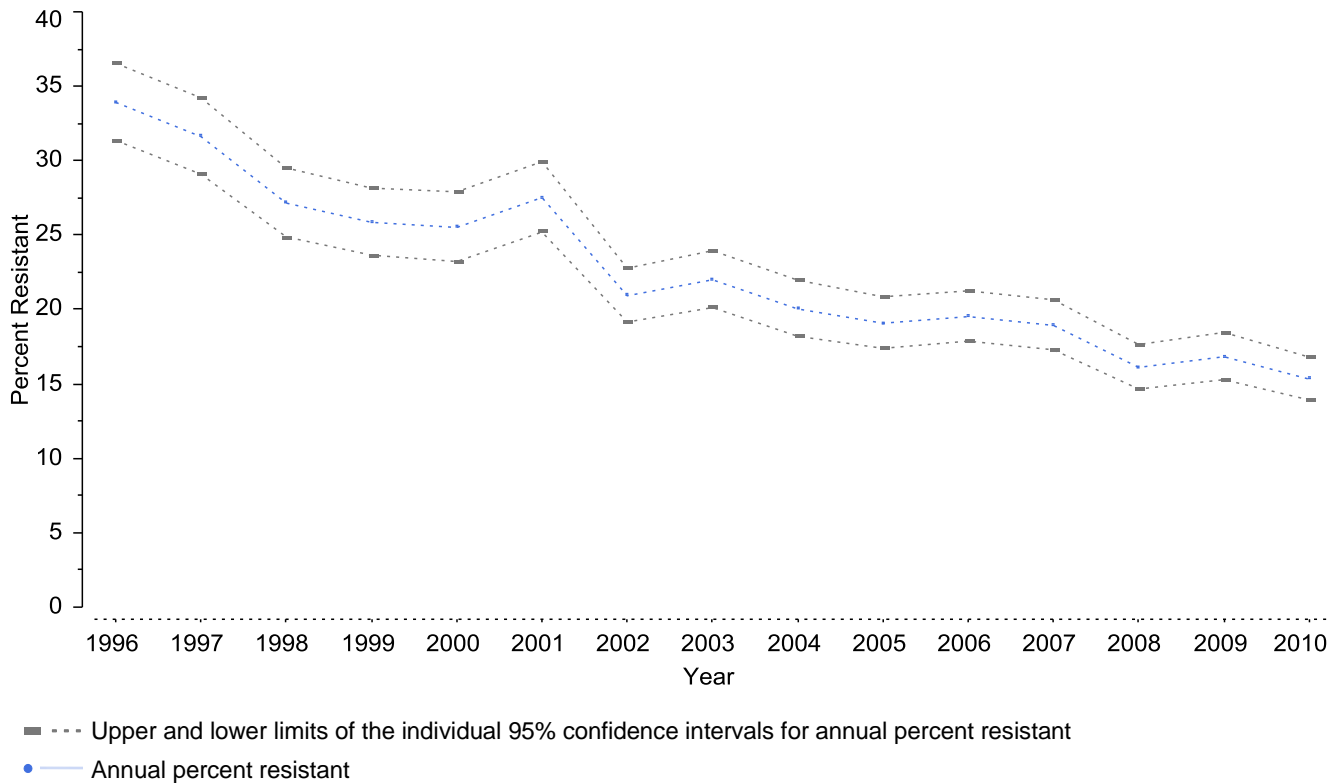


Figure 8. Percentage of non-typhoidal *Salmonella* isolates resistant to 3 or more antimicrobial classes, by year, 1996–2010

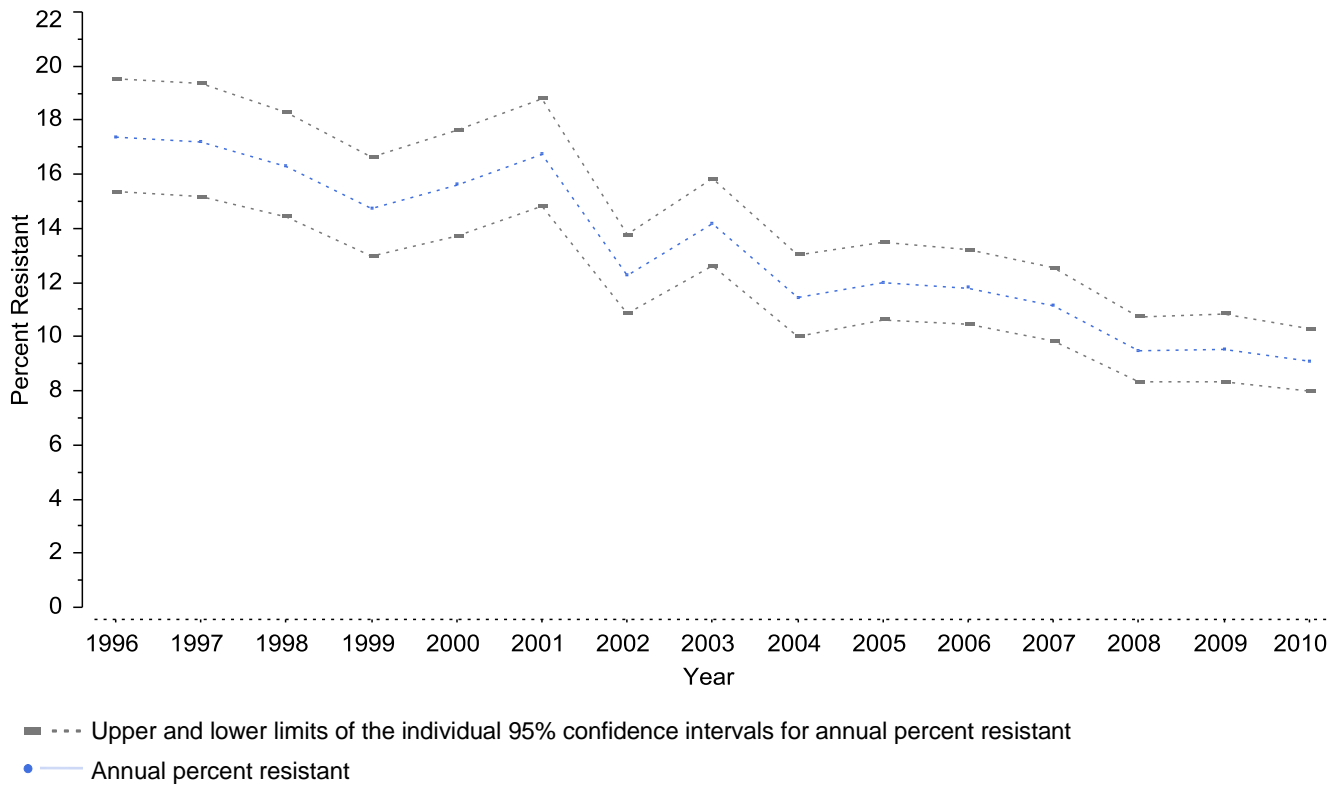


Figure 9. Percentage of *Salmonella ser. Typhi* isolates resistant to nalidixic acid, by year, 1999–2010

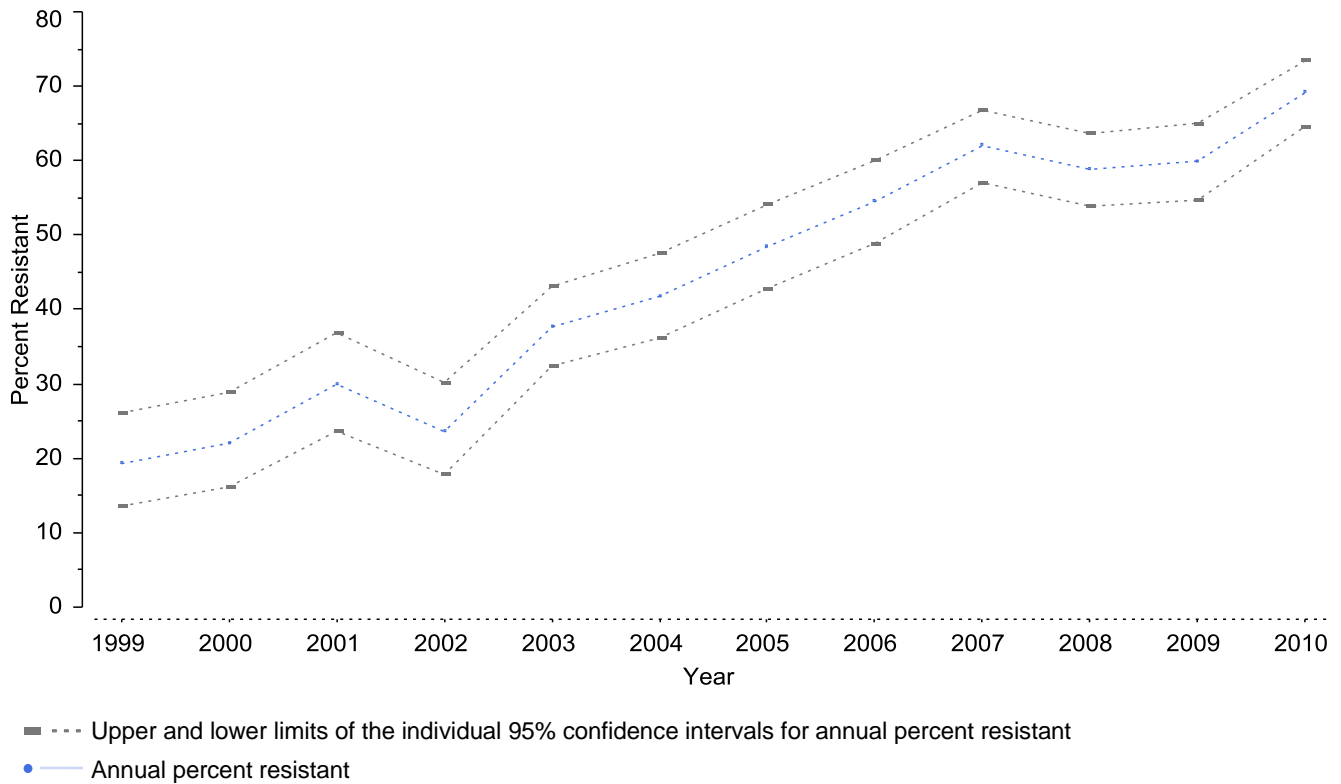
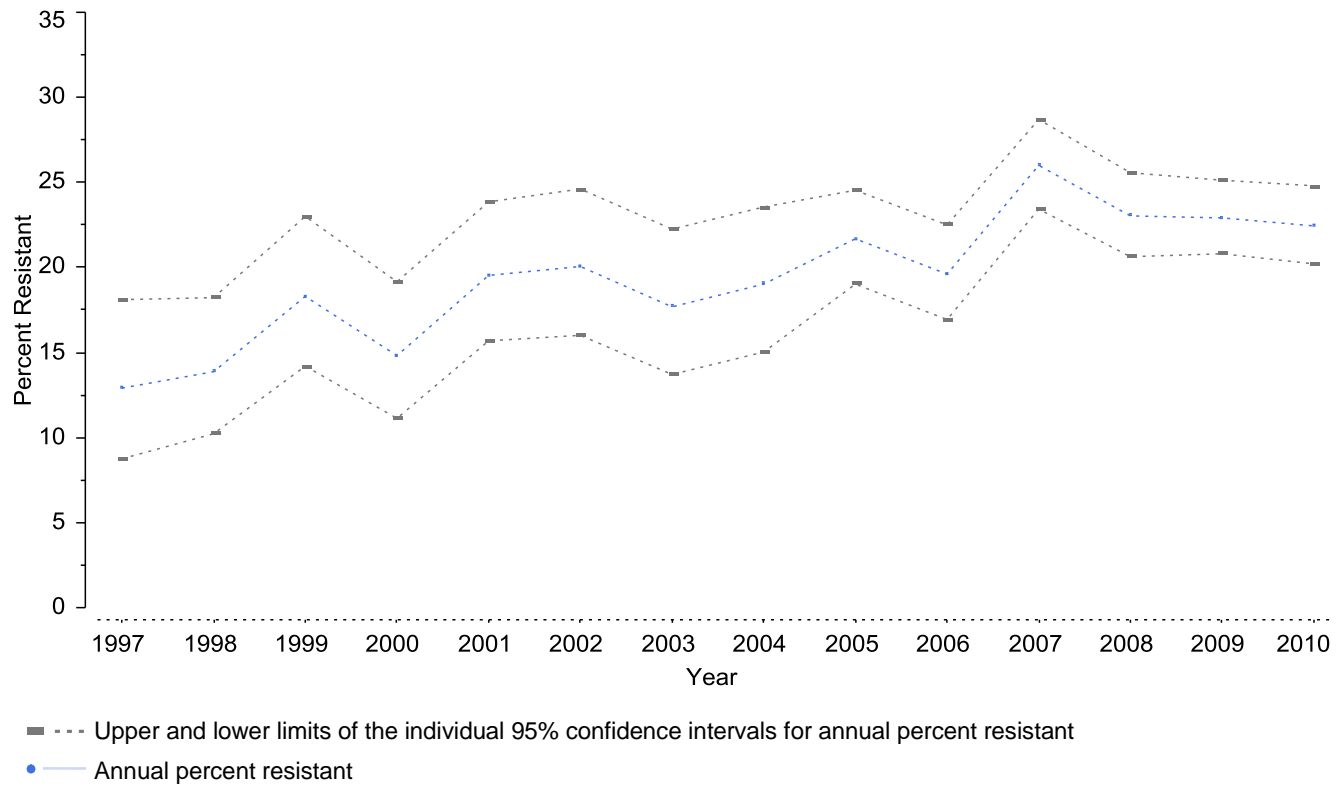


Figure 10. Percentage of *Campylobacter* isolates resistant to ciprofloxacin, by year, 1997–2010



WHO Categorization of Antimicrobial Agents

In 2009, the World Health Organization (WHO) convened a panel of experts to update a list of antimicrobial agents ranked according to their relative importance to human medicine ([WHO, 2009](#)). The participants categorized antimicrobial agents as either Critically Important, Highly Important, or Important based upon two criteria: (1) used as sole therapy or one of the few alternatives to treat serious human disease and (2) used to treat disease caused by either organisms that may be transmitted via non-human sources or diseases caused by organisms that may acquire resistance genes from non-human sources. In 2009, WHO recategorized tetracycline from highly important to critically important. Antimicrobial agents tested in NARMS have been included in the WHO categorization table.

- Antimicrobial agents are critically important if both criteria (1) and (2) are true.
- Antimicrobial agents are highly important if either criterion (1) or (2) is true.
- Antimicrobial agents are important if neither criterion is true.

Table 1. WHO categorization of antimicrobials of critical importance to human medicine

WHO Category Level	Importance	CLSI Class	Antimicrobial Agent tested in NARMS
I	Critically important	Aminoglycosides	Amikacin
			Gentamicin
			Streptomycin
		β-lactam / β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid
		Cephems	Ceftriaxone
		Ketolides	Telithromycin
		Macrolides	Azithromycin
		Penicillins	Ampicillin
		Quinolones	Ciprofloxacin
		Tetracyclines	Nalidixic acid
II	Highly important	Aminoglycosides	Kanamycin
		Cephems	Cefoxitin
		Folate pathway inhibitors	Cephalothin
		Phenicols	Sulfamethoxazole / Sulfisoxazole
		Tetracyclines	Trimethoprim-sulfamethoxazole
1. III	Important	Lincosamides	Chloramphenicol
1. III	Important	Lincosamides	Clindamycin

Surveillance Sites and Isolate Submissions

In 2010 NARMS conducted nationwide surveillance among approximately 309 million persons ([2010 U.S. Census Bureau estimates](#)). Public health laboratories systematically selected every 20th non-typhoidal *Salmonella*, *Shigella*, and *Escherichia coli* O157 isolate as well as every *Salmonella* ser. Typhi, *Salmonella* ser. Paratyphi A and *Salmonella* ser. Paratyphi C isolate received at their laboratories and forwarded these isolates to CDC for antimicrobial susceptibility testing. *Salmonella* ser. Paratyphi B was included in the every 20th sampling for non-typhoidal *Salmonella* because available laboratory methods do not always allow for consistent distinction between serotype Paratyphi B (which typically causes typhoidal illness) and serotype Paratyphi B var. L(+) tartrate+ (which does not typically cause typhoidal illness). Beginning in 2009, NARMS also performed susceptibility testing on isolates of *Vibrio* species other than *V. cholerae* submitted by the NARMS participating public health laboratories. Participants were asked to forward every isolate of *Vibrio* species other than *V. cholerae* that they received to CDC for antimicrobial susceptibility testing and confirmation by CDC's National Enteric Reference Laboratory.

Since 2005, public health laboratories of the 10 state health departments that participated in CDC's Foodborne Diseases Active Surveillance Network (FoodNet) have forwarded a representative sample of *Campylobacter* isolates to CDC for susceptibility testing. The FoodNet sites, representing approximately 47 million persons (2010 U.S. Census Bureau estimates), include California, Colorado, Connecticut, Georgia, Maryland, Minnesota, New Mexico, New York, Oregon, and Tennessee. Depending on the burden of *Campylobacter* in each FoodNet site, one of the following four methods was used to obtain and test a representative sample of *Campylobacter* isolates in 2010: all isolates received by Oregon and Tennessee; every other isolate from California, Colorado, Connecticut, Georgia, Maryland, and New York; every third isolate from New Mexico; and every fifth isolate from Minnesota. Isolates received from 2005 to 2009 had the same methods except all isolates were sent from Georgia, Maryland, and New Mexico. From 1997 to 2004, one *Campylobacter* isolate was submitted each week from participating FoodNet sites.

Table 2. Population size and number of isolates received and tested, NARMS, 2010

State/Site	Population Size	Non-typhoidal <i>Salmonella</i>		Typhoidal [†] <i>Salmonella</i>		<i>Shigella</i>		<i>E. coli</i> O157		<i>Campylobacter</i> [‡]	
		n	(%)	n	(%)	n	(%)	n	(%)	n	(%)
Alabama	4,779,736	67	(2.7)	5	(0.8)	12	(2.9)	2	(1.2)		
Alaska	710,231	3	(0.1)	0	(0)	1	(0.2)	1	(0.6)		
Arizona	6,392,017	54	(2.2)	11	(1.9)	20	(4.9)	0	(0)		
Arkansas	2,915,918	30	(1.2)	0	(0)	3	(0.7)	1	(0.6)		
California [§]	27,435,351	236	(9.5)	100	(16.9)	4	(1.0)	9	(5.4)	151	(11.5)
Colorado	5,029,196	32	(1.3)	3	(0.5)	7	(1.7)	4	(2.4)	52	(4.0)
Connecticut	3,574,097	28	(1.1)	9	(1.5)	1	(0.2)	2	(1.2)	124	(9.5)
Delaware	897,934	10	(0.4)	2	(0.3)	2	(0.5)	0	(0)		
District of Columbia	601,723	10	(0.4)	2	(0.3)	0	(0)	0	(0)		
Florida	18,801,310	29	(1.2)	22	(3.7)	0	(0)	0	(0)		
Georgia	9,687,653	155	(6.3)	15	(2.5)	37	(9.1)	25	(15.0)	218	(16.6)
Hawaii	1,360,301	17	(0.7)	2	(0.3)	4	(1.0)	1	(0.6)		
Houston, Texas [¶]	2,099,451	41	(1.7)	6	(1.0)	10	(2.5)	1	(0.6)		
Idaho	1,567,582	9	(0.4)	1	(0.2)	0	(0)	1	(0.6)		
Illinois	12,830,632	96	(3.9)	30	(5.1)	30	(7.4)	10	(6.0)		
Indiana	6,483,802	40	(1.6)	3	(0.5)	1	(0.2)	4	(2.4)		
Iowa	3,046,355	23	(0.9)	7	(1.2)	4	(1.0)	4	(2.4)		
Kansas	2,853,118	16	(0.6)	1	(0.2)	6	(1.5)	1	(0.6)		
Kentucky	4,339,367	24	(1.0)	0	(0)	1	(0.2)	1	(0.6)		
Los Angeles ^{**}	9,818,605	60	(2.4)	24	(4.1)	4	(1.0)	0	(0)		
Louisiana	4,533,372	24	(1.0)	1	(0.2)	2	(0.5)	0	(0)		
Maine	1,328,361	4	(0.2)	3	(0.5)	3	(0.7)	2	(1.2)		
Maryland	5,773,552	55	(2.2)	13	(2.2)	5	(1.2)	5	(3.0)	111	(8.5)
Massachusetts	6,547,629	37	(1.5)	10	(1.7)	5	(1.2)	1	(0.6)		
Michigan	9,883,640	42	(1.7)	10	(1.7)	10	(2.5)	1	(0.6)		
Minnesota	5,303,925	35	(1.4)	8	(1.4)	4	(1.0)	8	(4.8)	183	(14.0)
Mississippi	2,967,297	55	(2.2)	2	(0.3)	2	(0.5)	2	(1.2)		
Missouri	5,988,927	53	(2.1)	2	(0.3)	67	(16.5)	8	(4.8)		
Montana	989,415	7	(0.3)	0	(0)	1	(0.2)	2	(1.2)		
Nebraska	1,826,341	12	(0.5)	2	(0.3)	6	(1.5)	4	(2.4)		
Nevada	2,700,551	19	(0.8)	5	(0.8)	2	(0.5)	1	(0.6)		
New Hampshire	1,316,470	9	(0.4)	5	(0.8)	1	(0.2)	1	(0.6)		
New Jersey	8,791,894	60	(2.4)	46	(7.8)	13	(3.2)	8	(4.8)		
New Mexico	2,059,179	18	(0.7)	0	(0)	7	(1.7)	0	(0)	97	(7.4)
New York ^{††}	11,202,969	80	(3.2)	28	(4.7)	7	(1.7)	3	(1.8)	196	(15.0)
New York City ^{‡‡}	8,175,133	76	(3.1)	58	(9.8)	14	(3.4)	4	(2.4)		
North Carolina	9,535,483	133	(5.4)	11	(1.9)	4	(1.0)	0	(0)		
North Dakota	672,591	4	(0.2)	2	(0.3)	0	(0)	1	(0.6)		
Ohio	11,536,504	72	(2.9)	14	(2.4)	9	(2.2)	6	(3.6)		
Oklahoma	3,751,351	3	(0.1)	0	(0)	1	(0.2)	0	(0)		
Oregon	3,831,074	26	(1.1)	5	(0.8)	3	(0.7)	6	(3.6)	138	(10.5)
Pennsylvania	12,702,379	85	(3.4)	21	(3.6)	30	(7.4)	3	(1.8)		
Rhode Island	1,052,567	9	(0.4)	6	(1.0)	1	(0.2)	1	(0.6)		
South Carolina	4,625,364	82	(3.3)	2	(0.3)	4	(1.0)	1	(0.6)		
South Dakota	814,180	9	(0.4)	1	(0.2)	1	(0.2)	1	(0.6)		
Tennessee	6,346,105	54	(2.2)	6	(1.0)	12	(2.9)	3	(1.8)	40	(3.1)
Texas ^{§§}	23,046,110	207	(8.4)	36	(6.1)	13	(3.2)	2	(1.2)		
Utah	2,763,885	19	(0.8)	1	(0.2)	4	(1.0)	3	(1.8)		
Vermont	625,741	5	(0.2)	0	(0)	1	(0.2)	1	(0.6)		
Virginia	8,001,024	69	(2.8)	21	(3.6)	5	(1.2)	3	(1.8)		
Washington	6,724,540	44	(1.8)	22	(3.7)	6	(1.5)	5	(3.0)		
West Virginia	1,852,994	35	(1.4)	0	(0)	15	(3.7)	8	(4.8)		
Wisconsin	5,686,986	45	(1.8)	6	(1.0)	2	(0.5)	5	(3.0)		
Wyoming	563,626	7	(0.3)	0	(0)	0	(0)	1	(0.6)		
Total	308,745,538	2474	(100)	590	(100)	407	(100)	167	(100)	1310	(100)

[†] US Census Bureau, 2010

[†] Typhoidal *Salmonella* includes Typhi, Paratyphi A, Paratyphi B, and Paratyphi C

[‡] *Campylobacter* isolates are submitted only from FoodNet sites representing a total population 47,053,218. All *Campylobacter* isolates are received from Georgia, Maryland, New Mexico, Oregon, and Tennessee and every other isolate from California, Colorado, Connecticut, and New York; and every fifth isolate from Minnesota.

[§] Excluding Los Angeles County

[¶] Houston City

^{**} Los Angeles County

^{††} Excluding New York City

^{‡‡} Five boroughs of New York City (Bronx, Brooklyn, Manhattan, Queens, Staten Island)

^{§§} Excluding Houston, Texas

Testing of *Salmonella*, *Shigella*, and *Escherichia coli* O157

Antimicrobial Susceptibility Testing

Salmonella, *Shigella*, and *E. coli* O157 isolates were tested using broth microdilution (Sensititre[®], Trek Diagnostics, Cleveland, OH) according to manufacturer's instructions to determine the minimum inhibitory concentration (MIC) for each of 15 antimicrobial agents: amikacin, ampicillin, amoxicillin-clavulanic acid, ceftiofur, ceftriaxone, chloramphenicol, ciprofloxacin, gentamicin, kanamycin, nalidixic acid, streptomycin, sulfisoxazole, tetracycline, and trimethoprim-sulfamethoxazole ([Table 3](#)). Before 2004, sulfamethoxazole was used instead of sulfisoxazole to represent the sulfonamides. Interpretive criteria defined by CLSI were used when available. The resistance breakpoint for amikacin, according to CLSI guidelines, is ≥ 64 $\mu\text{g}/\text{mL}$. In 2002 and 2003, a truncated broth microdilution series was used for amikacin testing (0.5-4 $\mu\text{g}/\text{mL}$). For isolates that grew in all amikacin dilutions on the Sensititre panel (MIC > 4 $\mu\text{g}/\text{mL}$), Etest[®] (AB BIODISK, Solna, Sweden) was performed to determine amikacin MIC. The amikacin Etest[®] strip range of dilutions was 0.016-256 $\mu\text{g}/\text{mL}$. Since 2004, amikacin had a full range of dilutions (0.5-64 $\mu\text{g}/\text{mL}$) on the Sensititre panel (CMV1AGNF). Repeat testing of isolates was done based on criteria in [Appendix B](#).

In January 2010, CLSI published revised interpretive criteria for ceftriaxone and *Enterobacteriaceae*; the revised resistance breakpoint for ceftriaxone is MIC ≥ 4 $\mu\text{g}/\text{mL}$. Since the 2009 report, NARMS has applied the revised CLSI breakpoint for ceftriaxone resistance to data from all years. In January 2012, CLSI published revised ciprofloxacin breakpoints for invasive *Salmonella* infections. For those infections, ciprofloxacin susceptibility is defined as ≤ 0.06 $\mu\text{g}/\text{mL}$; the intermediate category is defined as 0.12 to 0.5 $\mu\text{g}/\text{mL}$; and resistance is defined as ≥ 1 $\mu\text{g}/\text{mL}$. This year's report includes a comparison of the frequency of resistance based on the revised breakpoints with the frequency of resistance based on the previous breakpoints. Since all *Salmonella* serotypes have the potential to cause invasive infection, the revised breakpoints are applied to all *Salmonella* in this comparison shown in [Box 2](#).

Table 3. Antimicrobial agents used for susceptibility testing for *Salmonella*, *Shigella*, and *Escherichia coli* O157 isolates, NARMS, 2010

CLSI class	Antimicrobial Agent	Antimicrobial Agent Concentration Range (µg/mL)	MIC Interpretive Standard (µg/mL)		
			Susceptible	Intermediate*	Resistant
Aminoglycosides	Amikacin	0.5–64	≤16	32	≥64
	Gentamicin	0.25–16	≤4	8	≥16
	Kanamycin	8–64	≤16	32	≥64
	Streptomycin [†]	32–64	≤32	N/A	≥64
β-lactam / β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid	1/0.5–32/16	≤8/4	16/8	≥32/16
Cephems	Cefoxitin	0.5–32	≤8	16	≥32
	Ceftiofur	0.12–8	≤2	4	≥8
	Ceftriaxone [‡]	0.25–64	≤1	2	≥4
	Cephalothin [§]	2–32	≤8	16	≥32
Folate pathway inhibitors	Sulfamethoxazole [¶]	16–512	≤256	N/A	≥512
	Sulfisoxazole	16–256	≤256	N/A	≥512
	Trimethoprim-sulfamethoxazole	0.12/2.38–4/76	≤2/38	N/A	≥4/76
Penicillins	Ampicillin	1–32	≤8	16	≥32
Phenicols	Chloramphenicol	2–32	≤8	16	≥32
Quinolones	Ciprofloxacin ^{**}	0.015–4	≤1	2	≥4
	Nalidixic acid	0.5–32	≤16	N/A	≥32
Tetracyclines	Tetracycline	4–32	≤4	8	≥16

* N/A indicates that no MIC range of intermediate susceptibility exists

† No CLSI breakpoints; resistance breakpoint used in NARMS is ≥64 µg/mL.

‡ CLSI updated the ceftriaxone interpretive standards in January, 2010. Previous standards that were used for NARMS Human Isolate reports from 1996-2008 were susceptible ≤8 µg/mL, intermediate 16-32 µg/mL, and resistant ≥64 µg/mL.

§ Cephalothin was tested from 1996 to 2003 for *Salmonella*, *Shigella*, and *E. coli* O157.

¶ Sulfamethoxazole, which was tested during 1996–2003 to represent sulfonamides, was replaced by sulfisoxazole in 2004.

** CLSI breakpoints for invasive *Salmonella* infections were updated, effective January 2012. For those infections, ciprofloxacin susceptibility is defined as ≤0.06 µg/mL; the intermediate category is defined as 0.12 to 0.5 µg/mL; and resistance is defined as ≥1 µg/mL.

Additional Testing of *Salmonella* Strains

Cephalosporin Retesting of Isolates from 1996-1998

Review of *Salmonella* isolates tested in NARMS during 1996 to 1998 gave conflicting cephalosporin susceptibility results. In particular, some isolates previously reported in NARMS as ceftiofur-resistant exhibited a low ceftriaxone MIC and, in some cases, did not exhibit an elevated MIC to other β-lactams. Because these findings suggested that some previously reported results were inaccurate, we retested, using the 2003 NARMS Sensititre[®] plate, isolates of *Salmonella* tested in NARMS during 1996 to 1998 that exhibited an MIC ≥2 µg/mL to ceftiofur or ceftriaxone. The retest results have been included in the NARMS annual reports since 2003.

Serotype Confirmation/Categorization

Salmonella serotype reported by the submitting laboratory was used for reporting with few exceptions. Serotype was confirmed by CDC for isolates that underwent subsequent molecular analysis for publication. Because of challenges associated with interpretation of tartrate fermentation assays, ability to ferment tartrate was confirmed for isolates reported as *Salmonella* ser. Paratyphi B by the submitting laboratory (serotype Paratyphi B is by definition unable to ferment L(+) tartrate). To distinguish *Salmonella* serotypes Paratyphi B and Paratyphi B var. L(+) tartrate+ (formerly serotype Java), CDC performed Jordan's tartrate test and/or Kauffmann's tartrate test on all *Salmonella* ser. Paratyphi B isolates from 1996 to 2010 for which the tartrate result was not reported or was reported to be negative. Isolates negative for tartrate fermentation by both assays were categorized as serotype Paratyphi B. Isolates that were positive for tartrate fermentation by either assay were categorized as serotype Paratyphi B var. L(+) tartrate+. Confirmation of other biochemical reactions or somatic and flagellar antigens was not performed at CDC.

Because of increased submissions of *Salmonella* ser. I 4,[5],12:i:- noted in previous years, and recognition of the possibility that this serotype may have been underreported in previous years, isolates reported as serogroup B and tested in NARMS during 1996 to 2010 were reviewed for additional information; isolates that could be clearly identified as serogroup B, first-phase flagellar antigen "i", second phase flagellar antigen absent were categorized in this report as *Salmonella* ser. I 4,[5],12:i:-.

Testing of *Campylobacter*

Changes in Sampling Scheme in 2010

The number of isolates received from Georgia, Maryland, and New Mexico increased over time. To avoid oversampling from these sites, instead of testing all isolates that had been received for 2010, the scheme for testing isolates was changed to every other isolate from Georgia and Maryland and every third from New Mexico. The sampling scheme was adjusted to reflect these changes.

Changes in Testing Methods in 2005

Starting in 2005, there were four changes in the methodology used for *Campylobacter*. First, a surveillance scheme for selecting a representative sample of *Campylobacter* isolates for submission by FoodNet sites was implemented. State public health laboratories within FoodNet sites receive *Campylobacter* isolates from reference and clinical laboratories within their state. In 2005, FoodNet sites changed from submitting the first isolate received each week to submitting every isolate (Georgia, Maryland, New Mexico, Oregon, and Tennessee), every other isolate (California, Colorado, Connecticut, and New York), or every fifth isolate received (Minnesota). The number of laboratories submitting isolates ranged from two to all. Second, the method of species identification was updated to parallel what is used by the CDC National *Campylobacter* Laboratory. Third, the susceptibility testing method changed from Etest® (AB bioMerieux, Solna, Sweden) to broth microdilution. Fourth, there were changes in the antimicrobial agents tested. Florfenicol replaced chloramphenicol as the phenicol class representative drug, and telithromycin was added to the NARMS panel of agents tested. These methods began in 2005 and continue through the current year's report except for noted changes to submissions from Georgia, Maryland, and New Mexico beginning in 2010.

Identification/Speciation and Antimicrobial Susceptibility Testing

From 2005 through 2010, isolates were confirmed as *Campylobacter* by determination of typical morphology and motility using dark-field microscopy and a positive oxidase test reaction. Identification of *C. jejuni* was performed using the hippurate hydrolysis test. Hippurate-positive isolates were identified as *C. jejuni*. Hippurate-negative isolates were further characterized with polymerase chain reaction (PCR) assays with specific targets for *C. jejuni* (*mapA* or *hipO* gene), *C. coli*-specific *ceuE* gene (Linton *et al.* 1997, Gonzales *et al.* 1997, Pruckler *et al.* 2006), or other species specific primers. The only changes for 2010 include all *jejuni* and suspected *coli* isolates were confirmed through a multiplex PCR (Vandamme *et al.* 1997) and the *ceuE* PCR was not used. From 2003 to 2004, putative *Campylobacter* isolates were identified as *C. jejuni* or *C. coli* using BAX® System PCR Assay according to the manufacturer's instructions (DuPont Qualicon, Wilmington, DE). Isolates not identified as *C. jejuni* or *C. coli* were further characterized by other PCR assays (Linton *et al.* 1996) or were characterized by the

CDC National *Campylobacter* Reference Laboratory. From 1997 to 2002, methodology similar to that used from 2005 to 2009 was used.

The methods for susceptibility testing *Campylobacter* and criteria for interpreting the results have changed during the course of NARMS surveillance. Beginning in 2005, broth microdilution using the Sensititre® system (Trek Diagnostics, Cleveland, OH) was performed according to manufacturer’s instructions to determine the MICs for nine antimicrobial agents: azithromycin, ciprofloxacin, clindamycin, erythromycin, florfenicol, gentamicin, nalidixic acid, telithromycin, and tetracycline (Table 4). CLSI recommendations for quality control were followed. From 1997 to 2004, Etest® (AB bioMerieux, Solna, Sweden) was used for susceptibility testing of *Campylobacter* isolates. *Campylobacter*-specific CLSI interpretive criteria were used for erythromycin, ciprofloxacin, and tetracycline beginning with the 2004 NARMS annual report. NARMS breakpoints were used when CLSI breakpoints were not available. Beginning in 2004, NARMS breakpoints were established based on the MIC distributions of NARMS isolates and the presence of known resistance genes or mutations. In pre-2004 annual reports, NARMS breakpoints used were based on those available for other organisms. Establishment of breakpoints based on MIC distributions resulted in higher MIC definitions for azithromycin and erythromycin resistance compared with those reported in pre-2004 annual reports. The breakpoints listed in Table 4 have been applied to MIC data collected for all years so that resistance prevalence is comparable over time. Repeat testing of isolates was done based on criteria in Appendix B.

Table 4. Antimicrobial agents used for susceptibility testing of *Campylobacter* isolates, NARMS, 1997–2010

CLSI class	Antimicrobial Agent	Antimicrobial Agent Concentration Range (µg/mL)	MIC Interpretive Standard (µg/mL)		
			Susceptible	Intermediate	Resistant
Aminoglycosides	Gentamicin	0.12–32 0.016–256*	≤2	4	≥8
Ketolides	Telithromycin†	0.015–8	≤4	8	≥16
Lincosamides	Clindamycin	0.03–16 0.016–256*	≤2	4	≥8
Macrolides	Azithromycin	0.015–64 0.016–256*	≤2	4	≥8
	Erythromycin	0.03–64 0.016–256*	≤8	16	≥32
Phenicol	Chloramphenicol‡	0.016–256*	≤8	16	≥32
	Florfenicol§	0.03–64	≤4	N/A	N/A
Quinolones	Ciprofloxacin	0.015–64 0.002–32*	≤1	2	≥4
	Nalidixic acid	4–64 0.016–256*	≤16	32	≥64
Tetracyclines	Tetracycline	0.06–64 0.016–256*	≤4	8	≥16

N/A indicates that no MIC range of either intermediate or resistant susceptibility exists

* Etest dilution range used from 1997–2004.

† Telithromycin added to NARMS panel in 2005.

‡ Chloramphenicol, tested from 1997–2004, was replaced by florfenicol in 2005.

§ Currently only a susceptible breakpoint (≤4 µg/mL) has been established. In this report isolates with a MIC ≥8 µg/mL are categorized as resistant.

Testing of *Vibrio* species other than *V. cholera*

NARMS participating public health laboratories were asked to forward every isolate of *Vibrio* species other than *V. cholerae* they received to CDC for antimicrobial susceptibility testing by the NARMS laboratory and, in some cases, confirmation of identity by CDC's National Enteric Reference Laboratory. Minimum inhibitory concentrations were determined by Etest® (AB bioMerieux, Solna, Sweden) according to manufacturer's instructions for 9 drugs: ampicillin, cephalothin, chloramphenicol, ciprofloxacin, kanamycin, nalidixic acid, streptomycin, tetracycline, and trimethoprim-sulfamethoxazole (Table 5). CLSI breakpoints specific for *Vibrio* species other than *V. cholera* were available for ampicillin, ciprofloxacin, tetracycline, and trimethoprim-sulfamethoxazole. Frequency of isolates susceptible, intermediate, and resistant for those drugs is shown in this report. MIC distributions are shown for drugs that do not have CLSI breakpoints. Identity confirmation is not yet complete for all isolates submitted in 2010, so results for isolates submitted in 2009 are presented in this report.

Table 5. Antimicrobial agents used for susceptibility testing of *Vibrio* species other than *V. cholerae* isolates, NARMS, 2009

CLSI class	Antimicrobial Agent	Antimicrobial Agent Concentration Range (µg/mL)	MIC Interpretive Standard (µg/mL)		
			Susceptible	Intermediate*	Resistant
Aminoglycosides	Kanamycin†	0.016-256			
	Streptomycin†	0.064-1024			
Cephems	Cephalothin†	0.016-256			
Folate pathway inhibitors	Trimethoprim-sulfamethoxazole	0.002-32	≤2/38	N/A	≥4/76
Penicillins	Ampicillin	0.016-256	≤8	16	≥32
Phenicol	Chloramphenicol†	0.016-256			
Quinolones	Ciprofloxacin	0.002-32	≤1	2	≥4
	Nalidixic acid†	0.016-256			
Tetracyclines	Tetracycline	0.016-256	≤4	8	≥16

* N/A indicates that no MIC range of intermediate susceptibility exists

† No CLSI or NARMS breakpoints established

Testing of Representative Bacteria from Outbreaks

CDC has often tested human clinical isolates of bacteria from selected foodborne disease outbreaks for various identification and subtyping purposes. Since 2004, efforts to characterize antimicrobial susceptibility of bacteria associated with foodborne disease outbreaks have increased, and CDC requests for state health departments to submit such isolates for this purpose have become more formal. Since 2006, all NARMS participating laboratories have been asked to forward 3 representative isolates from each outbreak of *Salmonella enterica* serotype Enteritidis, Newport, and Typhimurium. Also since 2006, FoodNet sites were asked to submit 3 representative isolates from all *Salmonella* outbreaks, regardless of serotype. The methods used for susceptibility testing were the same as those performed for *Salmonella* submitted for NARMS routine surveillance. A summary of antimicrobial susceptibility data of non-typhoidal *Salmonella* isolates tested in NARMS and available data from CDC's Foodborne Disease Outbreak Surveillance System for outbreaks from 2004 through 2008 are presented in this report in [Appendix A](#).

Data Analysis

For all pathogens, MICs were categorized as resistant, intermediate (if applicable), or susceptible. Analysis was restricted to the first isolate received (per genus under surveillance) per patient in the calendar year. If two or more isolates were received for the same patient for *Salmonella* ser. Typhi, the first blood isolate collected would be included in analysis. If no blood isolates were submitted, the first isolate collected would be included in

analysis. The 95% confidence intervals (CIs) for the percentage of resistant isolates are included in the MIC distribution tables. The 95% CIs were calculated using the Paulson-Camp-Pratt approximation method.

When describing results for several years, multidrug resistance for *Salmonella*, *Shigella*, and *E. coli* O157 isolates was limited to the eight CLSI classes ([Table 3](#)) represented by the following 15 agents: amikacin, amoxicillin-clavulanic acid, ampicillin, cefoxitin, ceftiofur, ceftriaxone, chloramphenicol, ciprofloxacin, gentamicin, kanamycin, nalidixic acid, streptomycin, sulfamethoxazole/sulfisoxazole, tetracycline, and trimethoprim-sulfamethoxazole. Isolates that were not resistant to any of these 15 agents were considered to have no resistance detected. When describing multidrug resistance for several years for *Campylobacter* isolates, multidrug resistance was limited to the six CLSI classes represented by azithromycin, ciprofloxacin, chloramphenicol/florfenicol, clindamycin, erythromycin, gentamicin, nalidixic acid, and tetracycline ([Table 4](#)). *Campylobacter* isolates that were not resistant to any of these agents were considered to have no resistance detected.

Logistic regression was used to compare the prevalence of antimicrobial resistance among *Salmonella* and *Campylobacter* isolates tested in 2010 with the reference, which was the average prevalence of resistance in the first five years that NARMS surveillance was nationwide (2003–07). The analysis included the following:

1. Non-typhoidal *Salmonella*: resistance to nalidixic acid, resistance to ceftriaxone, resistance to one or more CLSI classes, resistance to three more CLSI classes
2. *Salmonella* ser. Enteritidis: resistance to nalidixic acid
3. *Salmonella* ser. Typhimurium: resistance to at least ACSSuT (ampicillin, chloramphenicol, streptomycin, sulfonamide, and tetracycline)
4. *Salmonella* ser. Newport: resistance to at least ACSSuTAuCx (ACSSuT, amoxicillin-clavulanic acid, and ceftriaxone)
5. *Salmonella* ser. Typhi: resistance to nalidixic acid
6. *Campylobacter* species: resistance to ciprofloxacin
7. *Campylobacter jejuni*: resistance to ciprofloxacin

To account for site-to-site variation in the prevalence of antimicrobial resistance, we included main effects adjustments for site in the analysis. The final regression models for *Salmonella* adjusted for the submitting site using the nine geographic regions described by the [U.S. Census Bureau](#): East North Central, East South Central, Mid-Atlantic, Mountain, New England, Pacific, South Atlantic, West North Central, and West South Central. For *Campylobacter*, the final regression models adjusted for the submitting FoodNet site. Odds ratios (ORs) and 95% confidence intervals (CIs) were calculated using unconditional maximum likelihood estimation. The adequacy of model fit was assessed in several ways. The significance of the main effect of year was assessed using the likelihood ratio test. The likelihood ratio test was also used to test for significance of interaction between site and year, although the power of the test to detect a single site-specific interaction was low. The Hosmer and Lemeshow goodness-of-fit test was also used ([Fleiss, et al.](#)). Having assessed that the main effect of year was significant, we reported ORs with 95% CIs (for 2010 compared with reference) that did not include 1.00 as statistically significant.

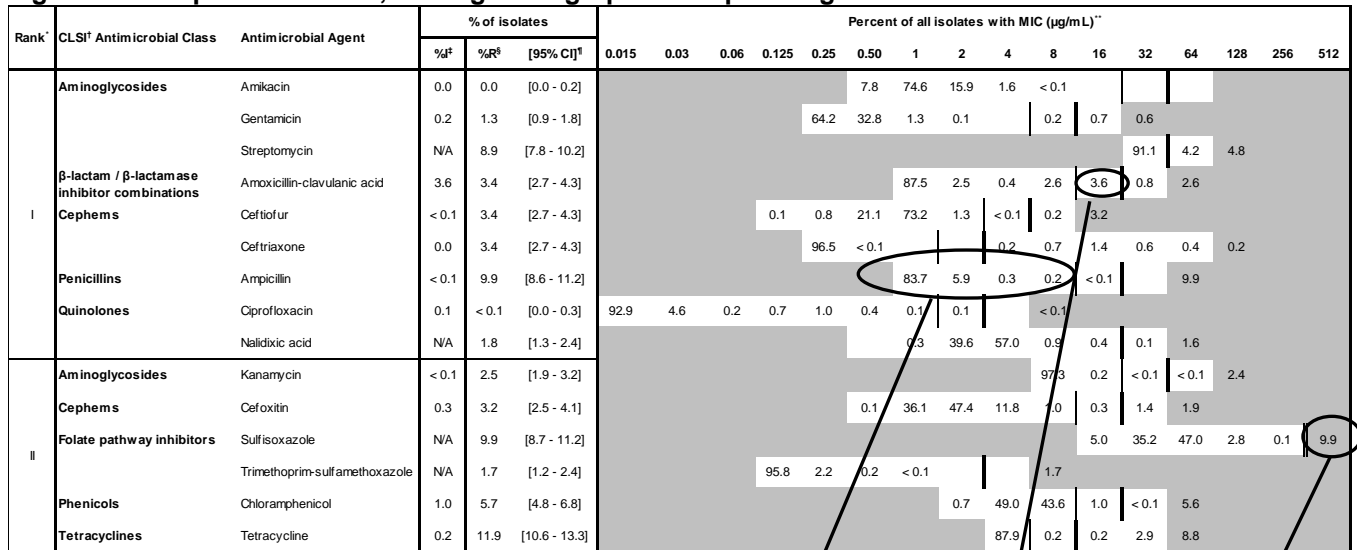
MIC Distribution Tables and Proportional Figures

An explanation on “how to read a squashtogram” has been provided to assist the reader with the different parts of each table (Figure 11). A squashtogram shows the distribution of MICs for antimicrobial agents tested. Proportional figures visually display data from squashtograms for an immediate comparative summary of resistance in specific pathogens and serotypes. These figures are a categorical visual aid for the interpretation of MIC values. For most antimicrobial agents tested, three categories (susceptible, intermediate, and resistant) are used to interpret MICs. The proportion representing each category is shown in a horizontal proportional bar chart (Figure 12).

Figure 11. How to read a squashtogram

Rank	CLSI [†] Antimicrobial Class	Antimicrobial Agent	% of isolates			Percent of all isolates with MIC (µg/mL) [‡]																				
			% [†]	%R [§]	[95% CI] [†]	0.015	0.03	0.06	0.125	0.25	0.5	1	2	4	8	16	32	64	128	256	512					
I	Aminoglycosides	Amikacin	0.0	0.0	[0.0–0.2]						7.4	70.1	20.8	1.6	0.1											
		Genamycin	0.1	2.1	[1.5–2.8]												0.1	0.9	1.2							
	β-lactam / β-lactamase inhibitor combinations	Streptomycin	N/A	10.4	[9.1–11.7]																	6.0				
		Amoxicillin-clavulanic acid	4.2	3.3	[2.6–4.1]							84.8	4.9	0.4	2.5		4.2	0.6	2.7							
	Cepheems	Ceftiofur	0.0	3.2	[2.6–4.1]			0.3	0.8	27.5	66.7	1.4				0.1	3.1									
		Ceftriaxone	2.3	0.4	[0.2–0.8]					96.7						0.1	0.5	1.4								
	Penicillins	Ampicillin	0.0	10.1	[8.9–11.5]							81.2	8.3	0.3	0.1							0.1	10.0			
		Quinolones	Ciprofloxacin	0.0	0.1	[0.0–0.3]	92.9	4.4	0.2	1.3	0.8	0.3														
			Nalidixic acid	N/A	2.2	[1.7–3.0]						0.1	0.2	34.4	61.9	0.9	0.2						2.2			
	II	Aminoglycosides	Kanamycin	< 0.1	2.8	[2.2–3.6]											96.8	0.2	< 0.1	0.2	2.6					
Cepheems		Cefepime	0.7	3.0	[2.3–3.7]							0.2	8.8	70.2	15.8	1.3						0.7	0.9	2.1		
		Sulfisoxazole	N/A	12.3	[11.0–13.8]																	19.0	52.1	15.0	0.5	0.1
Folate pathway inhibitors		Trimethoprim-sulfamethoxazole	N/A	1.6	[1.1–2.2]					79.7	18.3					1.5										
		Phenicol	Chloramphenicol	0.7	7.3	[6.2–8.5]								0.8	41.7	49.5	0.7									
Tetracyclines			Tetracycline	0.1	14.5	[13.0–16.0]										85.4	0.1	0.9	4.2	9.4						

Figure 12. Proportional chart, a categorical graph of a squashtogram

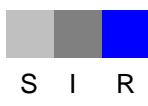
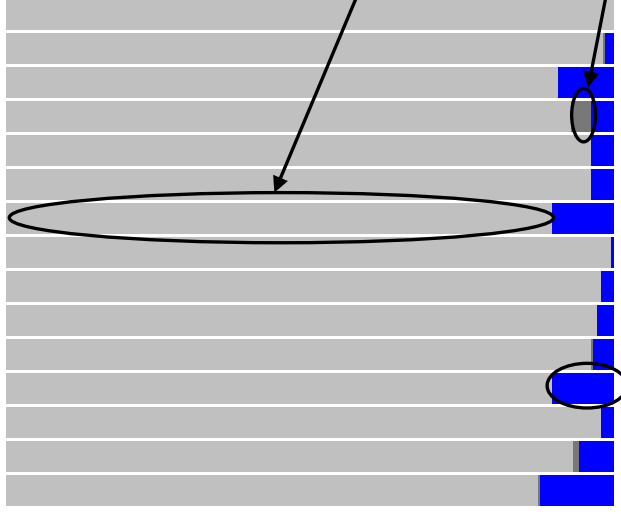


* Rank of antimicrobials based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically important; Rank 2, Highly important
 † CLSI: Clinical and Laboratory Standards Institute
 ‡ Percent of isolates w/ intermediate susceptibility, N/A if no MIC range of intermediate susceptibility exists
 § Percent of isolates that were resistant
 ¶ The 95% confidence intervals (CI) for percent resistant (%R) were calculated using the Paulson-Camp-Pratt approximation to the Clopper-Pearson exact method. The 95% CI is presented to summarize uncertainty in the observed resistance (R%).
 ** The unshaded areas indicate the dilution range of the Sensititre plates used to test isolates. Single vertical bars indicate the breakpoints for susceptibility, while double vertical bars indicate breakpoints for resistance. Numbers in the shaded areas indicate the percentages of isolates with MICs greater than the highest concentrations on the Sensititre plate. Numbers listed for the lowest tested concentrations represent the percentages of isolates with MICs equal to or less than the lowest tested concentration. CLSI breakpoints were used when available.

Antimicrobial Agent

- Amikacin
- Gentamicin
- Streptomycin
- Amoxicillin-clavulanic acid
- Ceftiofur
- Ceftriaxone
- Ampicillin
- Ciprofloxacin
- Nalidixic acid
- Kanamycin
- Cefoxitin
- Sulfisoxazole
- Trimethoprim-sulfamethoxazole
- Chloramphenicol
- Tetracycline

Susceptible, Intermediate, and Resistant Proportion



Results

1. Non-typhoidal *Salmonella*

Table 6. Number and percentage of isolates with resistance to at least ACSSuT, ACSSuTAuCx, nalidixic acid, and ceftriaxone among the 20 most common non-typhoidal *Salmonella* serotypes isolated in NARMS, 2010

Rank	Serotype	N	ACSSuT*		ACSSuTAuCx [†]		Nalidixic Acid		Ceftriaxone	
			n	(%)	n	(%)	n	(%)	n	(%)
1	Enteritidis	522	0	(0)	0	(0)	27	(55.1)	0	(0)
2	Typhimurium	366	68	(63.6)	7	(21.2)	5	(10.2)	18	(25.7)
3	Newport	305	22	(20.6)	22	(66.7)	1	(2.0)	22	(31.4)
4	Javiana	178	0	(0)	0	(0)	0	(0)	1	(1.4)
5	I 4,[5],12:i:-	77	1	(0.9)	0	(0)	2	(4.1)	2	(2.9)
6	Heidelberg	62	1	(0.9)	0	(0)	0	(0)	15	(21.4)
7	Saintpaul	60	0	(0)	0	(0)	0	(0)	0	(0)
8	Montevideo	60	0	(0)	0	(0)	0	(0)	0	(0)
9	Braenderup	57	0	(0)	0	(0)	0	(0)	0	(0)
10	Infantis	55	1	(0.9)	1	(3.0)	0	(0)	2	(2.9)
11	Paratyphi B var. L(+) tartrate+	54	7	(6.5)	0	(0)	0	(0)	0	(0)
12	Muenchen	52	0	(0)	0	(0)	0	(0)	0	(0)
13	Agona	43	0	(0)	0	(0)	1	(2.0)	0	(0)
14	Oranienburg	40	0	(0)	0	(0)	0	(0)	0	(0)
15	Thompson	24	0	(0)	0	(0)	0	(0)	0	(0)
16	Mbandaka	24	0	(0)	0	(0)	0	(0)	0	(0)
17	Mississippi	23	0	(0)	0	(0)	0	(0)	0	(0)
18	Anatum	20	0	(0)	0	(0)	0	(0)	0	(0)
19	Schwarzengrund	19	0	(0)	0	(0)	0	(0)	0	(0)
20	Stanley	18	0	(0)	0	(0)	0	(0)	0	(0)
Subtotal		2059	100	(93.5)	30	(90.9)	36	(73.5)	60	(85.7)
	All other serotypes	370	6	(5.6)	3	(9.1)	10	(20.4)	9	(12.9)
	Unknown serotype	18	1	(0.9)	0	(0)	0	(0)	0	(0)
	Partially serotyped	12	0	(0)	0	(0)	0	(0)	0	(0)
	Rough/Nonmotile isolates	15	0	(0)	0	(0)	3	(6.1)	1	(1.4)
Total		2474	107	(100)	33	(100)	49	(100)	70	(100)

* ACSSuT: at least resistant to ampicillin, chloramphenicol, streptomycin, sulfisoxazole, tetracycline

† ACSSuTAuCx: at least resistant to ACSSuT, amoxicillin-clavulanic acid, and ceftriaxone

Table 7. Minimum inhibitory concentrations (MICs) and resistance of non-typhoidal *Salmonella* isolates to antimicrobial agents, 2010 (N=2474)

Rank	CLSI [†] Antimicrobial Class	Antimicrobial Agent	% of isolates			Percent of all isolates with MIC (µg/mL)**																		
			% [‡]	%R [§]	[95% CI] [¶]	0.015	0.03	0.06	0.125	0.25	0.50	1	2	4	8	16	32	64	128	256	512			
I	Aminoglycosides	Amikacin	0.0	0.0	[0.0 - 0.1]						4.4	73.3	20.2	2.0	0.2									
		Gentamicin	0.2	1.0	[0.6 - 1.4]					66.9	30.3	1.4	0.2		0.2	0.4	0.6							
		Streptomycin	N/A	8.6	[7.5 - 9.7]													91.4	3.6	4.9				
	β-lactam / β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid	3.3	2.8	[2.2 - 3.6]							89.1	1.5	0.8	2.5	3.3	0.8	2.1						
		Cephems																						
		Ceftiofur	< 0.1	2.8	[2.2 - 3.5]			0.2	0.4	32.7	63.1	0.8		< 0.1	< 0.1	2.7								
		Ceftriaxone	0.0	2.8	[2.2 - 3.6]					97.1	< 0.1					0.2	1.2	1.1	0.2	0.2				
		Penicillins																						
		Ampicillin	< 0.1	9.1	[8.0 - 10.3]							85.1	5.5	0.2	< 0.1	< 0.1	< 0.1	9.0						
		Quinolones	Ciprofloxacin	0.0	0.2	[0.0 - 0.4]	93.5	3.6	0.2	0.9	0.9	0.7	< 0.1		< 0.1	0.1								
	Nalidixic acid		N/A	2.0	[1.5 - 2.6]							0.2	33.3	63.3	0.6	0.5	< 0.1	1.9						
	Tetracyclines	Tetracycline	0.1	11.0	[9.8 - 12.3]											88.8	0.1	0.4	2.7	7.9				
II	Aminoglycosides	Kanamycin	0.0	2.3	[1.7 - 2.9]												97.7	< 0.1	< 0.1	2.2				
	Cephems	Cefoxitin	0.4	2.5	[2.0 - 3.2]							19.2	65.2	11.8	0.9	0.4	1.0	1.5						
	Folate pathway inhibitors	Sulfisoxazole	N/A	9.0	[7.9 - 10.2]													5.0	36.1	47.6	2.1	< 0.1	9.0	
		Trimethoprim-sulfamethoxazole	N/A	1.6	[1.1 - 2.1]				97.0	1.3	0.1				< 0.1	1.5								
	Phenicol	Chloramphenicol	0.6	4.9	[4.1 - 5.9]									0.3	34.2	60.0	0.6	< 0.1	4.9					

* Rank of antimicrobial agents is based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important
 † CLSI: Clinical and Laboratory Standards Institute
 ‡ Percent of isolates with intermediate susceptibility; N/A indicates that no MIC range of intermediate susceptibility exists
 § Percent of isolates that were resistant
 ¶ The 95% confidence intervals (CI) for percent resistant (%R) were calculated using the Paulson-Camp-Pratt approximation to the Clopper-Pearson exact method
 ** The unshaded areas indicate the dilution range of the Sensititre plates used to test isolates. Single vertical bars indicate the breakpoints for susceptibility, while double vertical bars indicate breakpoints for resistance. Numbers in the shaded areas indicate the percentages of isolates with MICs greater than the highest concentrations on the Sensititre plate. Numbers listed for the lowest tested concentrations represent the percentages of isolates with MICs equal to or less than the lowest tested concentration. CLSI breakpoints were used when available.

Figure 13. Antimicrobial resistance pattern for non-typhoidal *Salmonella*, 2010

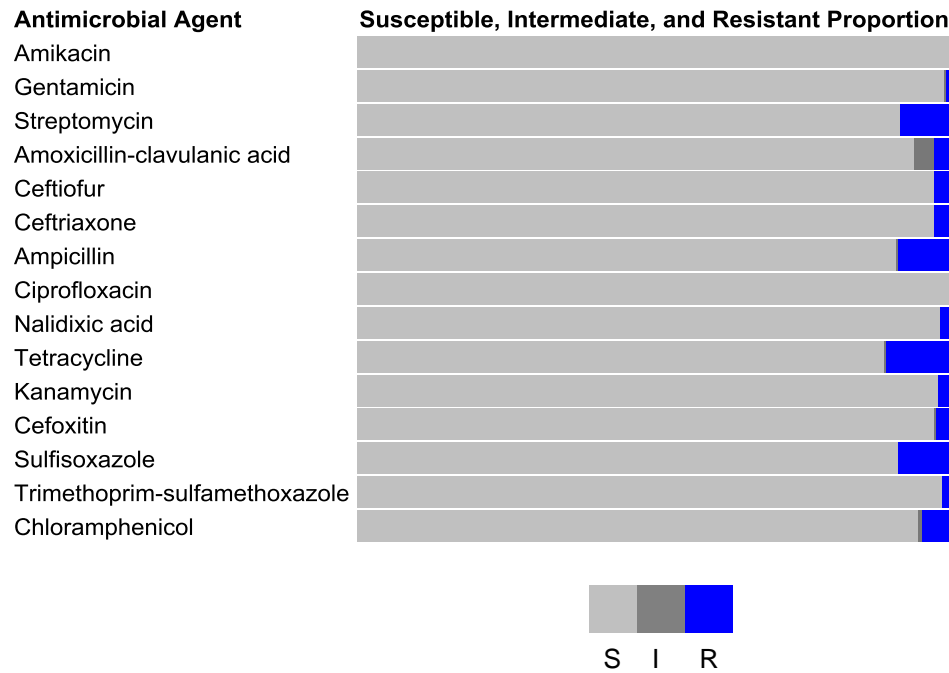


Table 8. Percentage and number of non-typhoidal *Salmonella* isolates resistant to antimicrobial agents, 2001–2010

Year		2001	2002	2003	2004	2005	2006	2007	2008	2009	2010	
Total Isolates		1410	1998	1855	1782	2034	2172	2145	2384	2193	2474	
Rank	CLSI [†] Antimicrobial Class	Antimicrobial Agent (Resistance breakpoint)										
I	Aminoglycosides	Amikacin (MIC ≥ 64)	0.0% 0	0.0% 0	0.0% 0	0.0% 0	< 0.1% 1	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0
		Gentamicin (MIC ≥ 16)	1.9% 27	1.4% 27	1.4% 26	1.3% 24	2.2% 44	2.0% 44	2.1% 45	1.5% 35	1.3% 28	1.0% 24
		Streptomycin (MIC ≥ 64)	17.1% 241	13.2% 264	15.0% 279	12.0% 213	11.1% 225	10.7% 233	10.3% 222	10.0% 238	8.9% 196	8.6% 212
	β-lactam/β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid (MIC ≥ 32/16)	4.7% 66	5.3% 106	4.6% 86	3.7% 66	3.2% 65	3.7% 81	3.3% 70	3.1% 73	3.4% 75	2.8% 70
		Cepheems	Ceftiofur (MIC ≥ 8)	4.1% 58	4.4% 87	4.5% 83	3.4% 60	2.9% 60	3.6% 79	3.3% 70	3.1% 73	3.4% 75
			Ceftriaxone (MIC ≥ 4)	3.7% 52	4.4% 87	4.4% 81	3.3% 59	2.9% 59	3.7% 80	3.3% 70	3.1% 73	3.4% 75
	Penicillins	Ampicillin (MIC ≥ 32)	17.5% 247	13.0% 259	13.6% 253	12.1% 216	11.4% 232	11.0% 238	10.1% 217	9.7% 232	9.8% 216	9.1% 224
	Quinolones	Ciprofloxacin (MIC ≥ 4)	0.2% 3	0.1% 1	0.2% 3	0.2% 4	< 0.1% 1	0.1% 2	0.1% 2	0.1% 2	< 0.1% 1	0.2% 4
		Nalidixic Acid (MIC ≥ 32)	2.3% 32	1.6% 32	1.9% 36	2.2% 39	1.9% 38	2.4% 52	2.2% 48	2.1% 49	1.8% 39	2.0% 49
	Tetracyclines	Tetracycline (MIC ≥ 16)	19.9% 280	14.9% 298	16.3% 303	13.6% 242	13.9% 282	13.5% 293	14.5% 310	11.5% 275	11.9% 261	11.0% 273
II	Aminoglycosides	Kanamycin (MIC ≥ 64)	4.8% 68	3.8% 76	3.5% 64	2.8% 50	3.4% 70	2.9% 63	2.8% 61	2.1% 50	2.5% 54	2.3% 56
		Cepheems	Cefoxitin (MIC ≥ 32)	3.4% 48	4.3% 86	4.3% 79	3.4% 61	3.0% 62	3.5% 77	2.9% 63	3.0% 72	3.2% 71
	Cephalothin (MIC ≥ 32)		4.0% 57	5.1% 101	5.3% 99	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested
	Folate pathway inhibitors	Sulfamethoxazole/Sulfisoxazole [‡] (MIC ≥ 5/12)	17.8% 251	12.9% 258	15.1% 280	13.3% 237	12.6% 256	12.1% 263	12.3% 264	10.1% 240	9.9% 217	9.0% 223
		Trimethoprim-sulfamethoxazole (MIC ≥ 4/76)	2.0% 28	1.4% 28	1.9% 36	1.7% 31	1.7% 34	1.7% 36	1.5% 33	1.6% 37	1.7% 38	1.6% 39
Phenicol	Chloramphenicol (MIC ≥ 32)	11.6% 164	8.6% 172	10.1% 187	7.6% 136	7.8% 159	6.4% 139	7.3% 156	6.1% 146	5.7% 125	4.9% 122	

* Rank of antimicrobial agents is based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important

† CLSI: Clinical and Laboratory Standards Institute

‡ Sulfamethoxazole, which was tested during 1996-2003 to represent sulfonamides, was replaced by sulfisoxazole in 2004

Table 9. Resistance patterns of non-typhoidal *Salmonella* isolates, 2001–2010

Year	2001	2002	2003	2004	2005	2006	2007	2008	2009	2010
Total Isolates	1410	1998	1855	1782	2034	2172	2145	2384	2193	2474
Resistance Pattern										
No resistance detected	72.5% 1022	79.1% 1580	78.0% 1447	80.0% 1425	80.9% 1646	80.5% 1748	81.1% 1739	83.9% 2000	83.2% 1824	84.7% 2095
Resistance ≥ 1 CLSI class*	27.5% 388	20.9% 418	22.0% 408	20.0% 357	19.1% 388	19.5% 424	18.9% 406	16.1% 384	16.8% 369	15.3% 379
Resistance ≥ 2 CLSI classes*	22.1% 311	15.8% 315	17.5% 325	15.0% 267	14.8% 302	14.7% 319	14.2% 305	12.5% 298	13.0% 284	11.3% 279
Resistance ≥ 3 CLSI classes*	16.7% 236	12.3% 245	14.2% 263	11.4% 204	12.0% 244	11.8% 256	11.1% 239	9.5% 226	9.5% 209	9.1% 225
Resistance ≥ 4 CLSI classes*	13.5% 191	9.8% 195	11.4% 211	9.3% 165	9.1% 185	8.1% 177	8.2% 176	7.4% 177	7.3% 159	6.8% 167
Resistance ≥ 5 CLSI classes*	10.3% 145	8.2% 164	9.8% 182	8.0% 142	7.2% 146	6.3% 137	6.9% 149	6.6% 157	6.2% 137	5.2% 128
At least ACSSuT [†]	10.1% 142	7.8% 156	9.3% 173	7.2% 129	6.9% 141	5.6% 121	6.3% 136	5.8% 138	5.1% 112	4.3% 107
At least ACT/S [‡]	0.5% 7	1.1% 21	1.2% 23	0.6% 10	0.9% 18	0.7% 15	0.7% 16	0.5% 11	0.7% 15	0.4% 11
At least ACSSuTAuCx [§]	2.6% 36	3.4% 67	3.2% 60	2.4% 42	2.0% 41	2.0% 43	2.1% 46	1.8% 44	1.4% 30	1.3% 33
At least ceftriaxone and nalidixic acid resistant	0.1% 2	0.2% 4	0.1% 1	0.1% 2	0.0% 1	0.2% 4	0.2% 5	0.0% 1	0.2% 4	0.1% 2

* CLSI: Clinical and Laboratory Standards Institute

† ACSSuT: resistance to ampicillin, chloramphenicol, streptomycin, sulfamethoxazole/sulfisoxazole, tetracycline

‡ ACT/S: resistance to ampicillin, chloramphenicol, trimethoprim-sulfamethoxazole

§ ACSSuTAuCx: resistance to ACSSuT, amoxicillin-clavulanic acid, ceftriaxone

A. Salmonella ser. Enteritidis

Table 10. Minimum inhibitory concentrations (MICs) and resistance of *Salmonella ser. Enteritidis* isolates to antimicrobial agents, 2010 (N=522)

Rank	CLSI† Antimicrobial Class	Antimicrobial Agent	% of Isolates			Percent of all isolates with MIC (µg/mL)**																	
			%‡	%R§	[95% CI]¶	0.015	0.03	0.06	0.125	0.25	0.50	1	2	4	8	16	32	64	128	256	512		
I	Aminoglycosides	Amikacin	0.0	0.0	[0.0 - 0.7]						10.3	79.1	9.4	1.1									
		Gentamicin	0.2	0.2	[0.0 - 1.1]				83.5	15.7	0.2	0.2		0.2			0.2						
		Streptomycin	N/A	0.6	[0.1 - 1.7]													99.4	0.4	0.2			
	β-lactam / β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid	0.2	0.4	[0.0 - 1.4]						95.2	2.1	0.2	1.9		0.2		0.2		0.2			
		Cephems							0.2	0.2	4.2	94.4	1.0										
		Ceftriaxone	0.0	0.0	[0.0 - 0.7]				100.0														
	Penicillins	Ampicillin	0.0	2.3	[1.2 - 4.0]						82.2	15.1	0.2	0.2							2.3		
	Quinolones	Ciprofloxacin	0.0	0.2	[0.0 - 1.1]	83.3	11.3	0.4	2.5	1.9	0.4							0.2					
		Nalidixic acid	N/A	5.2	[3.4 - 7.4]						0.4	10.9	82.8	0.6	0.2						5.2		
Tetracyclines	Tetracycline	0.0	2.1	[1.1 - 3.7]								97.9								2.1			
II	Aminoglycosides	Kanamycin	0.0	0.2	[0.0 - 1.1]												99.8				0.2		
	Cephems	Cefoxitin	0.2	0.0	[0.0 - 0.7]						11.3	82.4	5.0	1.1		0.2							
	Folate pathway inhibitors	Sulfisoxazole	N/A	1.9	[0.9 - 3.5]												2.7	26.2	66.3	2.9		1.9	
		Trimethoprim-sulfamethoxazole	N/A	1.0	[0.3 - 2.2]				98.5	0.6					1.0								
Phenicol	Chloramphenicol	0.4	0.6	[0.1 - 1.7]								35.8	63.2		0.4	0.2		0.4					

* Rank of antimicrobial agents is based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important
 † CLSI: Clinical and Laboratory Standards Institute
 ‡ Percent of isolates with intermediate susceptibility; N/A indicates that no MIC range of intermediate susceptibility exists
 § Percent of isolates that were resistant
 ¶ The 95% confidence intervals (CI) for percent resistant (%R) were calculated using the Paulson-Camp-Pratt approximation to the Clopper-Pearson exact method
 ** The unshaded areas indicate the dilution range of the Sensititre plates used to test isolates. Single vertical bars indicate the breakpoints for susceptibility, while double vertical bars indicate breakpoints for resistance. Numbers in the shaded areas indicate the percentages of isolates with MICs greater than the highest concentrations on the Sensititre plate. Numbers listed for the low est tested concentrations represent the percentages of isolates with MICs equal to or less than the low est tested concentration. CLSI breakpoints were used when available.

Figure 14. Antimicrobial resistance pattern for *Salmonella ser. Enteritidis*, 2010

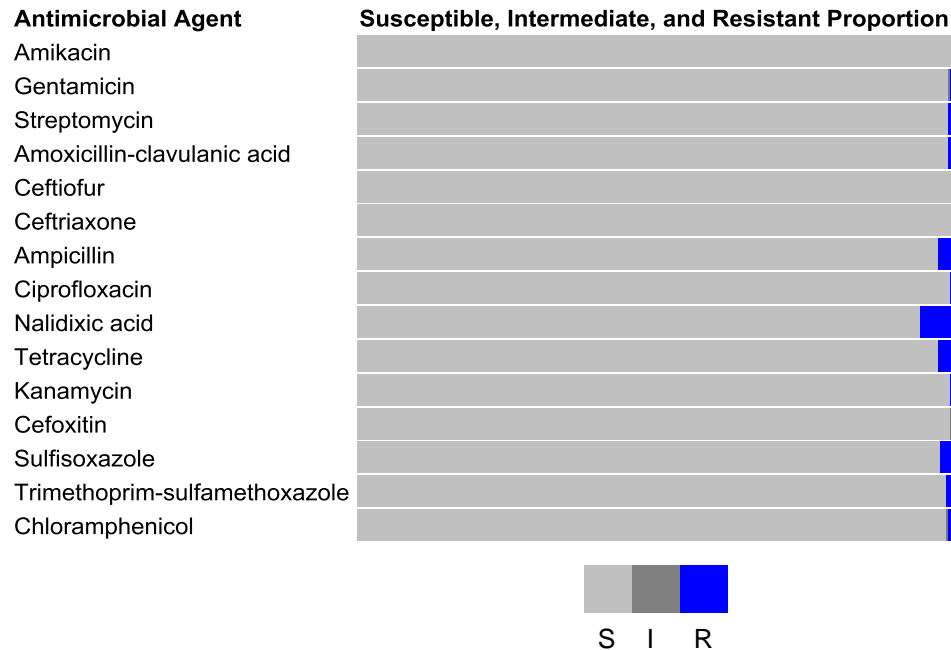


Table 11. Percentage and number of *Salmonella ser. Enteritidis* isolates resistant to antimicrobial agents, 2001–2010

Year		2001	2002	2003	2004	2005	2006	2007	2008	2009	2010	
Total Isolates		277	337	257	271	384	413	385	441	410	522	
Rank*	CLSI† Antimicrobial Class	Antimicrobial Agent (Resistance breakpoint)										
I	Aminoglycosides	Amikacin (MIC ≥ 64)	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0
		Gentamicin (MIC ≥ 16)	0.0% 0	0.3% 1	0.4% 1	0.4% 1	0.8% 3	0.2% 1	0.0% 0	0.2% 1	0.0% 0	0.2% 1
		Streptomycin (MIC ≥ 64)	1.4% 4	1.5% 5	1.2% 3	2.2% 6	1.0% 4	1.2% 5	0.5% 2	0.5% 2	1.2% 5	0.6% 3
	β-lactam/β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid (MIC ≥ 32/16)	1.4% 4	0.6% 2	0.0% 0	0.0% 0	0.8% 3	0.5% 2	0.5% 2	0.0% 0	0.0% 0	0.4% 2
	Cephems	Ceftiofur (MIC ≥ 8)	2.2% 6	0.0% 0	0.0% 0	0.0% 0	0.5% 2	0.5% 2	0.3% 1	0.2% 1	0.0% 0	0.0% 0
		Ceftriaxone (MIC ≥ 4)	1.4% 4	0.0% 0	0.0% 0	0.0% 0	0.3% 1	0.5% 2	0.3% 1	0.2% 1	0.0% 0	0.0% 0
	Penicillins	Ampicillin (MIC ≥ 32)	8.7% 24	6.8% 23	2.3% 6	4.1% 11	2.9% 11	4.4% 18	2.1% 8	3.9% 17	3.9% 16	2.3% 12
	Quinolones	Ciprofloxacin (MIC ≥ 4)	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.2% 1
		Nalidixic Acid (MIC ≥ 32)	4.3% 12	3.9% 13	4.7% 12	6.6% 18	4.7% 18	7.0% 29	5.7% 22	7.0% 31	3.7% 15	5.2% 27
	Tetracyclines	Tetracycline (MIC ≥ 16)	1.8% 5	4.2% 14	1.6% 4	3.3% 9	2.3% 9	1.7% 7	3.9% 15	1.8% 8	1.2% 5	2.1% 11
II	Aminoglycosides	Kanamycin (MIC ≥ 64)	0.7% 2	0.3% 1	0.0% 0	0.7% 2	0.3% 1	0.2% 1	0.5% 2	0.0% 0	0.2% 1	0.2% 1
	Cephems	Cefoxitin (MIC ≥ 32)	0.4% 1	0.0% 0	0.0% 0	0.0% 0	1.0% 4	0.5% 2	0.3% 1	0.0% 0	0.0% 0	0.0% 0
		Cephalothin (MIC ≥ 32)	1.1% 3	0.6% 2	1.2% 3	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested
	Folate pathway inhibitors	Sulfamethoxazole/Sulfisoxazole‡ (MIC ≥ 5/12)	2.2% 6	1.5% 5	1.2% 3	1.8% 5	1.6% 6	1.5% 6	1.6% 6	1.1% 5	1.7% 7	1.9% 10
		Trimethoprim-sulfamethoxazole (MIC ≥ 4/76)	0.7% 2	0.6% 2	0.8% 2	0.0% 0	0.5% 2	0.5% 2	1.0% 4	0.9% 4	0.7% 3	1.0% 5
	Phenicol	Chloramphenicol (MIC ≥ 32)	0.0% 0	0.3% 1	0.4% 1	0.4% 1	0.5% 2	0.0% 0	0.5% 2	0.5% 2	0.0% 0	0.6% 3

* Rank of antimicrobial agents is based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important

† CLSI: Clinical and Laboratory Standards Institute

‡ Sulfamethoxazole, which was tested during 1996-2003 to represent sulfonamides, was replaced by sulfisoxazole in 2004

Table 12. Resistance patterns of *Salmonella ser. Enteritidis* isolates, 2001–2010

Year	2001	2002	2003	2004	2005	2006	2007	2008	2009	2010
Total Isolates	277	337	257	271	384	413	385	441	410	522
Resistance Pattern										
No resistance detected	86.6% 240	87.5% 295	91.8% 236	87.1% 236	91.4% 351	88.6% 366	90.4% 348	87.5% 386	92.0% 377	92.1% 481
Resistance ≥ 1 CLSI class*	13.4% 37	12.5% 42	8.2% 21	12.9% 35	8.6% 33	11.4% 47	9.6% 37	12.5% 55	8.0% 33	7.9% 41
Resistance ≥ 2 CLSI classes*	4.7% 13	3.9% 13	2.3% 6	3.0% 8	3.6% 14	2.9% 12	3.4% 13	2.0% 9	2.4% 10	2.9% 15
Resistance ≥ 3 CLSI classes*	2.9% 8	2.1% 7	0.4% 1	1.1% 3	1.6% 6	1.7% 7	1.0% 4	0.5% 2	1.0% 4	2.1% 11
Resistance ≥ 4 CLSI classes*	1.1% 3	0.6% 2	0.4% 1	0.7% 2	1.0% 4	0.7% 3	0.3% 1	0.0% 0	0.5% 2	0.4% 2
Resistance ≥ 5 CLSI classes*	0.4% 1	0.0% 0	0.4% 1	0.7% 2	0.5% 2	0.2% 1	0.3% 1	0.0% 0	0.2% 1	0.0% 0
At least ACSSuT†	0.0% 0	0.0% 0	0.4% 1	0.4% 1	0.5% 2	0.0% 0	0.3% 1	0.0% 0	0.0% 0	0.0% 0
At least ACT/S‡	0.0% 0	0.0% 0	0.4% 1	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0
At least ACSSuTAuCx§	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.3% 1	0.0% 0	0.3% 1	0.0% 0	0.0% 0	0.0% 0
At least ceftriaxone and nalidixic acid resistant	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.3% 1	0.2% 1	0.0% 0	0.0% 0

* CLSI: Clinical and Laboratory Standards Institute

† ACSSuT: resistance to ampicillin, chloramphenicol, streptomycin, sulfamethoxazole/sulfisoxazole, tetracycline

‡ ACT/S: resistance to ampicillin, chloramphenicol, trimethoprim-sulfamethoxazole

§ ACSSuTAuCx: resistance to ACSSuT, amoxicillin-clavulanic acid, ceftriaxone

B. *Salmonella ser. Typhimurium*

Table 13. Minimum inhibitory concentrations (MICs) and resistance of *Salmonella ser. Typhimurium* isolates to antimicrobial agents, 2010 (N=366)

Rank*	CLSI† Antimicrobial Class	Antimicrobial Agent	% of isolates			Percent of all isolates with MIC (µg/mL)**																
			%‡	%R§	[95% CI]¶	0.015	0.03	0.06	0.125	0.25	0.50	1	2	4	8	16	32	64	128	256	512	
I	Aminoglycosides	Amikacin	0.0	0.0	[0.0 - 1.0]						1.9	71.3	24.3	2.5								
		Gentamicin	0.3	0.8	[0.2 - 2.4]				54.6	41.8	2.5				0.3	0.3	0.5					
		Streptomycin	N/A	25.7	[21.3 - 30.5]													74.3	13.9	11.7		
	β-lactam / β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid	16.1	4.4	[2.5 - 7.0]						73.2	0.3	1.1	4.9	16.1	0.5	3.8					
		Cephems	0.0	4.9	[2.9 - 7.7]			0.3	0.3	26.0	68.0	0.5			0.3	4.6						
		Ceftriaxone	0.0	4.9	[2.9 - 7.7]					94.8	0.3				0.5	1.6	1.6	0.5	0.5			
	Penicillins	Ampicillin	0.0	26.2	[21.8 - 31.1]							69.7	4.1				0.5	25.7				
	Quinolones	Ciprofloxacin	0.0	0.0	[0.0 - 1.0]	97.5	0.5		0.5	0.3	1.1											
		Nalidixic acid	N/A	1.4	[0.4 - 3.2]							0.3	42.3	54.9	0.5	0.5	0.3	1.1				
		Tetracyclines	Tetracycline	0.3	29.0	[24.4 - 33.9]									70.8	0.3	1.9	13.4	13.7			
II	Aminoglycosides	Kanamycin	0.0	7.4	[4.9 - 10.6]											92.6					7.4	
	Cephems	Cefoxitin	0.3	3.6	[1.9 - 6.0]						21.9	67.5	6.6	0.3	0.3	1.1	2.5					
	Folate pathway inhibitors	Sulfisoxazole	N/A	28.7	[24.1 - 33.6]											1.4	54.9	15.0				
		Trimethoprim-sulfamethoxazole	N/A	1.9	[0.8 - 3.9]					94.0	3.6	0.5				1.9						28.7
	Phenicols	Chloramphenicol	0.8	20.2	[16.2 - 24.7]											29.2	49.7	0.8				20.2

* Rank of antimicrobial agents is based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important

† CLSI: Clinical and Laboratory Standards Institute

‡ Percent of isolates with intermediate susceptibility; N/A indicates that no MIC range of intermediate susceptibility exists

§ Percent of isolates that were resistant

¶ The 95% confidence intervals (CI) for percent resistant (%R) were calculated using the Paulson-Camp-Pratt approximation to the Clopper-Pearson exact method

** The unshaded areas indicate the dilution range of the Sensititre plates used to test isolates. Single vertical bars indicate the breakpoints for susceptibility, while double vertical bars indicate breakpoints for resistance. Numbers in the shaded areas indicate the percentages of isolates with MICs greater than the highest concentrations on the Sensititre plate. Numbers listed for the low est tested concentrations represent the percentages of isolates with MICs equal to or less than the low est tested concentration. CLSI breakpoints were used when available.

Figure 15. Antimicrobial resistance pattern for *Salmonella ser. Typhimurium*, 2010

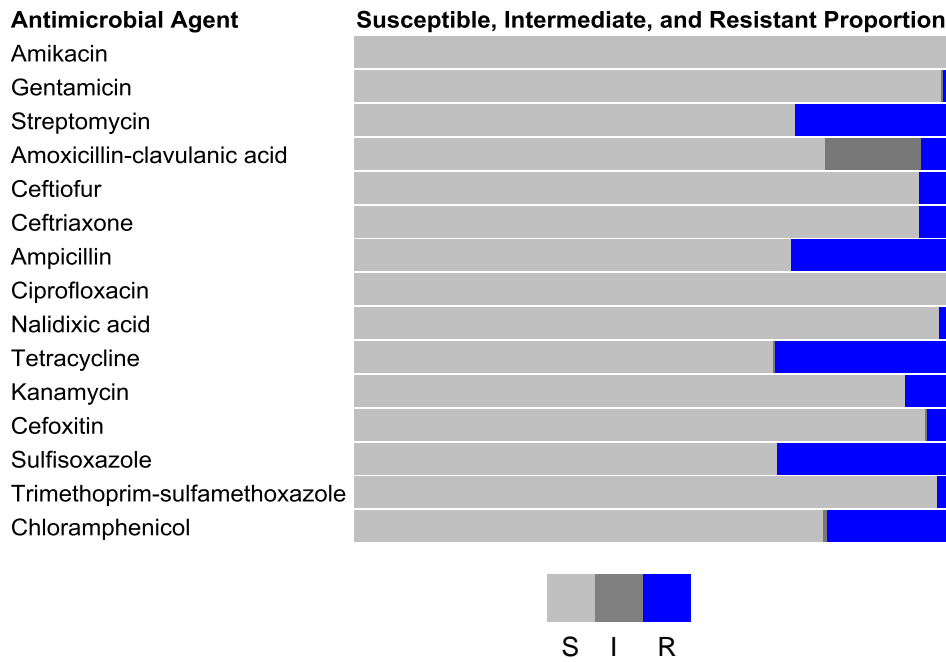


Table 14. Percentage and number of *Salmonella ser. Typhimurium* isolates resistant to antimicrobial agents, 2001–2010

Year			2001	2002	2003	2004	2005	2006	2007	2008	2009	2010
Total Isolates			325	394	408	383	438	408	405	397	371	366
Rank*	CLSI† Antimicrobial Class	Antimicrobial Agent (Resistance breakpoint)										
I	Aminoglycosides	Amikacin (MIC ≥ 64)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
		Gentamicin (MIC ≥ 16)	1.5%	2.3%	2.0%	2.1%	1.8%	2.7%	2.5%	1.5%	1.9%	0.8%
		Streptomycin (MIC ≥ 64)	40.0%	32.0%	35.5%	31.9%	28.1%	29.4%	32.3%	28.5%	25.9%	25.7%
	β-lactam/β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid (MIC ≥ 32/16)	6.2%	7.6%	5.6%	4.7%	3.2%	4.4%	6.7%	3.3%	6.2%	4.4%
		Cephems										
	Cephems	Cefotiofur (MIC ≥ 8)	3.1%	4.3%	4.9%	4.4%	2.5%	4.2%	6.4%	3.3%	6.5%	4.9%
		Ceftriaxone (MIC ≥ 4)	3.1%	4.3%	4.9%	4.4%	2.5%	4.2%	6.4%	3.3%	6.5%	4.9%
	Penicillins	Ampicillin (MIC ≥ 32)	42.5%	33.8%	36.3%	32.1%	29.0%	28.2%	31.6%	26.2%	28.0%	26.2%
		Quinolones										
	Quinolones	Ciprofloxacin (MIC ≥ 4)	0.3%	0.0%	0.0%	0.0%	0.0%	0.2%	0.0%	0.0%	0.0%	0.0%
Nalidixic Acid (MIC ≥ 32)		0.6%	1.3%	1.2%	0.5%	0.9%	0.7%	1.5%	1.3%	2.2%	1.4%	
Tetracyclines	Tetracycline (MIC ≥ 16)	43.4%	32.0%	38.2%	30.3%	30.4%	31.6%	36.8%	27.5%	28.8%	29.0%	
	II	Aminoglycosides	Kanamycin (MIC ≥ 64)	8.3%	7.6%	7.1%	5.7%	5.7%	5.1%	5.9%	2.3%	4.9%
Cephems												
Cephems		Cefoxitin (MIC ≥ 32)	3.1%	4.3%	4.4%	4.7%	2.5%	3.9%	5.7%	3.3%	5.4%	3.6%
		Cephalothin (MIC ≥ 32)	3.1%	5.6%	6.1%	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested
Folate pathway inhibitors		Sulfamethoxazole/Sulfisoxazole‡ (MIC ≥ 512)	43.1%	32.2%	38.7%	36.0%	32.0%	33.3%	37.3%	30.2%	29.9%	28.7%
		Trimethoprim-sulfamethoxazole (MIC ≥ 4/76)	2.5%	2.3%	3.4%	2.6%	2.7%	2.2%	2.5%	1.8%	3.0%	1.9%
Phenicol		Chloramphenicol (MIC ≥ 32)	31.7%	23.4%	28.2%	24.3%	24.4%	22.1%	25.4%	23.2%	20.5%	20.2%

* Rank of antimicrobial agents is based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important

† CLSI: Clinical and Laboratory Standards Institute

‡ Sulfamethoxazole, which was tested during 1996-2003 to represent sulfonamides, was replaced by sulfisoxazole in 2004

Table 15. Resistance patterns of *Salmonella ser. Typhimurium* isolates, 2001–2010

Year	2001	2002	2003	2004	2005	2006	2007	2008	2009	2010
Total Isolates	325	394	408	383	438	408	405	397	371	366
Resistance Pattern										
No resistance detected	49.2% 160	59.9% 236	54.7% 223	60.6% 232	65.1% 285	62.5% 255	57.5% 233	68.0% 270	63.6% 236	66.9% 245
Resistance ≥ 1 CLSI class*	50.8% 165	40.1% 158	45.3% 185	39.4% 151	34.9% 153	37.5% 153	42.5% 172	32.0% 127	36.4% 135	33.1% 121
Resistance ≥ 2 CLSI classes*	47.4% 154	36.3% 143	41.4% 169	37.1% 142	33.3% 146	34.1% 139	39.3% 159	31.2% 124	33.2% 123	30.3% 111
Resistance ≥ 3 CLSI classes*	41.5% 135	32.5% 128	37.3% 152	31.6% 121	30.1% 132	30.4% 124	34.3% 139	27.7% 110	28.0% 104	27.3% 100
Resistance ≥ 4 CLSI classes*	37.8% 123	28.4% 112	32.4% 132	27.7% 106	27.4% 120	27.0% 110	29.9% 121	24.7% 98	24.0% 89	24.3% 89
Resistance ≥ 5 CLSI classes*	29.5% 96	23.1% 91	27.7% 113	24.3% 93	22.8% 100	20.8% 85	24.9% 101	23.7% 94	22.1% 82	20.8% 76
At least ACSSuT†	29.5% 96	21.6% 85	26.5% 108	23.5% 90	22.4% 98	19.6% 80	22.7% 92	22.9% 91	19.4% 72	18.6% 68
At least ACT/S‡	0.9% 3	2.0% 8	3.2% 13	1.6% 6	2.1% 9	0.7% 3	2.0% 8	0.5% 2	2.2% 8	1.1% 4
At least ACSSuTAuCx§	1.2% 4	1.8% 7	2.2% 9	2.6% 10	1.8% 8	2.9% 12	3.7% 15	2.0% 8	1.6% 6	1.9% 7
At least ceftriaxone and nalidixic acid resistant	0.3% 1	0.5% 2	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.2% 1	0.0% 0	0.5% 2	0.3% 1

* CLSI: Clinical and Laboratory Standards Institute

† ACSSuT: resistance to ampicillin, chloramphenicol, streptomycin, sulfamethoxazole/sulfisoxazole, tetracycline

‡ ACT/S: resistance to ampicillin, chloramphenicol, trimethoprim-sulfamethoxazole

§ ACSSuTAuCx: resistance to ACSSuT, amoxicillin-clavulanic acid, ceftriaxone

C. *Salmonella ser. Newport*

Table 16. Minimum inhibitory concentrations (MICs) and resistance of *Salmonella ser. Newport* isolates to antimicrobial agents, 2010 (N=305)

Rank*	CLSI† Antimicrobial Class	Antimicrobial Agent	% of isolates			Percent of all isolates with MIC (µg/mL)**																	
			%‡	%R§	[95% CI]¶	0.015	0.03	0.06	0.125	0.25	0.50	1	2	4	8	16	32	64	128	256	512		
I	Aminoglycosides	Amikacin	0.0	0.0	[0.0 - 1.2]						1.3	78.0	19.3	1.3									
		Gentamicin	0.0	0.3	[0.0 - 1.8]				69.8	28.5	1.3							0.3					
		Streptomycin	N/A	8.2	[5.4 - 11.9]													91.8	0.7	7.5			
	β-lactam / β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid	0.0	7.5	[4.8 - 11.1]						90.5	1.3	0.3	0.3			2.6	4.9					
		Ceftiofur	0.0	7.2	[4.6 - 10.7]			0.3		29.8	62.0	0.7					7.2						
	Cephems	Ceftriaxone	0.0	7.2	[4.6 - 10.7]				92.8								2.3	3.6	1.0	0.3			
		Ampicillin	0.3	7.5	[4.8 - 11.1]						89.8	2.0	0.3				0.3			7.5			
	Quinolones	Ciprofloxacin	0.0	0.0	[0.0 - 1.2]	98.0	1.0		0.7	0.3													
		Nalidixic acid	N/A	0.3	[0.0 - 1.8]						0.3	37.4	61.0	0.7	0.3					0.3			
	Tetracyclines	Tetracycline	0.3	8.2	[5.4 - 11.9]									91.5	0.3		0.3	0.3	7.5				
II	Aminoglycosides	Kanamycin	0.0	0.7	[0.1 - 2.3]											99.0	0.3					0.7	
	Cephems	Cefoxitin	0.0	7.2	[4.6 - 10.7]						20.7	67.9	3.9	0.3			2.3	4.9					
	Folate pathway inhibitors	Sulfisoxazole	N/A	7.5	[4.8 - 11.1]												0.7	14.4	73.4	3.3	0.7	7.5	
		Trimethoprim-sulfamethoxazole	N/A	1.3	[0.4 - 3.3]				98.0	0.7						1.3							
Phenolics	Chloramphenicol	0.3	7.2	[4.6 - 10.7]										0.3	62.3	29.8	0.3				7.2		

* Rank of antimicrobial agents is based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important

† CLSI: Clinical and Laboratory Standards Institute

‡ Percent of isolates with intermediate susceptibility; N/A indicates that no MIC range of intermediate susceptibility exists

§ Percent of isolates that were resistant

¶ The 95% confidence intervals (CI) for percent resistant (%R) were calculated using the Paulson-Camp-Pratt approximation to the Clopper-Pearson exact method

** The unshaded areas indicate the dilution range of the Sensititre plates used to test isolates. Single vertical bars indicate the breakpoints for susceptibility, while double vertical bars indicate breakpoints for resistance. Numbers in the shaded areas indicate the percentages of isolates with MICs greater than the highest concentrations on the Sensititre plate. Numbers listed for the low end tested concentrations represent the percentages of isolates with MICs equal to or less than the low end tested concentration. CLSI breakpoints were used when available.

Figure 16. Antimicrobial resistance pattern for *Salmonella ser. Newport*, 2010

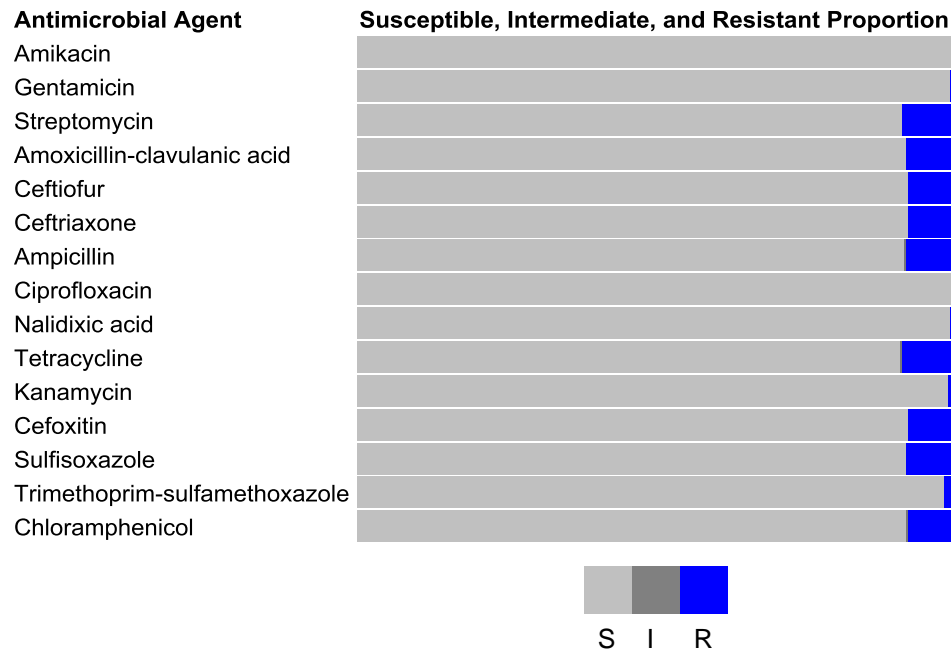


Table 17. Percentage and number of *Salmonella ser. Newport* isolates resistant to antimicrobial agents, 2001–2010

Year		2001	2002	2003	2004	2005	2006	2007	2008	2009	2010	
Total Isolates		124	241	223	191	207	217	221	255	236	305	
Rank	CLSI† Antimicrobial Class	Antimicrobial Agent (Resistance breakpoint)										
I	Aminoglycosides	Amikacin (MIC ≥ 64)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
		Gentamicin (MIC ≥ 16)	3.2%	3.3%	3.1%	0.5%	1.0%	0.9%	0.9%	0.4%	0.4%	0.3%
		Streptomycin (MIC ≥ 64)	31.5%	25.3%	24.2%	15.7%	14.0%	13.8%	10.4%	13.7%	7.6%	8.2%
	β-lactam/β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid (MIC ≥ 32/16)	26.6%	22.8%	21.5%	15.2%	12.6%	12.4%	8.1%	12.5%	6.8%	7.5%
		Cephems	Ceftiofur (MIC ≥ 8)	27.4%	22.8%	22.0%	15.2%	12.6%	12.4%	8.1%	12.5%	6.4%
	Cephems	Ceftriaxone (MIC ≥ 4)	25.8%	22.8%	21.5%	14.7%	12.6%	12.9%	8.1%	12.5%	6.4%	7.2%
		Penicillins	Ampicillin (MIC ≥ 32)	29.8%	24.9%	22.9%	15.7%	14.0%	15.2%	10.0%	14.5%	7.6%
	Quinolones	Ciprofloxacin (MIC ≥ 4)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
		Nalidixic Acid (MIC ≥ 32)	0.0%	0.8%	0.4%	0.5%	0.0%	0.5%	0.0%	0.4%	0.0%	0.3%
	Tetracyclines	Tetracycline (MIC ≥ 16)	30.6%	25.7%	24.2%	16.8%	14.5%	14.3%	10.0%	14.1%	8.1%	8.2%
II	Aminoglycosides	Kanamycin (MIC ≥ 64)	7.3%	10.0%	4.5%	2.6%	1.9%	2.3%	0.9%	3.5%	1.3%	0.7%
		Cephems	Cefoxitin (MIC ≥ 32)	25.8%	22.4%	21.5%	15.2%	12.6%	12.9%	8.1%	12.5%	5.9%
	Folate pathway inhibitors	Cephalothin (MIC ≥ 32)	26.6%	22.8%	22.4%	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested
		Sulfamethoxazole/Sulfisoxazole‡ (MIC ≥ 512)	32.3%	25.7%	24.7%	16.8%	15.5%	15.2%	10.4%	13.3%	8.1%	7.5%
	Phenicol	Trimethoprim-sulfamethoxazole (MIC ≥ 4/76)	1.6%	4.1%	0.9%	2.1%	1.9%	3.2%	1.8%	3.1%	0.4%	1.3%
		Chloramphenicol (MIC ≥ 32)	28.2%	25.3%	22.4%	15.2%	13.5%	12.4%	9.5%	12.2%	6.8%	7.2%

* Rank of antimicrobial agents is based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important

† CLSI: Clinical and Laboratory Standards Institute

‡ Sulfamethoxazole, which was tested during 1996-2003 to represent sulfonamides, was replaced by sulfisoxazole in 2004

Table 18. Resistance patterns of *Salmonella ser. Newport* isolates, 2001–2010

Year	2001	2002	2003	2004	2005	2006	2007	2008	2009	2010
Total Isolates	124	241	223	191	207	217	221	255	236	305
Resistance Pattern										
No resistance detected	65.3% 81	72.2% 174	73.5% 164	82.2% 157	84.1% 174	82.9% 180	89.1% 197	85.1% 217	89.8% 212	90.8% 277
Resistance ≥ 1 CLSI class*	34.7% 43	27.8% 67	26.5% 59	17.8% 34	15.9% 33	17.1% 37	10.9% 24	14.9% 38	10.2% 24	9.2% 28
Resistance ≥ 2 CLSI classes*	32.3% 40	25.3% 61	25.1% 56	17.3% 33	15.0% 31	16.6% 36	10.9% 24	13.7% 35	8.5% 20	7.9% 24
Resistance ≥ 3 CLSI classes*	31.5% 39	25.3% 61	23.3% 52	16.2% 31	14.5% 30	15.2% 33	10.9% 24	13.7% 35	7.6% 18	7.5% 23
Resistance ≥ 4 CLSI classes*	31.5% 39	25.3% 61	22.9% 51	15.7% 30	14.0% 29	13.4% 29	9.5% 21	13.7% 35	6.8% 16	7.5% 23
Resistance ≥ 5 CLSI classes*	26.6% 33	23.7% 57	22.4% 50	14.7% 28	12.6% 26	12.9% 28	8.6% 19	12.9% 33	6.4% 15	7.2% 22
At least ACSSuT†	25.8% 32	23.7% 57	22.0% 49	14.7% 28	12.6% 26	12.0% 26	8.6% 19	11.8% 30	6.4% 15	7.2% 22
At least ACT/S‡	0.8% 1	3.7% 9	0.9% 2	1.0% 2	1.9% 4	2.3% 5	0.5% 1	2.7% 7	0.4% 1	1.3% 4
At least ACSSuTAuCx§	25.0% 31	22.8% 55	21.1% 47	14.7% 28	12.6% 26	10.6% 23	8.1% 18	11.8% 30	6.4% 15	7.2% 22
At least ceftriaxone and nalidixic acid resistant	0.0% 0	0.4% 1	0.0% 0	0.5% 1	0.0% 0	0.5% 1	0.0% 0	0.0% 0	0.0% 0	0.0% 0

* CLSI: Clinical and Laboratory Standards Institute

† ACSSuT: resistance to ampicillin, chloramphenicol, streptomycin, sulfamethoxazole/sulfisoxazole, tetracycline

‡ ACT/S: resistance to ampicillin, chloramphenicol, trimethoprim-sulfamethoxazole

§ ACSSuTAuCx: resistance to ACSSuT, amoxicillin-clavulanic acid, ceftriaxone

D. *Salmonella ser. Heidelberg*

Table 19. Minimum inhibitory concentrations (MICs) and resistance of *Salmonella ser. Heidelberg* isolates to antimicrobial agents, 2010 (N=62)

Rank*	CLSI† Antimicrobial Class	Antimicrobial Agent	% of isolates			Percent of all isolates with MIC (µg/mL)**																	
			%‡	%R§	[95% CI]¶	0.015	0.03	0.06	0.125	0.25	0.50	1	2	4	8	16	32	64	128	256	512		
I	Aminoglycosides	Amikacin	0.0	0.0	[0.0 - 5.8]						6.5	64.5	24.2	4.8									
		Gentamicin	0.0	8.1	[2.7 - 17.8]					62.9	24.2	4.8					4.8	3.2					
		Streptomycin	N/A	27.4	[16.8 - 40.2]														72.6	12.9	14.5		
	β-lactam / β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid	0.0	24.2	[14.2 - 36.7]						61.3		1.6	12.9				4.8	19.4				
		Cephems	0.0	24.2	[14.2 - 36.7]					1.6	32.3	41.9						24.2					
		Ceftriaxone	0.0	24.2	[14.2 - 36.7]					75.8								16.1	6.5	1.6			
	Penicillins	Ampicillin	0.0	38.7	[26.6 - 51.9]							59.7	1.6								38.7		
	Quinolones	Ciprofloxacin	0.0	0.0	[0.0 - 5.8]	100.0																	
		Nalidixic acid	N/A	0.0	[0.0 - 5.8]								19.4	80.6									
		Tetracyclines	Tetracycline	0.0	24.2	[14.2 - 36.7]												75.8			1.6	22.6	
II	Aminoglycosides	Kanamycin	0.0	22.6	[12.9 - 35.0]																1.6	21.0	
	Cephems	Cefoxitin	0.0	24.2	[14.2 - 36.7]																16.1	8.1	
	Folate pathway inhibitors	Sulfisoxazole	N/A	11.3	[4.6 - 21.9]																19.4	53.2	
		Trimethoprim-sulfamethoxazole	N/A	0.0	[0.0 - 5.8]																	16.1	11.3
	Phenolics	Chloramphenicol	0.0	1.6	[0.0 - 8.7]																		1.6

* Rank of antimicrobial agents is based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important

† CLSI Clinical and Laboratory Standards Institute

‡ Percent of isolates with intermediate susceptibility; N/A indicates that no MIC range of intermediate susceptibility exists

§ Percent of isolates that were resistant

¶ The 95% confidence intervals (CI) for percent resistant (%R) were calculated using the Paulson-Camp-Pratt approximation to the Clopper-Pearson exact method

** The unshaded areas indicate the dilution range of the Sensititre plates used to test isolates. Single vertical bars indicate the breakpoints for susceptibility, while double vertical bars indicate breakpoints for resistance. Numbers in the shaded areas indicate the percentages of isolates with MICs greater than the highest concentrations on the Sensititre plate. Numbers listed for the lowest tested concentrations represent the percentages of isolates with MICs equal to or less than the lowest tested concentration. CLSI breakpoints were used when available.

Figure 17. Antimicrobial resistance pattern for *Salmonella ser. Heidelberg*, 2010

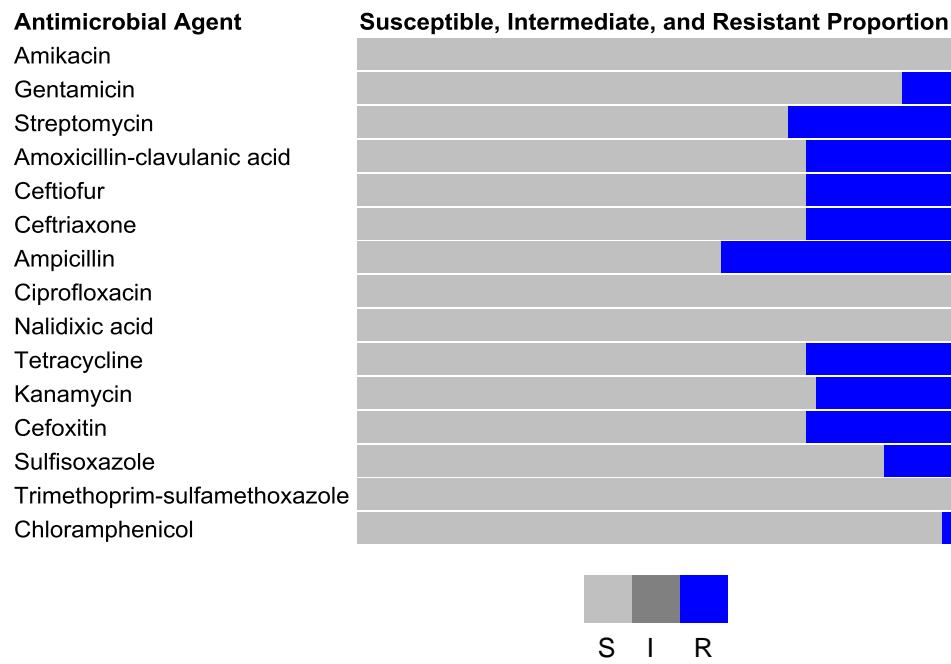


Table 20. Percentage and number of *Salmonella ser. Heidelberg* isolates resistant to antimicrobial agents, 2001–2010

Year	2001	2002	2003	2004	2005	2006	2007	2008	2009	2010	
Total Isolates	102	105	96	92	125	102	98	75	86	62	
Rank	CLSI [†] Antimicrobial Class	Antimicrobial Agent (Resistance breakpoint)									
I	Aminoglycosides	Amikacin (MIC ≥ 64)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	
		Gentamicin (MIC ≥ 16)	7.8%	3.8%	5.2%	4.3%	6.4%	4.9%	16.3%	14.7%	2.3%
		Streptomycin (MIC ≥ 64)	25.5%	17.1%	12.5%	15.2%	13.6%	11.8%	12.2%	30.7%	23.3%
	β-lactam/β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid (MIC ≥ 32/16)	2.9%	9.5%	5.2%	9.8%	8.8%	9.8%	7.1%	8.0%	20.9%
		Cephems	Ceftiofur (MIC ≥ 8)	2.9%	7.6%	5.2%	8.7%	8.8%	9.8%	7.1%	8.0%
	Cephems	Ceftriaxone (MIC ≥ 4)	3	8	5	8	11	10	7	6	18
		Penicillins	Ampicillin (MIC ≥ 32)	9.8%	12.4%	10.4%	25.0%	20.0%	18.6%	18.4%	28.0%
	Quinolones	Ciprofloxacin (MIC ≥ 4)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
		Nalidixic Acid (MIC ≥ 32)	0.0%	0.0%	1.0%	0.0%	0.8%	0.0%	0.0%	0.0%	0.0%
	Tetracyclines	Tetracycline (MIC ≥ 16)	24.5%	19.0%	16.7%	19.6%	18.4%	13.7%	22.4%	36.0%	27.9%
II	Aminoglycosides	Kanamycin (MIC ≥ 64)	19.6%	10.5%	8.3%	8.7%	12.8%	8.8%	11.2%	26.7%	20.9%
		Cephems	Cefoxitin (MIC ≥ 32)	2.9%	8.6%	5.2%	7.6%	8.8%	8.8%	7.1%	8.0%
	Cephems	Cephalothin (MIC ≥ 32)	3.9%	10.5%	7.3%	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested
		Folate pathway inhibitors	Sulfamethoxazole/Sulfisoxazole [‡] (MIC ≥ 512)	8.8%	6.7%	7.3%	7.6%	8.0%	4.9%	18.4%	12.0%
	Trimethoprim-sulfamethoxazole (MIC ≥ 4/76)		2.0%	1.0%	2.1%	0.0%	0.8%	0.0%	0.0%	2.7%	
	Phenicol	Chloramphenicol (MIC ≥ 32)	1.0%	1.0%	0.0%	1.1%	0.8%	0.0%	3.1%	1.3%	
			1	1	0	1	1	0	3	1	

* Rank of antimicrobial agents is based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important

[†] CLSI: Clinical and Laboratory Standards Institute

[‡] Sulfamethoxazole, which was tested during 1996-2003 to represent sulfonamides, was replaced by sulfisoxazole in 2004

Table 21. Resistance patterns of *Salmonella ser. Heidelberg* isolates, 2001–2010

Year	2001	2002	2003	2004	2005	2006	2007	2008	2009	2010
Total Isolates	102	105	96	92	125	102	98	75	86	62
Resistance Pattern										
No resistance detected	64.7% 66	67.6% 71	68.8% 66	56.5% 52	62.4% 78	67.6% 69	58.2% 57	57.3% 43	60.5% 52	51.6% 32
Resistance ≥ 1 CLSI class*	35.3% 36	32.4% 34	31.3% 30	43.5% 40	37.6% 47	32.4% 33	41.8% 41	42.7% 32	39.5% 34	48.4% 30
Resistance ≥ 2 CLSI classes*	28.4% 29	25.7% 27	17.7% 17	22.8% 21	24.8% 31	23.5% 24	28.6% 28	40.0% 30	34.9% 30	43.5% 27
Resistance ≥ 3 CLSI classes*	7.8% 8	12.4% 13	10.4% 10	13.0% 12	15.2% 19	12.7% 13	17.3% 17	28.0% 21	25.6% 22	33.9% 21
Resistance ≥ 4 CLSI classes*	2.0% 2	1.9% 2	0.0% 0	4.3% 4	4.8% 6	2.0% 2	5.1% 5	13.3% 10	17.4% 15	11.3% 7
Resistance ≥ 5 CLSI classes*	1.0% 1	1.9% 2	0.0% 0	3.3% 3	1.6% 2	2.0% 2	4.1% 4	6.7% 5	15.1% 13	9.7% 6
At least ACSSuT†	1.0% 1	1.0% 1	0.0% 0	1.1% 1	0.0% 0	0.0% 0	3.1% 3	1.3% 1	3.5% 3	1.6% 1
At least ACT/S‡	0.0% 0	1.0% 1	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	3.5% 3	0.0% 0
At least ACSSuTAuCx§	1.0% 1	1.0% 1	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	1.2% 1	0.0% 0
At least ceftriaxone and nalidixic acid resistant	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0

* CLSI: Clinical and Laboratory Standards Institute

† ACSSuT: resistance to ampicillin, chloramphenicol, streptomycin, sulfamethoxazole/sulfisoxazole, tetracycline

‡ ACT/S: resistance to ampicillin, chloramphenicol, trimethoprim-sulfamethoxazole

§ ACSSuTAuCx: resistance to ACSSuT, amoxicillin-clavulanic acid, ceftriaxone

E. *Salmonella ser. I 4,[5],12:i:-*

Table 22. Minimum inhibitory concentrations (MICs) and resistance of *Salmonella ser. I 4,[5],12:i:-* isolates to antimicrobial agents, 2010 (N=77)

Rank*	CLSI† Antimicrobial Class	Antimicrobial Agent	% of isolates			Percent of all isolates with MIC (µg/mL)**												
			%‡	%R§	[95% CI]¶	0.015	0.03	0.06	0.125	0.25	0.50	1	2	4	8	16	32	64
I	Aminoglycosides	Amikacin	0.0	0.0	[0.0 - 4.7]	[Shaded area from 0.015 to 0.50, with values 77.9, 20.8, 1.3]												
		Gentamicin	0.0	1.3	[0.0 - 7.0]	[Shaded area from 0.015 to 0.50, with values 76.6, 20.8, 1.3]												
		Streptomycin	N/A	19.5	[11.3 - 30.1]	[Shaded area from 0.015 to 0.50, with values 80.5, 2.6, 16.9]												
	β-lactam / β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid	3.9	3.9	[0.8 - 11.0]	[Shaded area from 0.015 to 0.50, with values 77.9, 6.5, 7.8, 3.9, 1.3, 2.6]												
		Cephems	Ceftiofur	0.0	2.6	[0.3 - 9.1]	[Shaded area from 0.015 to 0.50, with values 29.9, 67.5, 2.6]											
	Ceftriaxone	0.0	2.6	[0.3 - 9.1]	[Shaded area from 0.015 to 0.50, with values 97.4, 2.6]													
	Penicillins	Ampicillin	0.0	22.1	[13.4 - 33.0]	[Shaded area from 0.015 to 0.50, with values 74.0, 3.9, 22.1]												
		Quinolones	Ciprofloxacin	0.0	1.3	[0.0 - 7.0]	[Shaded area from 0.015 to 0.50, with values 96.1, 1.3, 1.3]											
	Nalidixic acid	N/A	2.6	[0.3 - 9.1]	[Shaded area from 0.015 to 0.50, with values 46.8, 50.6, 2.6]													
	Tetracyclines	Tetracycline	0.0	28.6	[18.8 - 40.0]	[Shaded area from 0.015 to 0.50, with values 71.4, 1.3, 27.3]												
II	Aminoglycosides	Kanamycin	0.0	1.3	[0.0 - 7.0]	[Shaded area from 0.015 to 0.50, with values 98.7, 1.3, 1.3]												
		Cephems	Cefoxitin	1.3	2.6	[0.3 - 9.1]	[Shaded area from 0.015 to 0.50, with values 22.1, 63.6, 9.1, 1.3, 1.3, 2.6]											
	Folate pathway inhibitors	Sulfisoxazole	N/A	19.5	[11.3 - 30.1]	[Shaded area from 0.015 to 0.50, with values 1.3, 50.6, 27.3, 1.3, 19.5]												
		Trimethoprim-sulfamethoxazole	N/A	1.3	[0.0 - 7.0]	[Shaded area from 0.015 to 0.50, with values 97.4, 1.3, 1.3]												
	Phenicols	Chloramphenicol	1.3	1.3	[0.0 - 7.0]	[Shaded area from 0.015 to 0.50, with values 35.1, 62.3, 1.3, 1.3]												

* Rank of antimicrobial agents is based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important

† CLSI: Clinical and Laboratory Standards Institute

‡ Percent of isolates with intermediate susceptibility; N/A indicates that no MIC range of intermediate susceptibility exists

§ Percent of isolates that were resistant

¶ The 95% confidence intervals (CI) for percent resistant (%R) were calculated using the Paulson-Camp-Pratt approximation to the Clopper-Pearson exact method

** The unshaded areas indicate the dilution range of the Sensititre plates used to test isolates. Single vertical bars indicate the breakpoints for susceptibility, while double vertical bars indicate breakpoints for resistance. Numbers in the shaded areas indicate the percentages of isolates with MICs greater than the highest concentrations on the Sensititre plate. Numbers listed for the low est tested concentrations represent the percentages of isolates with MICs equal to or less than the low est tested concentration. CLSI breakpoints were used when available.

Figure 18. Antimicrobial resistance pattern for *Salmonella ser. I 4,[5],12:i:-*, 2010

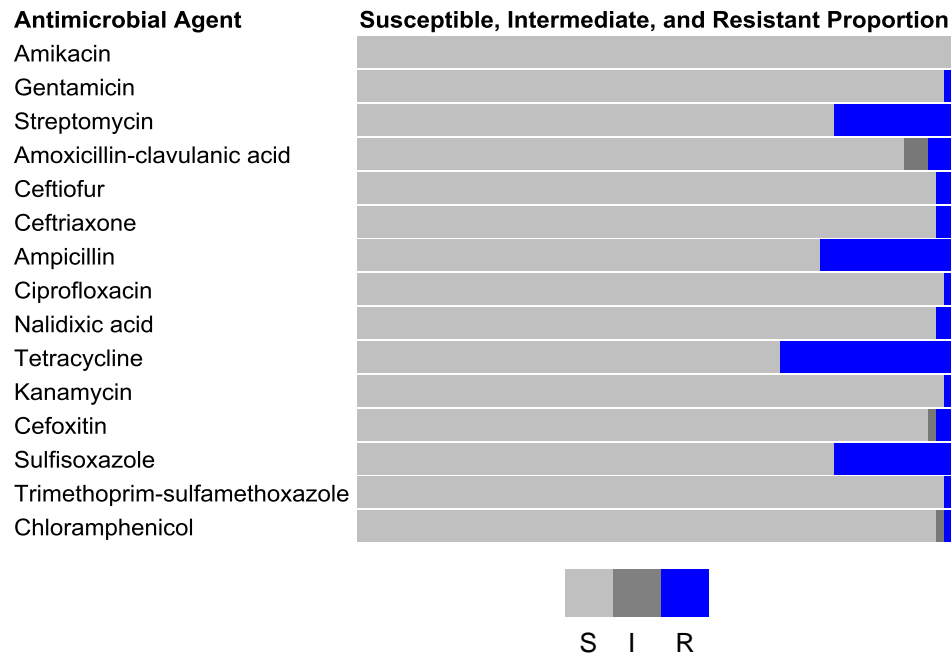


Table 23. Percentage and number of *Salmonella ser. I 4,[5],12:i:-* isolates resistant to antimicrobial agents, 2001–2010

Year			2001	2002	2003	2004	2005	2006	2007	2008	2009	2010	
Total Isolates			14	35	37	36	33	105	73	84	72	77	
Rank*	CLSI† Antimicrobial Class	Antimicrobial Agent (Resistance breakpoint)											
I	Aminoglycosides	Amikacin (MIC ≥ 64)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	
		Gentamicin (MIC ≥ 16)	7.1%	0.0%	5.4%	5.6%	0.0%	4.8%	1.4%	3.6%	2.8%	1.3%	
		Streptomycin (MIC ≥ 64)	14.3%	2.9%	8.1%	5.6%	3.0%	3.8%	8.2%	10.7%	12.5%	19.5%	
	β-lactam/β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid (MIC ≥ 32/16)	0.0%	2.9%	5.4%	2.8%	3.0%	3.8%	1.4%	4.8%	4.2%	3.9%	
		Cephems											
	Cephems	Ceftiofur (MIC ≥ 8)	7.1%	2.9%	5.4%	2.8%	3.0%	3.8%	2.7%	4.8%	2.8%	2.6%	
		Ceftriaxone (MIC ≥ 4)	0.0%	2.9%	5.4%	2.8%	3.0%	3.8%	2.7%	4.8%	2.8%	2.6%	
	Penicillins	Ampicillin (MIC ≥ 32)	7.1%	8.6%	8.1%	5.6%	6.1%	6.7%	5.5%	9.5%	11.1%	22.1%	
	Quinolones	Ciprofloxacin (MIC ≥ 4)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	1.3%
		Nalidixic Acid (MIC ≥ 32)	0.0%	0.0%	2.7%	2.8%	0.0%	1.0%	1.4%	1.2%	0.0%	2.6%	
Tetracyclines	Tetracycline (MIC ≥ 16)	7.1%	5.7%	0.0%	11.1%	3.0%	8.6%	9.6%	16.7%	16.7%	28.6%		
II	Aminoglycosides	Kanamycin (MIC ≥ 64)	7.1%	0.0%	0.0%	0.0%	0.0%	0.0%	1.4%	1.2%	0.0%	1.3%	
		Cephems											
	Cephems	Cefoxitin (MIC ≥ 32)	0.0%	2.9%	5.4%	2.8%	3.0%	3.8%	1.4%	4.8%	2.8%	2.6%	
		Cephalothin (MIC ≥ 32)	7.1%	2.9%	5.4%	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	
	Folate pathway inhibitors	Sulfamethoxazole/Sulfisoxazole‡ (MIC ≥ 512)	14.3%	2.9%	5.4%	11.1%	0.0%	8.6%	4.1%	13.1%	13.9%	19.5%	
		Trimethoprim-sulfamethoxazole (MIC ≥ 4/76)	7.1%	2.9%	0.0%	2.8%	0.0%	0.0%	1.4%	4.8%	1.4%	1.3%	
	Phenicol	Chloramphenicol (MIC ≥ 32)	7.1%	2.9%	0.0%	2.8%	0.0%	1.9%	1.4%	6.0%	8.3%	1.3%	

* Rank of antimicrobial agents is based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important

† CLSI: Clinical and Laboratory Standards Institute

‡ Sulfamethoxazole, which was tested during 1996-2003 to represent sulfonamides, was replaced by sulfisoxazole in 2004

Table 24. Resistance patterns of *Salmonella ser. I 4,[5],12:i-* isolates, 2001–2010

Year	2001	2002	2003	2004	2005	2006	2007	2008	2009	2010
Total Isolates	14	35	37	36	33	105	73	84	72	77
Resistance Pattern										
No resistance detected	78.6% 11	91.4% 32	78.4% 29	80.6% 29	87.9% 29	85.7% 90	82.2% 60	76.2% 64	76.4% 55	66.2% 51
Resistance ≥ 1 CLSI class*	21.4% 3	8.6% 3	21.6% 8	19.4% 7	12.1% 4	14.3% 15	17.8% 13	23.8% 20	23.6% 17	33.8% 26
Resistance ≥ 2 CLSI classes*	14.3% 2	8.6% 3	10.8% 4	13.9% 5	3.0% 1	11.4% 12	6.8% 5	17.9% 15	16.7% 12	22.1% 17
Resistance ≥ 3 CLSI classes*	7.1% 1	5.7% 2	5.4% 2	8.3% 3	3.0% 1	9.5% 10	5.5% 4	10.7% 9	12.5% 9	22.1% 17
Resistance ≥ 4 CLSI classes*	7.1% 1	2.9% 1	0.0% 0	2.8% 1	0.0% 0	3.8% 4	2.7% 2	7.1% 6	9.7% 7	19.5% 15
Resistance ≥ 5 CLSI classes*	7.1% 1	2.9% 1	0.0% 0	2.8% 1	0.0% 0	2.9% 3	1.4% 1	4.8% 4	6.9% 5	3.9% 3
At least ACSSuT [†]	7.1% 1	2.9% 1	0.0% 0	2.8% 1	0.0% 0	1.9% 2	1.4% 1	3.6% 3	6.9% 5	1.3% 1
At least ACT/S [‡]	7.1% 1	2.9% 1	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0
At least ACSSuTAuCx [§]	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	2.4% 2	0.0% 0	0.0% 0
At least ceftriaxone and nalidixic acid resistant	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0

* CLSI: Clinical and Laboratory Standards Institute

† ACSSuT: resistance to ampicillin, chloramphenicol, streptomycin, sulfamethoxazole/sulfisoxazole, tetracycline

‡ ACT/S: resistance to ampicillin, chloramphenicol, trimethoprim-sulfamethoxazole

§ ACSSuTAuCx: resistance to ACSSuT, amoxicillin-clavulanic acid, ceftriaxone

2. Typhoidal *Salmonella*

A. *Salmonella ser. Typhi*

Table 25. Minimum inhibitory concentrations (MICs) and resistance of *Salmonella ser. Typhi* isolates to antimicrobial agents, 2010 (N=444)

Rank [*]	CLSI [†] Antimicrobial Class	Antimicrobial Agent	% of isolates			Percent of all isolates with MIC (µg/mL) ^{**}																	
			% [‡]	%R [§]	[95% CI] [¶]	0.015	0.03	0.06	0.125	0.25	0.50	1	2	4	8	16	32	64	128	256	512		
I	Aminoglycosides	Amikacin	0.0	0.0	[0.0 - 0.8]						26.4	66.2	7.2	0.2									
		Gentamicin	0.0	0.0	[0.0 - 0.8]				92.3	7.4	0.2												
		Streptomycin	N/A	10.1	[7.5 - 13.3]												89.9						10.1
	β-lactam / β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid	0.2	0.0	[0.0 - 0.8]							87.6	0.2	2.5	9.5	0.2							
		Cephems																					
		Ceftiofur	0.0	0.0	[0.0 - 0.8]				1.4	11.3	79.1	8.1	0.2										
		Ceftriaxone	0.0	0.0	[0.0 - 0.8]					100.0													
		Penicillins																					
		Ampicillin	0.0	12.4	[9.5 - 15.8]							87.4	0.2						0.2				12.2
		Quinolones																					
	Ciprofloxacin	1.1	2.7	[1.4 - 4.7]	28.2	0.2	2.7	14.0	46.2	4.5	0.5	1.1				2.7							
	Nalidixic acid	N/A	69.1	[64.6 - 73.4]									2.9	24.3	2.5	1.1			1.4			67.8	
	Tetracyclines																						
	Tetracycline	0.0	3.6	[2.1 - 5.8]											96.4							3.6	
II	Aminoglycosides	Kanamycin	0.0	0.2	[0.0 - 1.2]											99.8						0.2	
	Cephems	Cefoxitin	0.2	0.0	[0.0 - 0.8]																		
	Folate pathway inhibitors	Sulfisoxazole	N/A	12.4	[9.5 - 15.8]																		
		Trimethoprim-sulfamethoxazole	N/A	11.9	[9.1 - 15.3]																		
	Phenicol																						
	Chloramphenicol	0.0	11.7	[8.9 - 15.1]																			

* Rank of antimicrobial agents is based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important

† CLSI: Clinical and Laboratory Standards Institute

‡ Percent of isolates with intermediate susceptibility; N/A indicates that no MIC range of intermediate susceptibility exists

§ Percent of isolates that were resistant

¶ The 95% confidence intervals (CI) for percent resistant (%R) were calculated using the Paulson-Camp-Pratt approximation to the Clopper-Pearson exact method

** The unshaded areas indicate the dilution range of the Sensititre plates used to test isolates. Single vertical bars indicate the breakpoints for susceptibility, while double vertical bars indicate breakpoints for resistance. Numbers in the shaded areas indicate the percentages of isolates with MICs greater than the highest concentrations on the Sensititre plate. Numbers listed for the low est tested concentrations represent the percentages of isolates with MICs equal to or less than the low est tested concentration. CLSI breakpoints were used when available.

Figure 19. Antimicrobial resistance pattern for *Salmonella ser. Typhi*, 2010

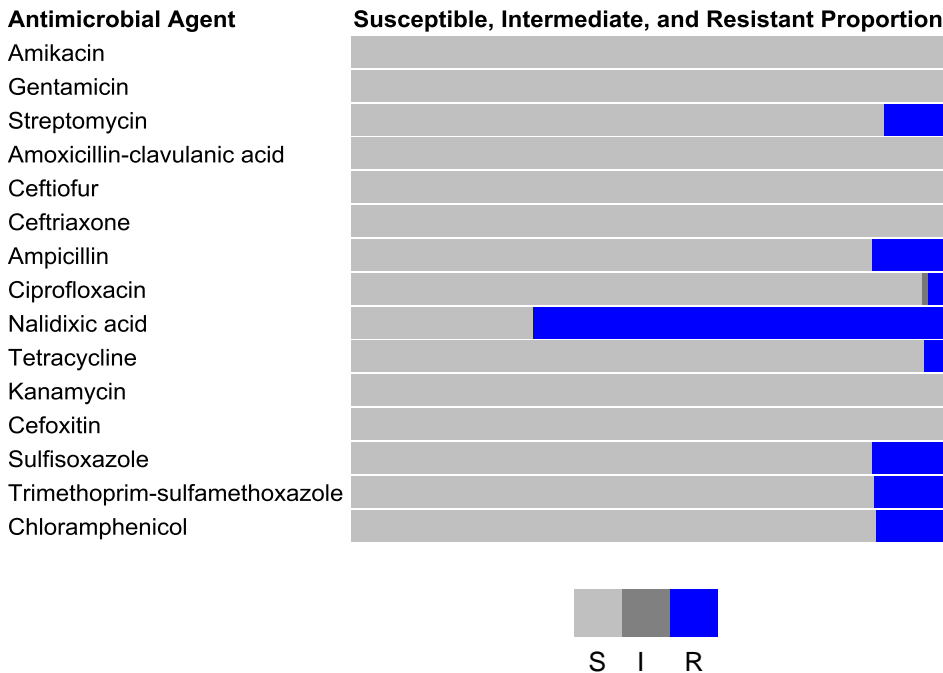


Table 26. Percentage and number of *Salmonella ser. Typhi* isolates resistant to antimicrobial agents, 2001–2010

Year		2001	2002	2003	2004	2005	2006	2007	2008	2009	2010	
Total Isolates		197	195	332	304	318	323	400	408	362	444	
Rank	CLSI [†] Antimicrobial Class	Antimicrobial Agent (Resistance breakpoint)										
I	Aminoglycosides	Amikacin (MIC ≥ 64)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
		Gentamicin (MIC ≥ 16)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
		Streptomycin (MIC ≥ 64)	20.3%	7.2%	14.5%	11.8%	13.2%	18.9%	15.8%	11.5%	10.8%	10.1%
	β-lactam/β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid (MIC ≥ 32/16)	0.0%	0.0%	0.0%	0.0%	0.0%	0.3%	0.3%	0.0%	0.3%	0.0%
		Cephems	Ceftiofur (MIC ≥ 8)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
	Penicillins	Ceftriaxone (MIC ≥ 4)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
		Ampicillin (MIC ≥ 32)	20.3%	5.6%	16.0%	11.8%	13.2%	20.4%	17.0%	13.2%	12.4%	12.4%
	Quinolones	Ciprofloxacin (MIC ≥ 4)	0.0%	0.0%	0.3%	0.0%	0.3%	0.9%	1.0%	0.0%	3.3%	2.7%
		Nalidixic Acid (MIC ≥ 32)	29.9%	23.6%	37.7%	41.8%	48.4%	54.5%	62.0%	58.8%	59.9%	69.1%
	Tetracyclines	Tetracycline (MIC ≥ 16)	20.8%	6.7%	15.4%	8.9%	10.1%	8.4%	6.3%	4.7%	5.8%	3.6%
		41	13	51	27	32	27	25	19	21	16	
II	Aminoglycosides	Kanamycin (MIC ≥ 64)	0.5%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.2%
			1	0	0	0	0	0	0	0	0	1
	Cephems	Cefoxitin (MIC ≥ 32)	0.5%	0.0%	0.3%	0.0%	0.0%	0.3%	0.5%	0.0%	0.0%	0.0%
			1	0	1	0	0	1	2	0	0	0
	Folate pathway inhibitors	Cephalothin (MIC ≥ 32)	0.5%	1.5%	0.0%	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested
		Sulfamethoxazole/Sulfisoxazole [‡] (MIC ≥ 512)	20.8%	6.2%	16.9%	11.8%	14.2%	20.7%	17.5%	13.2%	13.5%	12.4%
		41	12	56	36	45	67	70	54	49	55	
Phenicol	Trimethoprim-sulfamethoxazole (MIC ≥ 4/76)	20.8%	6.7%	16.9%	13.2%	14.5%	20.7%	16.3%	12.7%	12.4%	11.9%	
		41	13	56	40	46	67	65	52	45	53	
	Chloramphenicol (MIC ≥ 32)	20.8%	6.2%	16.6%	13.2%	13.2%	19.5%	15.8%	13.0%	11.6%	11.7%	
		41	12	55	40	42	63	63	53	42	52	

* Rank of antimicrobial agents is based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important

† CLSI: Clinical and Laboratory Standards Institute

‡ Sulfamethoxazole, which was tested during 1996-2003 to represent sulfonamides, was replaced by sulfisoxazole in 2004

Table 27. Resistance patterns of *Salmonella ser. Typhi* isolates, 2001–2010

Year	2001	2002	2003	2004	2005	2006	2007	2008	2009	2010
Total Isolates	197	195	332	304	318	323	400	408	362	444
Resistance Pattern										
No resistance detected	58.9%	74.4%	56.6%	56.6%	48.1%	40.2%	35.5%	38.2%	37.6%	29.5%
	116	145	188	172	153	130	142	156	136	131
Resistance ≥ 1 CLSI class*	41.1%	25.6%	43.4%	43.4%	51.9%	59.8%	64.5%	61.8%	62.4%	70.5%
	81	50	144	132	165	193	258	252	226	313
Resistance ≥ 2 CLSI classes*	22.8%	7.2%	17.5%	13.2%	14.5%	21.7%	18.0%	14.5%	14.4%	13.7%
	45	14	58	40	46	70	72	59	52	61
Resistance ≥ 3 CLSI classes*	21.8%	6.7%	16.6%	12.8%	13.8%	20.7%	17.5%	13.5%	13.0%	13.7%
	43	13	55	39	44	67	70	55	47	61
Resistance ≥ 4 CLSI classes*	21.3%	6.2%	16.3%	12.5%	12.9%	19.2%	17.0%	13.0%	12.4%	11.7%
	42	12	54	38	41	62	68	53	45	52
Resistance ≥ 5 CLSI classes*	16.8%	5.6%	14.2%	11.8%	11.9%	16.7%	14.8%	10.8%	10.2%	9.7%
	33	11	47	36	38	54	59	44	37	43
At least ACSSuT [†]	16.8%	5.6%	12.7%	7.9%	9.1%	5.9%	3.8%	2.5%	2.8%	1.6%
	33	11	42	24	29	19	15	10	10	7
At least ACT/S [‡]	17.8%	5.6%	15.7%	11.8%	12.9%	18.6%	15.3%	12.3%	10.8%	10.6%
	35	11	52	36	41	60	61	50	39	47
At least ACSSuTAuCx [§]	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
	0	0	0	0	0	0	0	0	0	0
At least ceftriaxone and nalidixic acid resistant	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
	0	0	0	0	0	0	0	0	0	0

* CLSI: Clinical and Laboratory Standards Institute

† ACSSuT: resistance to ampicillin, chloramphenicol, streptomycin, sulfamethoxazole/sulfisoxazole, tetracycline

‡ ACT/S: resistance to ampicillin, chloramphenicol, trimethoprim-sulfamethoxazole

§ ACSSuTAuCx: resistance to ACSSuT, amoxicillin-clavulanic acid, ceftriaxone

B. Salmonella ser. Paratyphi A, Paratyphi B, and Paratyphi C

Table 28. Frequency of Salmonella ser. Paratyphi A, Paratyphi B, and Paratyphi C, 2010

Species	2010	
	n	(%)
Paratyphi A	143	(97.9)
Paratyphi B	3	(2.1)
Paratyphi C	0	(0)
Total	146	(100)

Table 29. Minimum inhibitory concentrations (MICs) and resistance of Salmonella ser. Paratyphi A, Paratyphi B, and Paratyphi C isolates to antimicrobial agents, 2010 (N=146)

Rank	CLSI [†] Antimicrobial Class	Antimicrobial Agent	% of isolates			Percent of all isolates with MIC (µg/mL) ^{**}													
			% [‡]	%R [§]	[95% CI] [¶]	0.015	0.03	0.06	0.125	0.25	0.50	1	2	4	8	16	32	64	128
I	Aminoglycosides	Amikacin	0.0	0.0	[0.0 - 2.5]	[Shaded area from 0.015 to 0.50]													
		Gentamicin	0.0	0.7	[0.0 - 3.8]	[Shaded area from 0.015 to 0.25]													
		Streptomycin	N/A	2.1	[0.4 - 5.9]	[Shaded area from 0.015 to 0.06]													
	β-lactam / β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid	0.0	0.7	[0.0 - 3.8]	[Shaded area from 0.015 to 0.50]													
		Cephems	Ceftiofur	0.0	0.0	[0.0 - 2.5]	[Shaded area from 0.015 to 0.125]												
		Ceftriaxone	0.0	0.0	[0.0 - 2.5]	[Shaded area from 0.015 to 0.25]													
	Penicillins	Ampicillin	0.0	2.1	[0.4 - 5.9]	[Shaded area from 0.015 to 0.125]													
		Quinolones	Ciprofloxacin	0.0	0.0	[0.0 - 2.5]	[Shaded area from 0.015 to 0.06]												
		Nalidixic acid	N/A	90.4	[84.4 - 94.7]	[Shaded area from 0.015 to 0.015]													
		Tetracyclines	Tetracycline	0.0	2.1	[0.4 - 5.9]	[Shaded area from 0.015 to 0.06]												
II	Aminoglycosides	Kanamycin	0.0	0.7	[0.0 - 3.8]	[Shaded area from 0.015 to 0.50]													
	Cephems	Cefoxitin	3.4	0.0	[0.0 - 2.5]	[Shaded area from 0.015 to 0.50]													
	Folate pathway inhibitors	Sulfisoxazole	N/A	1.4	[0.2 - 4.9]	[Shaded area from 0.015 to 0.06]													
		Trimethoprim-sulfamethoxazole	N/A	2.1	[0.4 - 5.9]	[Shaded area from 0.015 to 0.06]													
	Phenolics	Chloramphenicol	15.8	1.4	[0.2 - 4.9]	[Shaded area from 0.015 to 0.06]													

* Rank of antimicrobial agents is based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important
 † CLSI: Clinical and Laboratory Standards Institute
 ‡ Percent of isolates with intermediate susceptibility; N/A indicates that no MIC range of intermediate susceptibility exists
 § Percent of isolates that were resistant
 ¶ The 95% confidence intervals (CI) for percent resistant (%R) were calculated using the Paulson-Camp-Pratt approximation to the Clopper-Pearson exact method
 ** The unshaded areas indicate the dilution range of the Sensititre plates used to test isolates. Single vertical bars indicate the breakpoints for susceptibility, while double vertical bars indicate breakpoints for resistance. Numbers in the shaded areas indicate the percentages of isolates with MICs greater than the highest concentrations on the Sensititre plate. Numbers listed for the low est tested concentrations represent the percentages of isolates with MICs equal to or less than the low est tested concentration. CLSI breakpoints were used when available.

Figure 20. Antimicrobial resistance pattern for Salmonella ser. Paratyphi A, Paratyphi B, and Paratyphi C, 2010

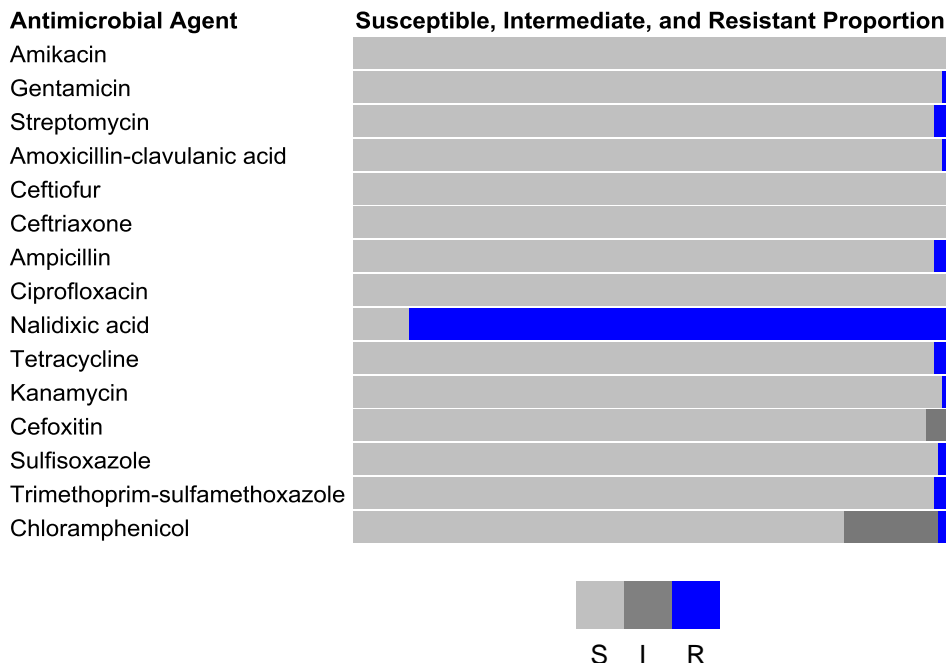


Table 30. Percentage and number of *Salmonella ser. Paratyphi A, Paratyphi B, and Paratyphi C* isolates resistant to antimicrobial agents, 2001–2010

Year			2001	2002	2003	2004	2005	2006	2007	2008	2009	2010	
Total Isolates			9	10	8	11	18	15	17	92	101	146	
Rank*	CLSI† Antimicrobial Class	Antimicrobial Agent (Resistance breakpoint)											
I	Aminoglycosides	Amikacin (MIC ≥ 64)	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	
		Gentamicin (MIC ≥ 16)	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.7% 1	
		Streptomycin (MIC ≥ 64)	0.0% 0	10.0% 1	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	1.0% 1	2.1% 3
	β-lactam/β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid (MIC ≥ 32/16)	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.7% 1
	Cepheems	Ceftiofur (MIC ≥ 8)	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0
		Ceftriaxone (MIC ≥ 4)	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0
	Penicillins	Ampicillin (MIC ≥ 32)	0.0% 0	0.0% 0	12.5% 1	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	1.0% 1	2.1% 3
	Quinolones	Ciprofloxacin (MIC ≥ 4)	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0
		Nalidixic Acid (MIC ≥ 32)	55.6% 5	40.0% 4	75.0% 6	72.7% 8	66.7% 12	53.3% 8	94.1% 16	87.0% 80	86.1% 87	90.4% 132	
	Tetracyclines	Tetracycline (MIC ≥ 16)	0.0% 0	10.0% 1	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	1.1% 1	1.0% 1	2.1% 3
II	Aminoglycosides	Kanamycin (MIC ≥ 64)	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.7% 1
		Cepheems	Cefoxitin (MIC ≥ 32)	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0
		Cephalothin (MIC ≥ 32)	0.0% 0	0.0% 0	0.0% 0	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested
	Folate pathway inhibitors	Sulfamethoxazole/Sulfisoxazole‡ (MIC ≥ 512)	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	1.0% 1	1.4% 2
		Trimethoprim-sulfamethoxazole (MIC ≥ 4/76)	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	1.0% 1	2.1% 3
	Phenicol	Chloramphenicol (MIC ≥ 32)	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	1.0% 1	1.4% 2

* Rank of antimicrobial agents is based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important

† CLSI: Clinical and Laboratory Standards Institute

‡ Sulfamethoxazole, which was tested during 1996-2003 to represent sulfonamides, was replaced by sulfisoxazole in 2004

Table 31. Resistance patterns of *Salmonella ser. Paratyphi A, Paratyphi B, and Paratyphi C* isolates, 2001–2010

Year	2001	2002	2003	2004	2005	2006	2007	2008	2009	2010
Total Isolates	9	10	8	11	18	15	17	92	101	146
Resistance Pattern										
No resistance detected	44.4% 4	50.0% 5	12.5% 1	27.3% 3	33.3% 6	46.7% 7	5.9% 1	12.0% 11	12.9% 13	6.8% 10
Resistance ≥ 1 CLSI class*	55.6% 5	50.0% 5	87.5% 7	72.7% 8	66.7% 12	53.3% 8	94.1% 16	88.0% 81	87.1% 88	93.2% 136
Resistance ≥ 2 CLSI classes*	0.0% 0	10.0% 1	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	1.0% 1	3.4% 5
Resistance ≥ 3 CLSI classes*	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	1.0% 1	2.1% 3
Resistance ≥ 4 CLSI classes*	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	1.0% 1	1.4% 2
Resistance ≥ 5 CLSI classes*	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	1.0% 1	0.7% 1
At least ACSSuT†	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	1.0% 1	0.7% 1
At least ACT/S‡	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	1.0% 1	0.7% 1
At least ACSSuTAuCx§	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0
At least ceftriaxone and nalidixic acid resistant	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0

* CLSI: Clinical and Laboratory Standards Institute

† ACSSuT: resistance to ampicillin, chloramphenicol, streptomycin, sulfamethoxazole/sulfisoxazole, tetracycline

‡ ACT/S: resistance to ampicillin, chloramphenicol, trimethoprim-sulfamethoxazole

§ ACSSuTAuCx: resistance to ACSSuT, amoxicillin-clavulanic acid, ceftriaxone

Figure 21. Antimicrobial resistance pattern for *Shigella*, 2010

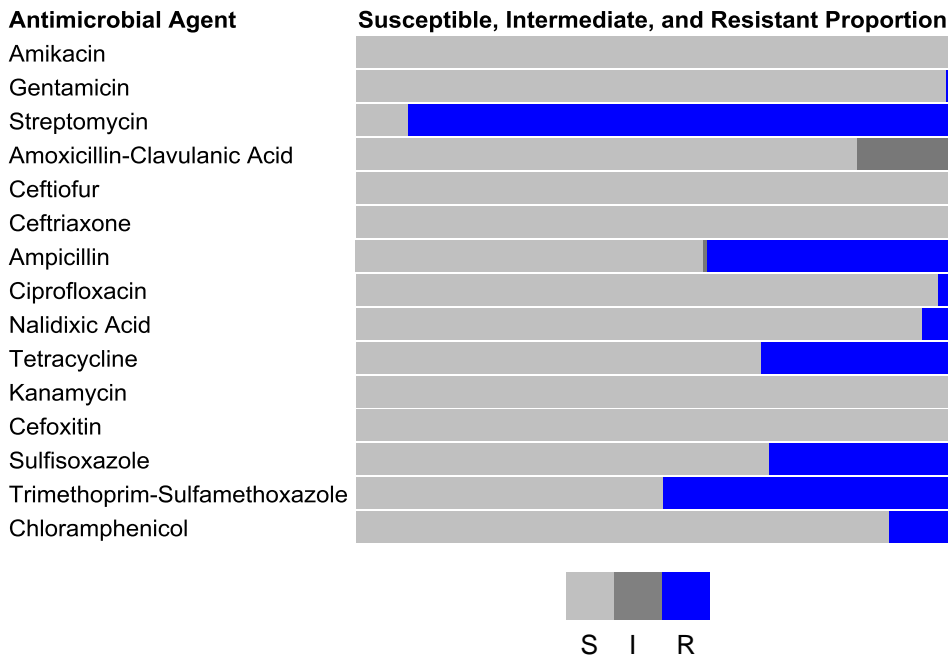


Table 34. Percentage and number of *Shigella* isolates resistant to antimicrobial agents, 2001–2010

Year	2001	2002	2003	2004	2005	2006	2007	2008	2009	2010		
Total Isolates	344	620	495	316	396	402	480	551	475	407		
Rank*	CLSI† Antimicrobial Class	Antimicrobial Agent (Resistance breakpoint)										
I	Aminoglycosides	Amikacin (MIC ≥ 64)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	
		Gentamicin (MIC ≥ 16)	0.0%	0.2%	0.0%	0.0%	1.0%	0.2%	0.8%	0.4%	0.6%	0.5%
		Streptomycin (MIC ≥ 64)	53.2%	54.4%	57.0%	59.8%	68.7%	60.7%	73.3%	80.6%	89.1%	91.2%
	β-lactam/β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid (MIC ≥ 32/16)	4.4%	2.6%	1.4%	1.6%	1.0%	1.5%	0.4%	3.3%	2.1%	0.0%
		Cephems	Ceftiofur (MIC ≥ 8)	0.0%	0.2%	0.2%	0.3%	0.5%	0.2%	0.0%	0.0%	0.6%
	Cephems	Ceftriaxone (MIC ≥ 4)	0.0%	0.2%	0.2%	0.3%	0.5%	0.2%	0.0%	0.0%	0.6%	0.2%
		Penicillins	Ampicillin (MIC ≥ 32)	79.7%	76.6%	79.4%	77.5%	70.7%	62.4%	63.8%	62.4%	46.3%
	Quinolones	Ciprofloxacin (MIC ≥ 4)	0.3%	0.0%	0.0%	0.0%	0.0%	0.2%	0.2%	0.7%	0.6%	1.7%
		Nalidixic Acid (MIC ≥ 32)	1.7%	1.6%	1.0%	1.6%	1.5%	3.5%	1.7%	1.6%	2.1%	4.4%
	Tetracyclines	Tetracycline (MIC ≥ 16)	59.3%	30.6%	29.1%	49.4%	38.4%	34.6%	25.6%	24.3%	29.5%	31.7%
II	Aminoglycosides	Kanamycin (MIC ≥ 64)	0.6%	0.8%	0.4%	0.0%	0.8%	0.0%	0.2%	0.5%	0.4%	0.0%
		Cephems	Cefoxitin (MIC ≥ 32)	1.2%	0.3%	0.0%	0.3%	0.3%	0.0%	0.0%	0.0%	0.6%
	Cephems	Cephalothin (MIC ≥ 32)	9.0%	6.6%	9.3%	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested
		Folate pathway inhibitors	Sulfamethoxazole/Sulfisoxazole‡ (MIC ≥ 512)	56.4%	31.8%	33.9%	52.5%	57.6%	40.3%	25.8%	28.5%	30.5%
	Folate pathway inhibitors	Trimethoprim-sulfamethoxazole (MIC ≥ 4/76)	46.8%	37.3%	38.6%	46.8%	53.3%	46.0%	25.8%	31.2%	40.4%	48.2%
		Phenicol	Chloramphenicol (MIC ≥ 32)	21.5%	7.6%	8.5%	15.2%	10.9%	10.9%	8.3%	6.9%	9.3%

* Rank of antimicrobial agents is based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important

† CLSI: Clinical and Laboratory Standards Institute

‡ Sulfamethoxazole, which was tested during 1996-2003 to represent sulfonamides, was replaced by sulfisoxazole in 2004

Figure 22. Antimicrobial resistance pattern for *Shigella sonnei*, 2010

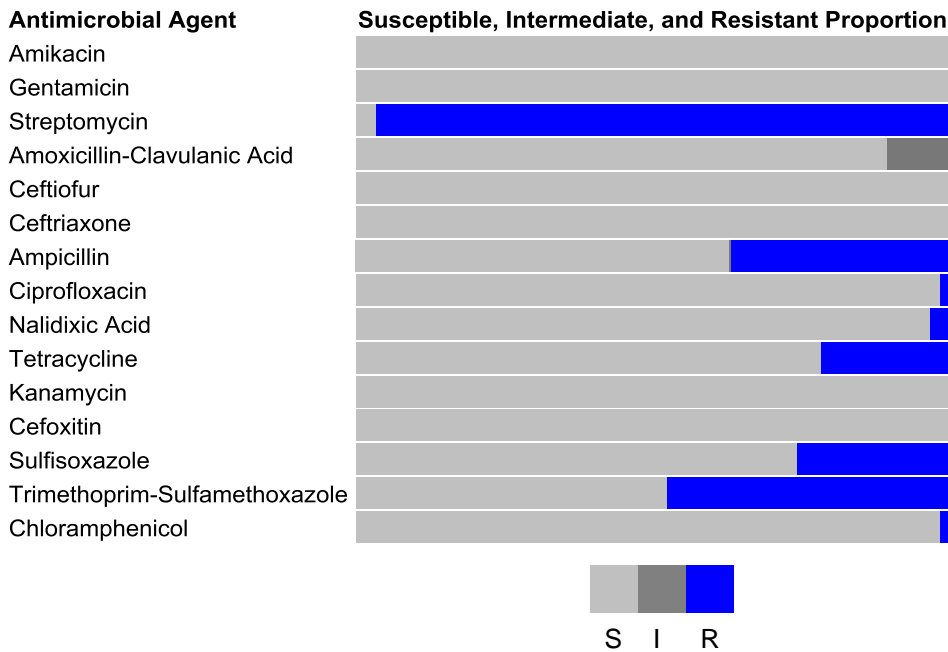


Table 37. Percentage and number of *Shigella sonnei* isolates resistant to antimicrobial agents, 2001–2010

Year	2001	2002	2003	2004	2005	2006	2007	2008	2009	2010		
Total Isolates	239	536	434	241	340	321	414	497	410	333		
Rank*	CLSI† Antimicrobial Class	Antimicrobial Agent (Resistance breakpoint)										
I	Aminoglycosides	Amikacin (MIC ≥ 64)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	
		Gentamicin (MIC ≥ 16)	0.0%	0.0%	0.0%	0.0%	1.2%	0.0%	1.0%	0.4%	0.7%	0.0%
		Streptomycin (MIC ≥ 64)	54.0%	55.4%	56.5%	56.8%	70.3%	61.7%	76.8%	82.3%	91.5%	96.4%
	β-lactam/β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid (MIC ≥ 32/16)	4.6%	2.2%	1.4%	1.7%	1.2%	1.9%	0.5%	3.2%	2.0%	0.0%
			11	12	6	4	4	6	2	16	8	0
	Cephems	Ceftiofur (MIC ≥ 8)	0.0%	0.0%	0.0%	0.4%	0.6%	0.0%	0.0%	0.0%	0.5%	0.3%
			0	0	0	1	2	0	0	0	2	1
	Ceftriaxone (MIC ≥ 4)		0.0%	0.0%	0.0%	0.4%	0.6%	0.0%	0.0%	0.0%	0.5%	0.3%
			0	0	0	1	2	0	0	0	2	1
	Penicillins	Ampicillin (MIC ≥ 32)	82.8%	77.6%	79.7%	79.3%	70.6%	62.6%	64.0%	61.4%	43.2%	36.6%
			198	416	346	191	240	201	265	305	177	122
Quinolones	Ciprofloxacin (MIC ≥ 4)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.6%	0.0%	1.5%	
		0	0	0	0	0	0	0	3	0	5	
Nalidixic Acid (MIC ≥ 32)		0.8%	1.5%	0.5%	1.7%	1.2%	2.8%	1.2%	1.6%	1.7%	3.3%	
		2	8	2	4	4	9	5	8	7	11	
Tetracyclines	Tetracycline (MIC ≥ 16)	44.8%	23.5%	22.1%	36.1%	29.4%	22.7%	16.2%	17.3%	20.7%	21.6%	
		107	126	96	87	100	73	67	86	85	72	
II	Aminoglycosides	Kanamycin (MIC ≥ 64)	0.4%	0.4%	0.0%	0.0%	0.0%	0.0%	0.2%	0.6%	0.2%	0.0%
			1	2	0	0	0	0	1	3	1	0
	Cephems	Cefoxitin (MIC ≥ 32)	1.7%	0.4%	0.0%	0.4%	0.3%	0.0%	0.0%	0.0%	0.7%	0.0%
			4	2	0	1	1	0	0	0	3	0
	Cephalothin (MIC ≥ 32)		12.6%	7.3%	10.1%	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested
			30	39	44							
	Folate pathway inhibitors	Sulfamethoxazole/Sulfisoxazole‡ (MIC ≥ 512)	54.4%	29.9%	31.3%	49.0%	57.9%	33.3%	20.0%	24.9%	23.9%	25.5%
		130	160	136	118	197	107	83	124	98	85	
Trimethoprim-sulfamethoxazole (MIC ≥ 4/76)		50.6%	37.9%	38.5%	46.9%	55.0%	42.7%	22.0%	29.4%	36.1%	47.4%	
		121	203	167	113	187	137	91	146	148	158	
Phenicols	Chloramphenicol (MIC ≥ 32)	1.3%	0.2%	1.2%	2.5%	2.4%	0.9%	1.2%	1.0%	1.2%	1.5%	
		3	1	5	6	8	3	5	5	5	5	

* Rank of antimicrobial agents is based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important

† CLSI: Clinical and Laboratory Standards Institute

‡ Sulfamethoxazole, which was tested during 1996-2003 to represent sulfonamides, was replaced by sulfisoxazole in 2004

Figure 23. Antimicrobial resistance pattern for *Shigella flexneri*, 2010

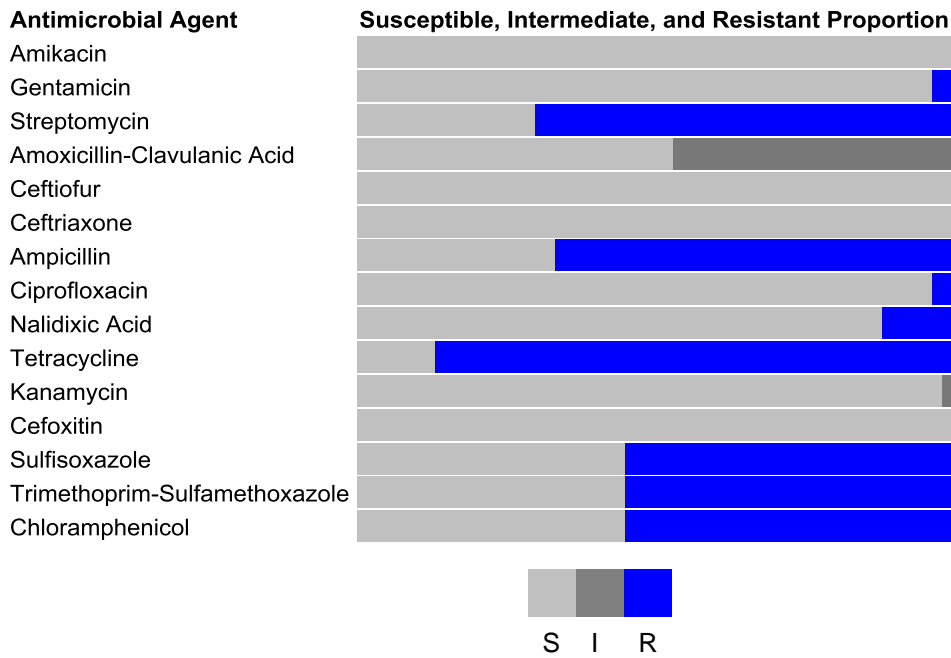


Table 40. Percentage and number of *Shigella flexneri* isolates resistant to antimicrobial agents, 2001–2010

Year	2001	2002	2003	2004	2005	2006	2007	2008	2009	2010		
Total Isolates	91	73	51	62	52	74	61	46	57	60		
Rank*	CLSI† Antimicrobial Class	Antimicrobial Agent (Resistance breakpoint)										
I	Aminoglycosides	Amikacin (MIC ≥ 64)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	
		Gentamicin (MIC ≥ 16)	0.0%	1.4%	0.0%	0.0%	0.0%	1.4%	0.0%	0.0%	3.3%	
		Streptomycin (MIC ≥ 64)	47.3%	43.8%	60.8%	71.0%	57.7%	58.1%	52.5%	63.0%	73.7%	70.0%
	β-lactam/β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid (MIC ≥ 32/16)	4.4%	5.5%	2.0%	1.6%	0.0%	0.0%	0.0%	4.3%	3.5%	0.0%
	Cephems	Cefotiofur (MIC ≥ 8)	0.0%	1.4%	2.0%	0.0%	0.0%	1.4%	0.0%	0.0%	1.8%	0.0%
		Ceftriaxone (MIC ≥ 4)	0.0%	1.4%	2.0%	0.0%	0.0%	1.4%	0.0%	0.0%	1.8%	0.0%
	Penicillins	Ampicillin (MIC ≥ 32)	72.5%	75.3%	84.3%	80.6%	75.0%	63.5%	63.9%	76.1%	70.2%	66.7%
	Quinolones	Ciprofloxacin (MIC ≥ 4)	1.1%	0.0%	0.0%	0.0%	0.0%	1.4%	1.6%	2.2%	3.5%	3.3%
		Nalidixic Acid (MIC ≥ 32)	3.3%	2.7%	5.9%	1.6%	3.8%	5.4%	4.9%	2.2%	3.5%	11.7%
	Tetracyclines	Tetracycline (MIC ≥ 16)	94.5%	78.1%	82.4%	95.2%	94.2%	83.8%	83.6%	87.0%	87.7%	86.7%
II	Aminoglycosides	Kanamycin (MIC ≥ 64)	1.1%	4.1%	3.9%	0.0%	3.8%	0.0%	0.0%	0.0%	1.8%	0.0%
	Cephems	Cefoxitin (MIC ≥ 32)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
		Cephalothin (MIC ≥ 32)	1.1%	2.7%	3.9%	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested
	Folate pathway inhibitors	Sulfamethoxazole/Sulfisoxazole‡ (MIC ≥ 512)	57.1%	41.1%	52.9%	66.1%	55.8%	68.9%	62.3%	60.9%	73.7%	55.0%
		Trimethoprim-sulfamethoxazole (MIC ≥ 4/76)	34.1%	28.8%	39.2%	46.8%	44.2%	59.5%	49.2%	47.8%	68.4%	55.0%
	Phenicols	Chloramphenicol (MIC ≥ 32)	74.7%	63.0%	68.6%	61.3%	65.4%	54.1%	55.7%	67.4%	66.7%	55.0%

* Rank of antimicrobial agents is based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important

† CLSI: Clinical and Laboratory Standards Institute

‡ Sulfamethoxazole, which was tested during 1996-2003 to represent sulfonamides, was replaced by sulfisoxazole in 2004

Table 41. Resistance patterns of *Shigella flexneri* isolates, 2001–2010

Year	2001	2002	2003	2004	2005	2006	2007	2008	2009	2010
Total Isolates	91	73	51	62	52	74	61	46	57	60
Resistance Pattern										
No resistance detected	3.3% 3	15.1% 11	7.8% 4	0.0% 0	5.8% 3	5.4% 4	9.8% 6	4.3% 2	5.3% 3	10.0% 6
Resistance ≥ 1 CLSI class*	96.7% 88	84.9% 62	92.2% 47	100.0% 62	94.2% 49	94.6% 70	90.2% 55	95.7% 44	94.7% 54	90.0% 54
Resistance ≥ 2 CLSI classes*	89.0% 81	76.7% 56	86.3% 44	93.5% 58	80.8% 42	85.1% 63	80.3% 49	93.5% 43	86.0% 49	83.3% 50
Resistance ≥ 3 CLSI classes*	79.1% 72	75.3% 55	80.4% 41	90.3% 56	78.8% 41	75.7% 56	68.9% 42	84.8% 39	82.5% 47	80.0% 48
Resistance ≥ 4 CLSI classes*	62.6% 57	57.5% 42	62.7% 32	64.5% 40	65.4% 34	47.3% 35	55.7% 34	56.5% 26	63.2% 36	56.7% 34
Resistance ≥ 5 CLSI classes*	25.3% 23	19.2% 14	31.4% 16	29.0% 18	30.8% 16	28.4% 21	27.9% 17	28.3% 13	49.1% 28	28.3% 17
At least ACSSuT†	22.0% 20	15.1% 11	29.4% 15	27.4% 17	28.8% 15	27.0% 20	26.2% 16	23.9% 11	47.4% 27	26.7% 16
At least ACT/S‡	23.1% 21	21.9% 16	27.5% 14	24.2% 15	32.7% 17	28.4% 21	26.2% 16	26.1% 12	47.4% 27	26.7% 16
At least AT/S§	25.3% 23	27.4% 20	37.3% 19	35.5% 22	38.5% 20	43.2% 32	36.1% 22	32.6% 15	52.6% 30	40.0% 24
At least ANT/S¶	1.1% 1	1.4% 1	5.9% 3	0.0% 0	1.9% 1	2.7% 2	1.6% 1	0.0% 0	1.8% 1	8.3% 5
At least ACSSuTAuCx**	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0	0.0% 0
At least ceftriaxone and nalidixic acid resistant	0.0% 0	0.0% 0	2.0% 1	0.0% 0	0.0% 0	1.4% 1	0.0% 0	0.0% 0	0.0% 0	0.0% 0

* CLSI: Clinical and Laboratory Standards Institute

† ACSSuT: resistance to ampicillin, chloramphenicol, streptomycin, sulfamethoxazole/sulfisoxazole, tetracycline

‡ ACT/S: resistance to ampicillin, chloramphenicol, trimethoprim-sulfamethoxazole

§ AT/S: resistance to ampicillin, trimethoprim-sulfamethoxazole

¶ ANT/S: resistance to AT/S, nalidixic acid

** ACSSuTAuCx: resistance to ACSSuT, amoxicillin-clavulanic acid, ceftriaxone

Table 43. Percentage and number of *Escherichia coli* O157 isolates resistant to antimicrobial agents, 2001–2010

Year		2001	2002	2003	2004	2005	2006	2007	2008	2009	2010	
Total Isolates		277	399	158	169	194	233	190	160	187	167	
Rank*	CLSI† Antimicrobial Class	Antimicrobial Agent (Resistance breakpoint)										
I	Aminoglycosides	Amikacin (MIC ≥ 64)	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
		Gentamicin (MIC ≥ 16)	0.4%	0.0%	0.0%	0.6%	0.5%	0.0%	0.0%	1.3%	0.5%	0.6%
		Streptomycin (MIC ≥ 64)	1.8%	2.3%	1.9%	1.8%	2.1%	2.6%	2.1%	1.9%	4.8%	1.8%
	β-lactam/β-lactamase inhibitor combinations	Amoxicillin-clavulanic acid (MIC ≥ 32/16)	0.7%	0.0%	1.3%	0.0%	0.0%	1.3%	0.5%	0.6%	0.5%	0.0%
		Cephems	Ceftiofur (MIC ≥ 8)	1.1%	0.0%	1.3%	0.0%	0.0%	1.3%	0.0%	0.6%	0.0%
	Penicillins	Ceftriaxone (MIC ≥ 4)	0.7%	0.0%	1.3%	0.0%	0.0%	1.3%	0.0%	0.6%	0.0%	0.0%
		Ampicillin (MIC ≥ 32)	2.2%	1.5%	3.2%	1.2%	4.1%	2.6%	2.1%	3.8%	4.3%	1.8%
	Quinolones	Ciprofloxacin (MIC ≥ 4)	0.0%	0.0%	0.0%	0.0%	0.0%	0.4%	0.5%	0.0%	0.5%	0.0%
		Nalidixic Acid (MIC ≥ 32)	1.1%	1.0%	0.6%	1.8%	1.5%	2.1%	2.1%	1.3%	2.1%	1.2%
	Tetracyclines	Tetracycline (MIC ≥ 16)	5.4%	3.0%	5.7%	1.8%	8.8%	4.7%	4.7%	1.9%	7.5%	4.2%
II		Aminoglycosides	Kanamycin (MIC ≥ 64)	0.0%	0.5%	0.0%	0.0%	0.5%	0.4%	0.0%	0.0%	0.5%
	Cephems		Cefoxitin (MIC ≥ 32)	0.7%	0.0%	1.3%	0.6%	0.0%	1.3%	0.0%	1.3%	0.5%
	Folate pathway inhibitors	Cephalothin (MIC ≥ 32)	1.4%	1.5%	3.2%	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested
		Sulfamethoxazole/Sulfisoxazole‡ (MIC ≥ 512)	5.1%	3.5%	3.8%	1.8%	6.7%	3.0%	2.6%	3.1%	6.4%	4.2%
	Phenicol	Trimethoprim-sulfamethoxazole (MIC ≥ 4/76)	0.7%	0.5%	0.6%	0.0%	0.5%	0.4%	1.1%	1.3%	4.3%	1.2%
		Chloramphenicol (MIC ≥ 32)	1.4%	1.3%	1.3%	0.6%	1.0%	1.3%	0.5%	0.6%	1.1%	0.6%

* Rank of antimicrobial agents is based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important

† CLSI: Clinical and Laboratory Standards Institute

‡ Sulfamethoxazole, which was tested during 1996-2003 to represent sulfonamides, was replaced by sulfisoxazole in 2004

Table 44. Resistance patterns of *Escherichia coli* O157 isolates, 2001–2010

Year	2001	2002	2003	2004	2005	2006	2007	2008	2009	2010
Total Isolates	277	399	158	169	194	233	190	160	187	167
Resistance Pattern										
No resistance detected	91.3%	94.0%	90.5%	94.7%	87.6%	91.8%	92.1%	91.9%	89.8%	94.0%
	253	375	143	160	170	214	175	147	168	157
Resistance ≥ 1 CLSI class*	8.7%	6.0%	9.5%	5.3%	12.4%	8.2%	7.9%	8.1%	10.2%	6.0%
	24	24	15	9	24	19	15	13	19	10
Resistance ≥ 2 CLSI classes*	5.4%	3.8%	5.1%	2.4%	6.7%	4.7%	3.2%	3.1%	7.5%	4.2%
	15	15	8	4	13	11	6	5	14	7
Resistance ≥ 3 CLSI classes*	2.2%	2.0%	3.2%	1.2%	5.2%	3.4%	2.1%	2.5%	5.9%	3.6%
	6	8	5	2	10	8	4	4	11	6
Resistance ≥ 4 CLSI classes*	1.4%	0.8%	1.3%	0.6%	1.0%	2.1%	1.1%	1.3%	4.3%	1.8%
	4	3	2	1	2	5	2	2	8	3
Resistance ≥ 5 CLSI classes*	0.4%	0.0%	0.0%	0.0%	0.0%	0.9%	0.5%	0.0%	0.5%	0.0%
	1	0	0	0	0	2	1	0	1	0
At least ACSSuT†	0.4%	0.0%	0.0%	0.0%	0.0%	0.9%	0.0%	0.0%	0.0%	0.0%
	1	0	0	0	0	2	0	0	0	0
At least ACT/S‡	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.6%	0.0%	0.0%
	0	0	0	0	0	0	0	1	0	0
At least ACSSuTAuCx§	0.4%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
	1	0	0	0	0	0	0	0	0	0
At least ceftriaxone and nalidixic acid resistant	0.0%	0.0%	0.0%	0.0%	0.0%	0.4%	0.0%	0.0%	0.0%	0.0%
	0	0	0	0	0	1	0	0	0	0

* CLSI: Clinical and Laboratory Standards Institute

† ACSSuT: resistance to ampicillin, chloramphenicol, streptomycin, sulfamethoxazole/sulfisoxazole, tetracycline

‡ ACT/S: resistance to ampicillin, chloramphenicol, trimethoprim-sulfamethoxazole

§ ACSSuTAuCx: resistance to ACSSuT, amoxicillin-clavulanic acid, ceftriaxone

Table 47. Percentage and number of *Campylobacter* isolates resistant to antimicrobial agents, 2001–2010

Year			2001	2002	2003	2004	2005	2006	2007	2008	2009	2010
Total Isolates			384	354	328	347	890	816	1100	1155	1497	1310
Rank*	CLSI† Antimicrobial Class	Antimicrobial Agent (Resistance breakpoint)										
I	Aminoglycosides	Gentamicin (MIC ≥ 8)	0.0% 0	0.0% 0	0.3% 1	0.3% 1	0.7% 6	0.1% 1	0.6% 7	1.1% 13	0.9% 13	1.6% 21
		Ketolides	Not Tested	Not Tested	Not Tested	Not Tested	1.0% 9	1.6% 13	1.5% 16	2.5% 29	1.5% 22	1.6% 21
	Macrolides	Azithromycin (MIC ≥ 8)	2.1% 8	2.0% 7	0.9% 3	0.6% 2	1.9% 17	1.7% 14	2.0% 22	3.0% 35	1.7% 25	1.5% 19
		Erythromycin (MIC ≥ 32)	2.1% 8	1.4% 5	0.9% 3	0.3% 1	1.8% 16	1.7% 14	2.0% 22	3.0% 35	1.7% 25	1.5% 19
	Quinolones	Ciprofloxacin (MIC ≥ 4)	19.5% 75	20.1% 71	17.7% 58	19.0% 66	21.7% 193	19.6% 160	26.0% 286	23.0% 266	22.9% 343	22.4% 294
		Nalidixic Acid (MIC ≥ 64)	20.3% 78	20.6% 73	18.9% 62	19.6% 68	22.4% 199	20.1% 164	26.5% 291	23.5% 272	23.2% 347	22.7% 298
	Tetracyclines	Tetracycline (MIC ≥ 16)	40.9% 157	41.2% 146	38.4% 126	46.1% 160	40.6% 361	46.0% 375	44.4% 488	43.6% 504	43.6% 652	42.1% 551
II	Phenicol	Chloramphenicol (MIC ≥ 32)	0.3% 1	0.3% 1	0.0% 0	1.4% 5	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested	Not Tested
		Florfenicol† Susceptible breakpoint: (MIC ≤ 4)	Not Tested	Not Tested	Not Tested	Not Tested	0.6% 5	0.0% 0	0.0% 0	0.5% 6	0.5% 8	1.3% 17
III	Lincosamides	Clindamycin (MIC ≥ 8)	2.1% 8	2.0% 7	0.6% 2	2.0% 7	1.5% 13	2.0% 16	1.7% 19	2.8% 32	1.4% 21	1.7% 22

* Rank of antimicrobial agents is based on World Health Organization's categorization of critical importance in human medicine (Table I): Rank 1, Critically Important; Rank 2, Highly Important; Rank 3, Important

† CLSI: Clinical and Laboratory Standards Institute

‡ Only a susceptible breakpoint (≤ 4 µg/ml) has been established. In this report, isolates with an MIC ≥ 8 µg/ml are categorized as resistant

Table 48. Resistance patterns of *Campylobacter* isolates, 2001–2010

Year	2001	2002	2003	2004	2005	2006	2007	2008	2009	2010
Total Isolates	384	354	328	347	890	816	1100	1155	1497	1310
Resistance Pattern										
No resistance detected	49.2% 189	48.0% 170	50.9% 167	46.1% 160	48.4% 431	43.9% 358	45.2% 497	45.9% 530	46.4% 694	47.3% 620
Resistance ≥ 1 CLSI class*	50.8% 195	52.0% 184	49.1% 161	53.9% 187	51.6% 459	56.1% 458	54.8% 603	54.1% 625	53.6% 803	52.7% 690
Resistance ≥ 2 CLSI classes*	13.3% 51	12.7% 45	8.5% 28	14.1% 49	13.8% 123	12.0% 98	17.5% 192	15.6% 180	14.2% 212	14.3% 187
Resistance ≥ 3 CLSI classes*	1.6% 6	1.4% 5	0.9% 3	1.7% 6	1.8% 16	1.5% 12	1.7% 19	2.7% 31	1.7% 25	2.1% 28
Resistance ≥ 4 CLSI classes*	0.3% 1	0.0% 0	0.3% 1	0.3% 1	0.4% 4	0.5% 4	0.9% 10	1.4% 16	1.1% 16	0.8% 10
Resistance ≥ 5 CLSI classes*	0.0% 0	0.0% 0	0.3% 1	0.0% 0	0.1% 1	0.1% 1	0.6% 7	0.7% 8	0.5% 8	0.6% 8

* CLSI: Clinical and Laboratory Standards Institute

Box 1. Changes in Antimicrobial Resistance: 2010 vs. 2003–07

To understand changes in prevalence of antimicrobial resistance over time, we used logistic regression to compare the prevalence of specific antimicrobial resistance patterns among *Salmonella* and *Campylobacter* isolates tested in 2010 with the average prevalence of resistance in 2003–2007. Since 2003, all 50 states have participated in *Salmonella* surveillance and all 10 FoodNet sites have participated in *Campylobacter* surveillance. A description of the methods is included in this report (refer to Surveillance and Laboratory Testing Methods).

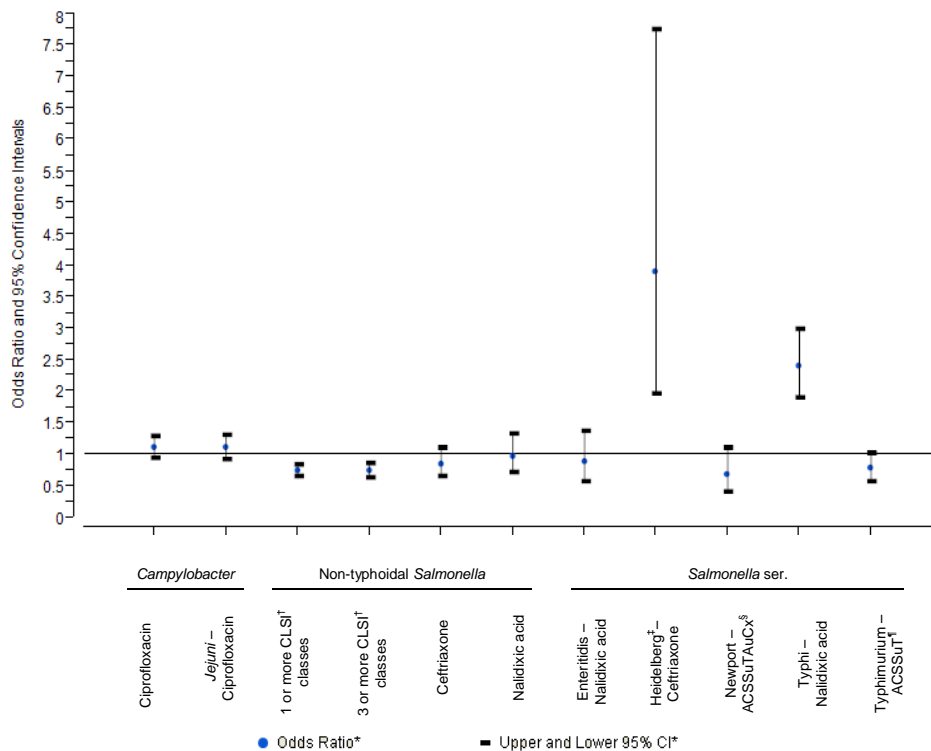
The differences between the prevalence of resistance in 2010 and the average prevalence of resistance in 2003–07 (Figure 1) were statistically significant for the following:

- Resistance to one or more CLSI classes in non-typhoidal *Salmonella* (NTS) was lower in 2010 than in 2003–2007 (Odds ratio [OR]=0.75, 95% Confidence interval [CI] 0.66–0.84)
 - Resistance to three or more CLSI classes in NTS was lower in 2010 than in 2003–2007 (OR=0.74, 95% CI 0.64–0.86)
 - Nalidixic acid resistance in *Salmonella* ser. Typhi was higher in 2010 than in 2003–2007 (OR=2.39, 95% CI 1.91–2.99)
 - Ceftriaxone resistance among *Salmonella* ser. Heidelberg was higher in 2010 than in 2003–2007 (OR=3.90, 95% CI 1.96–7.75)
- Descriptive analysis suggests that resistance in 2010 was mainly driven by New York, California, and Wisconsin. When trend analysis excluded these 3 states, there was no significant change (OR=2.26, 95% CI 0.86–5.93). Thus, the reported OR represents a summary of possibly unequal trends across sites.

The differences between the prevalence of resistance in 2010 and the average prevalence of resistance in 2003–07 (Figure 1) were not statistically significant for the following:

- Among *Campylobacter*
 - Ciprofloxacin resistance (OR=1.11, 95% CI 0.94–1.30)
 - Ciprofloxacin resistance in *Campylobacter jejuni* (OR=1.11, 95% CI 0.93–1.32)
- Among non-typhoidal *Salmonella* in general
 - Ceftriaxone resistance (OR=0.85, 95% CI 0.65–1.11)
 - Nalidixic acid resistance (OR=0.97, 95% CI 0.71–1.34)
- Among *Salmonella* of particular serotypes
 - Nalidixic acid resistance in ser. Enteritidis (OR=0.88, 95% CI 0.57–1.37)
 - ACSSuTAuCx resistance in ser. Newport (OR=0.67, 95% CI 0.41–1.11)
 - ACSSuT resistance in ser. Typhimurium (OR=0.77, 95% CI 0.58–1.03)

Figure 1. Summary of trend analysis of the prevalence of specific resistance patterns among *Salmonella* and *Campylobacter* isolates, 2010 compared with 2003–2007*



* The reference is the average prevalence of resistance in 2003–2007. Logistic regression models adjusted for site. The odds ratios (ORs) and 95% confidence intervals (CIs) for 2010 compared with the reference were calculated by using unconditional maximum likelihood estimation. ORs that do not include 1.00 in the 95% CIs are reported as statistically significant.

† Clinical and Laboratory Standards Institute (CLSI) antimicrobial classes of agents are used

‡ Descriptive analysis suggests that increased resistance in 2010 was mainly driven by New York, California, and Wisconsin. Thus, the reported OR represents a summary of possibly unequal trends across sites.

§ ACSSuTAuCx: resistance to at least ampicillin, chloramphenicol, streptomycin, sulfonamide, tetracycline, amoxicillin-clavulanic acid, and ceftriaxone

¶ ACSSuT: resistance to at least ampicillin, chloramphenicol, streptomycin, sulfonamide, and tetracycline

Box 2. Ciprofloxacin Breakpoint Changes for *Salmonella*

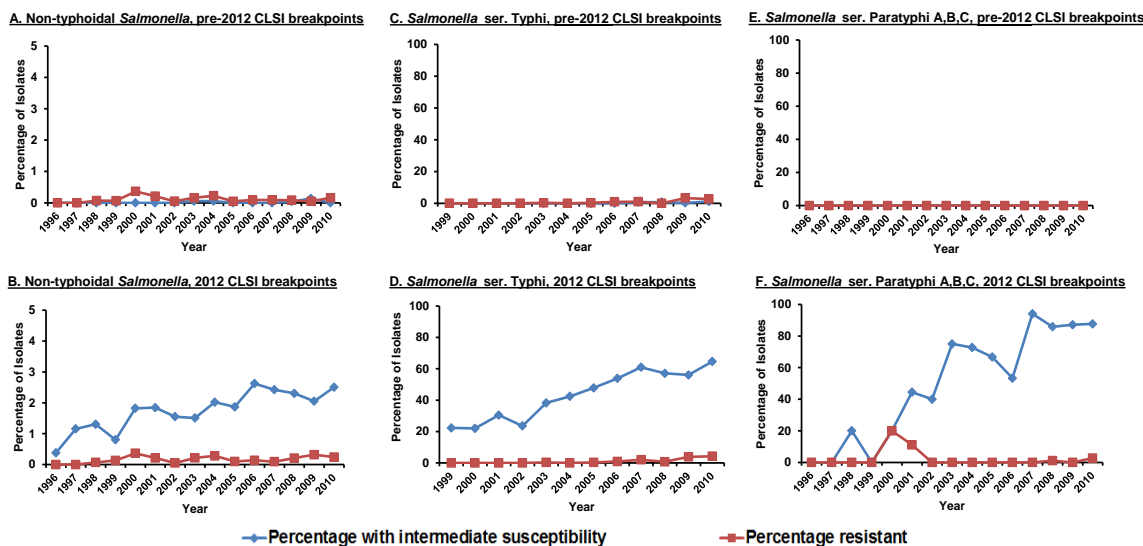
The Clinical and Laboratory Standards Institute (CLSI) is a consensus organization that publishes methods and interpretive criteria pertinent to clinical antimicrobial susceptibility testing. CLSI approved standards are used by NARMS and other entities throughout the world. CLSI reviewed fluoroquinolone interpretive criteria for *Enterobacteriaceae*. This process began with a review of the breakpoints for *Salmonella* infections. CLSI determined, after review of clinical and microbiologic data, that the MIC criteria for intermediate and resistant categories should be lowered for invasive *Salmonella* because patients whose isolates showed MICs in the susceptible range do not always respond to therapy with that class of agents; therefore, for invasive *Salmonella*, CLSI updated ciprofloxacin MIC ranges and disk diffusion correlates for susceptible (S), intermediate (I), and resistant (R) categories. These ranges appeared in the [January 2012 CLSI M100 supplement](#). Pre-2012 breakpoints defined isolates with MICs ≤ 1 $\mu\text{g/mL}$ as susceptible, isolates with an MIC of 2 $\mu\text{g/mL}$ as intermediate, and isolates with an MIC of ≥ 4 $\mu\text{g/mL}$ as resistant. The updated 2012 breakpoints defined the susceptible MIC range as ≤ 0.064 $\mu\text{g/mL}$, the intermediate range 0.12-0.5 $\mu\text{g/mL}$, and resistance as ≥ 1 $\mu\text{g/mL}$. To show how the data will change once the 2012 breakpoints are applied, in this report, we show S, I, and R frequencies for *Salmonella* (typhoidal and non-typhoidal) using both the outgoing and new breakpoints.

Table 1. Percentage of *Salmonella* isolates with intermediate susceptibility and resistance to ciprofloxacin, by pre-2012 and 2012 CLSI breakpoints, 1996–2010

Non-typhoidal <i>Salmonella</i>						<i>Salmonella</i> ser. Typhi				<i>Salmonella</i> ser. Paratyphi A, Paratyphi B, Paratyphi C							
Year	Total Isolates	Pre-2012 CLSI Breakpoints*		2012 CLSI Breakpoints [†]		Year	Total Isolates	Pre-2012 CLSI Breakpoints*		2012 CLSI Breakpoints [†]		Year	Total Isolates	Pre-2012 CLSI Breakpoints*		2012 CLSI Breakpoints [†]	
		%I [‡]	%R [§]	%I [‡]	%R [§]			%I [‡]	%R [§]	%I [‡]	%R [§]			%I [‡]	%R [§]		
1996	1318	0.0	0.0	0.4	0.0							1996	6	0.0	0.0	0.0	0.0
1997	1297	0.0	0.0	1.2	0.0							1997	4	0.0	0.0	0.0	0.0
1998	1455	0.0	0.1	1.3	0.1							1998	5	0.0	0.0	20.0	0.0
1999	1493	0.0	0.1	0.8	0.1	1999	166	0.0	0.0	22.3	0.0	1999	2	0.0	0.0	0.0	0.0
2000	1372	0.0	0.4	1.8	0.4	2000	177	0.0	0.0	22.0	0.0	2000	5	0.0	0.0	20.0	20.0
2001	1410	0.0	0.2	1.8	0.2	2001	197	0.0	0.0	30.5	0.0	2001	9	0.0	0.0	44.4	11.1
2002	1998	0.0	0.1	1.6	0.1	2002	195	0.0	0.0	23.6	0.0	2002	10	0.0	0.0	40.0	0.0
2003	1855	0.1	0.2	1.5	0.2	2003	332	0.0	0.3	38.3	0.3	2003	8	0.0	0.0	75.0	0.0
2004	1782	0.1	0.2	2.0	0.3	2004	304	0.0	0.0	42.4	0.0	2004	11	0.0	0.0	72.7	0.0
2005	2034	0.0	<0.1	1.9	0.1	2005	318	0.0	0.3	47.8	0.3	2005	18	0.0	0.0	66.7	0.0
2006	2172	0.0	0.1	2.6	0.1	2006	323	0.0	0.9	53.9	0.9	2006	15	0.0	0.0	53.3	0.0
2007	2145	0.0	0.1	2.4	0.1	2007	400	0.8	1.0	61.0	2.0	2007	17	0.0	0.0	94.1	0.0
2008	2384	<0.1	0.1	2.3	0.2	2008	408	0.7	0.0	57.1	0.7	2008	92	0.0	0.0	85.9	1.1
2009	2193	0.1	<0.1	2.1	0.3	2009	362	0.3	3.3	56.1	3.9	2009	101	0.0	0.0	87.1	0.0
2010	2474	0.0	0.2	2.5	0.2	2010	444	1.1	2.7	64.6	4.3	2010	146	0.0	0.0	87.7	2.7

* The current CLSI breakpoints used for ciprofloxacin in this report are: Resistant (R) MIC ≥ 4 $\mu\text{g/mL}$, Intermediate (I) MIC=2 $\mu\text{g/mL}$
[†] The new CLSI breakpoints for ciprofloxacin that will be used in the 2011 NARMS Reports are: Resistant (R) MIC ≥ 1 $\mu\text{g/mL}$, Intermediate (I) MIC=0.12-0.5 $\mu\text{g/mL}$
[‡] Percentage of isolates with intermediate susceptibility to ciprofloxacin
[§] Percentage of isolates that were resistant to ciprofloxacin

Figure 1. Percentage of *Salmonella* isolates with intermediate susceptibility and resistance to ciprofloxacin, by pre-2012 and 2012 CLSI breakpoints, 1996–2010



References

- CDC. [National Antimicrobial Resistance Monitoring System for Enteric Bacteria \(NARMS\): 2005 human isolates final report](#). Atlanta, Georgia: U.S. Department of Health and Human Services, CDC, 2007.
- Clinical and Laboratory Standards Institute. [Methods for antimicrobial dilution and disk susceptibility testing of infrequently isolated or fastidious bacteria: approved guideline—Second Edition](#). CLSI Document M45-A2. CLSI, Wayne, Pennsylvania, 2010.
- Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing; Twenty-First Informational Supplement. CLSI Document M100-S21. CLSI, Wayne, Pennsylvania, 2011.
- Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing; Twenty-Second Informational Supplement. CLSI Document M100-S22. CLSI, Wayne, Pennsylvania, 2012.
- Clinical and Laboratory Standards Institute. [Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically; Approved Standard—Eighth Edition](#). CLSI Document M07-A8. CLSI, Wayne, Pennsylvania, 2009.
- Clinical and Laboratory Standards Institute. [Performance standards for antimicrobial disk and dilution susceptibility tests for bacteria isolated from animals: Approved Standard—Third Edition](#). CLSI Document M31-A3. CLSI, Wayne, Pennsylvania, 2008.
- Fleiss JL, Levin B, Paik MC. [Statistical methods in for rates and proportions](#). In: Shewart WA, Wilks SS, eds. [Wiley Series in Probability and Statistics](#). Published Online; 2004:284–308.
- Gonzalez, I, Grant KA, Richardson PT, Park SF, Collins MD. [Specific identification of the enteropathogens *Campylobacter jejuni* and *Campylobacter coli* by using a PCR test based on the *ceuE* gene encoding a putative virulence determinant](#). Journal of Clinical Microbiology 1997;35:759–63.
- Linton D, Lawson AJ, Owen RJ, Stanley J. [PCR detection, identification to species level, and fingerprinting of *Campylobacter jejuni* and *Campylobacter coli* direct from diarrheic samples](#). Journal of Clinical Microbiology 1997;35:2568–72.
- Linton D, Owen RJ, Stanley J. [Rapid Identification by PCR of the genus *Campylobacter* and of five *Campylobacter* species enteropathogenic for man and animals](#). Research in Microbiology 1996;147:707–18.
- Pruckler J et al., Comparison of four real-time PCR methods for the identification of the genus *Campylobacter* and speciation of *C. jejuni* and *C. coli*. ASM 106th General meeting; Poster C282.
- U.S. Census Bureau. [Guide to State and Local Geography – Selected Data from the 2010 Census](#). Washington, D.C.: U.S. Department of Commerce, U.S. Census Bureau, 2011.
- U.S. Census Bureau. [Census Regions and Divisions of the United States](#). Washington, D.C.: U.S. Department of Commerce, U.S. Census Bureau, 2011.
- World Health Organization (WHO). [Critically Important Antimicrobials for Human Medicine. 2nd Revision](#). Switzerland, 2009.
- Vandamme P, Van Doorn LJ, al Rashid ST, Quint WG, van der Plas J, Chan VL, On SL. [*Campylobacter hyoilei* Alderton et al. 1995 and *Campylobacter coli* Veron and Chatelain 1973 are subjective synonyms](#). Inter. J. Syst. Bacteriol 1997; 47:1055–60.

Folster JP, Pecic G, Krueger A, Rickert R, Burger K, Carattoli A, Whichard JM. [Identification and characterization of CTX-M-producing *Shigella* isolates in the United States](#). *Antimicrob. Agents Chemother.* 2010;54 (5):2269-70.

Folster JP, Pecic G, Bolcen S, Theobald L, Hise K, Carattoli A, Zhao S, McDermott PF, Whichard JM. [Characterization of extended-spectrum cephalosporin-resistant *Salmonella enterica* serovar Heidelberg isolated from humans in the United States](#). *Foodborne Pathog. Dis.* 2010;7 (2):181-7.

Howie RL, Folster JP, Bowen A, Barzilay EJ, Whichard JM. [Reduced Azithromycin susceptibility in *Shigella sonnei*, United States](#). *Microb. Drug Resist.* 2010;16(4):245-8.

Krueger AL, Folster J, Medalla F, Joyce K, Perri MB, Johnson L, Zervoz M, Whichard JM, Barzilay EJ. [Commensal *Escherichia coli* isolate resistant to eight classes of antimicrobial agents in the United States](#). *Foodborne Pathog. Dis.* 2011;8(2):329-32.

M'ikanatha NM, Sandt CH, Localio AR, Tewari D, Rankin SC, Whichard JM, Altekruze SF, Lautenbach E, Folster JP, Russo A, Chiller TM, Reynolds SM, McDermott PF. [Multidrug-resistant *Salmonella* isolates from retail chicken meat compared with human clinical isolates](#). *Foodborne Pathog. Dis.* 2010;7 (8):929-34.

Sjölund-Karlsson M, Howie R, Rickert R, Krueger A, Tran TT, Zhao S, Ball T, Haro J, Pecic G, Joyce K, Fedorka-Cray PJ, Whichard JM, McDermott PF. [Plasmid-mediated quinolone resistance among non-Typhi *Salmonella enterica* isolates, USA](#). *Emerging Infectious Diseases* 2010;16 (11):1789-91.

Sjölund-Karlsson M, Rickert R, Matar C, Pecic G, Howie RL, Joyce K, Medalla F, Barzilay EJ, Whichard JM. [*Salmonella* isolates with decreased susceptibility to extended-spectrum cephalosporins in the United States](#). *Foodborne Pathog. Dis.* 2010;7 (12):1503-9.

Whichard JM, Medalla F, Hoekstra RM, McDermott PF, Joyce K, Chiller T, Barrett TJ, White DG. [Evaluation of antimicrobial resistance phenotypes for predicting multidrug-resistant *Salmonella* recovered from retail meats and humans in the United States](#). *J. Food Prot.* 2010;73 (3):445-51.

Appendix A

Summary of Non-Typhoidal *Salmonella* Strains that Caused Outbreaks, United States, 2004–2008

BACKGROUND

Antimicrobial resistance among *Salmonella* has important public health implications. Treatment with antimicrobial agents is critical for persons with severe *Salmonella* infections, especially older adults, children, and immunocompromised patients. First-line agents for the treatment of severe *Salmonella* infections include fluoroquinolones (e.g., ciprofloxacin) and extended-spectrum cephalosporins (e.g., ceftriaxone).^{1, 2} Monitoring resistance to these and other important antimicrobial agents is crucial because antimicrobial use in food-producing animals may result in resistance among enteric bacteria, which can be transmitted to humans through food. Surveillance of resistance among enteric bacteria transmitted commonly through food is performed by the National Antimicrobial Resistance Monitoring System (NARMS).

To aid in *Salmonella* outbreak investigations, NARMS collects isolates and performs antimicrobial susceptibility testing to determine resistance patterns. Antimicrobial susceptibility testing during outbreak investigations can help determine which food vehicles are associated with certain resistant patterns and provide information about food source attribution. We examined antimicrobial resistance among those isolates that were submitted to NARMS from non-typhoidal *Salmonella* outbreaks in the United States from 2004 through 2008.

METHODS

CDC asked public health laboratories to submit representative isolates to NARMS for antimicrobial susceptibility testing from all outbreaks caused by *Salmonella* serotypes Enteritidis, Newport, and Typhimurium that occurred from 2004 through 2008. CDC also asked sites in the Foodborne Diseases Active Surveillance Network (FoodNet) to submit representative isolates from all *Salmonella* outbreaks. CDC tested isolates using broth microdilution to determine the minimum inhibitory concentration (MIC) for 15 antimicrobial agents, which were categorized into eight classes: aminoglycosides (amikacin, gentamicin, kanamycin, streptomycin); β -lactam/ β -lactamase inhibitor combinations (amoxicillin-clavulanic acid); cepheims (cefoxitin, ceftiofur, ceftriaxone); penicillins (ampicillin); quinolones (ciprofloxacin, nalidixic acid); folate pathway inhibitors (sulfamethoxazole/sulfisoxazole, trimethoprim-sulfamethoxazole); phenicols (chloramphenicol); and tetracyclines (tetracycline). Antimicrobial classes and MIC resistance breakpoints were defined by using criteria established by the Clinical and Laboratory Standards Institute (CLSI).

A foodborne disease outbreak is defined as the occurrence of two or more similar illnesses that resulted from ingestion of a common food.³ Local, state, and territorial health departments voluntarily report outbreaks to CDC's Foodborne Disease Outbreak Surveillance System by submitting a standard web-based form.³ Data collected for each outbreak include the number of illnesses, hospitalizations, and deaths; etiologic agent; and the implicated food.³ CDC classifies foods into 1 of 17 commodities, which are categorized into three groups: aquatic animals (finfish, crustaceans, mollusks); land animals (dairy, eggs, beef, game, pork, poultry); and plants (grains-beans, oils-sugars, fruits-nuts, fungi, leafy, root, sprout, vine-stalk).³ Food items that contain ingredients from only one commodity were assigned to that commodity.³ Food items that contain ingredients from more than one commodity were classified as "complex" if the contaminated commodity was not determined, and food items were classified as "unknown" when the outbreak report provided insufficient information.³

Non-typhoidal *Salmonella* outbreak data were linked to isolate resistance data using a combination of variables including outbreak identification number, state, year, month, and serotype. The PulseNet-assigned *Xba*I pattern and PulseNet cluster code were used to validate if an isolate was part of a reported outbreak. Outbreaks were considered to be caused by a resistant bacterium if at least one isolate was resistant to ≥ 1 antimicrobial agents; outbreaks were considered to have no resistance detected if results for all drugs were either susceptible or intermediate. Additionally, multidrug resistance patterns were defined: ACSSuT if resistant to at least ampicillin, chloramphenicol, streptomycin, sulfamethoxazole/sulfisoxazole, and tetracycline; ACSSuTAuCx if resistant to at least ACSSuT, amoxicillin-clavulanic acid, and ceftriaxone; and ACT/S if resistant to at least ampicillin, chloramphenicol, and trimethoprim-sulfamethoxazole.

RESULTS

From 2004–2008, 592 non-typhoidal *Salmonella* outbreaks with known serotype information were reported to the Foodborne Disease Outbreak Surveillance System (FDOSS), and 484 outbreak isolates were submitted to NARMS. Isolates were submitted to NARMS for 103 (17%) of the outbreaks reported to FDOSS. The strain was resistant for 18 (17%) ([Table 1](#)) and no resistance detected for 85 (83%) ([Table 2](#)).

Of the 18 outbreaks with a resistant strain, 9 (50%) were caused by a strain resistant to at least ceftriaxone and 2 (11%) by a strain resistant to at least nalidixic acid ([Table 3](#)). Resistance was observed most often to tetracycline (15 outbreaks), followed by sulfamethoxazole/sulfisoxazole (13 outbreaks), amoxicillin-clavulanic acid (11 outbreaks), ampicillin (11 outbreaks), and streptomycin (11 outbreaks).

Seven (7%) of the 103 outbreaks were caused by a strain resistant to 1–4 agents and 11 (11%) by a strain resistant to ≥ 5 agents ([Table 4](#)). The multidrug resistance pattern ACSSuT was observed in 8 (8%) outbreaks; strains from 6 (75%) of these were also resistant to amoxicillin-clavulanic acid and ceftriaxone (ACSSuTAuCx).

Among the 47 outbreaks attributed to a single food commodity, 8 (17%) were caused by a resistant strain and 39 (83%) by strains with no resistance detected. Of the 8 outbreaks with a resistant strain, 4 (50%) were caused by strains that were resistant to ≥ 5 agents, including one caused by an ACSSuTAuCx resistant strain. Outbreaks attributed to a land animal commodity (e.g., beef, poultry, eggs, dairy) accounted for 6 (75%) of the 8 outbreaks caused by resistant strains and 22 (56%) of the 39 outbreaks caused by strains with no resistance detected.

CONCLUSIONS

Among *Salmonella* outbreaks attributed to a single food commodity and with information on resistance, land animal foods were identified as the predominant source of outbreaks caused by both resistant (6 of 8 outbreaks, 75%) and susceptible (22 of 39 outbreaks, 56%) strains. However, an isolate was received for a small proportion of outbreaks, so these findings may not be representative of all outbreaks. These data suggest that obtaining isolates from more outbreaks and determining their antimicrobial susceptibility could provide important information for food source attribution analyses.

REFERENCES

1. Guerrant RL, Van Gilder T, Steiner TS, Thielman NM, Slutsker L, Tauxe RV, Hennessy T, Griffin PM, DuPont H, Sack RB, et al. [Practice guidelines for the management of infectious diarrhea](#). Clin. Infect. Dis. 2001; 32: 331-51.
2. Hohmann EL. [Nontyphoidal salmonellosis](#). Clin. Infect. Dis. 2001; 32: 263-9.
3. [Surveillance for foodborne disease outbreaks - United States, 2008](#). In: MMWR Morb. Mortal. Wkly. Rep. United States, 2011: 1197-1202.

Table 1. Non-typhoidal *Salmonella* outbreaks caused by antimicrobial resistant strains (N=18), 2004–2008

Food Commodity	Year	No. of Cases	Serotype	Resistance Patterns ^{†, ‡}	Multistate Outbreak
<u>Land animal</u>					
Beef	2007	43	Newport	ACSSuTAuCxCfFox	Yes
Dairy	2004	100	Newport	ACSSuTAuCxCfFox	No
Dairy	2006	20	Typhimurium	ASuTAuCxCfFoxKan	No
Poultry	2004	24	Agona	SuT	No
Poultry	2004	42	Istanbul	T	No
Poultry	2005	4	Heidelberg	SSuGen	No
<u>Plants</u>					
Root	2006	3	Typhimurium	ACSSuT, ACSSuTAu	No
Vine-stalk	2006	84	Braenderup	ASuTGen, Gen	Yes
<u>Other</u>					
Complex	2005	25	Typhimurium	ACSSuTSxt, ACSuTSxt, ACSSuTAuSxt	No
Complex	2006	24	Newport	ACSSuTAuCxCfFox	No
Unknown	2004	2	Newport	ACSSuTAuCxCfFox, ACSSuTAuCxCfFoxKan	No
Unknown	2005	19	Heidelberg	STGen, STNal, ST	No
Unknown	2005	100	Typhimurium	ACSSuTAuCxCfFox	No
Unknown	2005	6	Schwarzengrund	AAuCxCfFox	No
Unknown	2006	9	Hadar	T, ST	No
Unknown	2006	14	I 4,[5],12:i:-	Nal	No
Unknown	2006	9	Newport	ACSSuTAuCxCfFox	No
Unknown	2007	11	Newport	ACSSuTAuCxCfFox	No

* Outbreaks were considered to be caused by a resistant isolate if ≥1 isolate was resistant to ≥1 antimicrobial agent

† A: ampicillin; Au: amoxicillin-clavulanic acid; C: chloramphenicol; Cf: ceftiofur; Cx: ceftriaxone; Fox: ceftiofur; Gen: gentamicin; Kan: kanamycin; Nal: nalidixic acid; S: streptomycin; Su: sulfonamide; Sxt: trimethoprim-sulfamethoxazole; T: tetracycline

‡ Multiple isolates from each outbreak were tested; all different resistance patterns observed are listed and separated by a comma

Table 2. Non-typhoidal *Salmonella* outbreaks caused by strains with no resistance detected (N=85), 2004–2008*

Food Commodity	Year	No. of Cases	Serotype	Multistate Outbreak	Food Commodity	Year	No. of Cases	Serotype	Multistate Outbreak
<u>Land animals</u>					<u>Other</u>				
Beef	2004	155	Berta	Yes	Complex	2004	31	Amager	No
Beef	2004	34	Typhimurium	Yes	Complex	2004	19	Enteritidis	No
Beef	2004	108	Anatum	No	Complex	2004	4	Heidelberg	No
Beef	2006	72	Montevideo	No	Complex	2004	4	I4,[5],12:i:-	No
Beef	2008	87	Newport	No	Complex	2004	12	I4,[5],12:i:-	No
Dairy	2005	3	Typhimurium	No	Complex	2005	24	Newport	No
Dairy	2006	4	Dublin	No	Complex	2005	34	Enteritidis	No
Dairy	2007	20	Montevideo	Yes	Complex	2005	57	Typhimurium	No
Eggs	2005	38	Enteritidis	No	Complex	2005	12	Enteritidis	No
Eggs	2005	23	Enteritidis	Yes	Complex	2005	5	Manhattan	No
Eggs	2006	113	Enteritidis	No	Complex	2005	34	Heidelberg	No
Eggs	2006	9	Enteritidis	No	Complex	2005	27	Enteritidis	Yes
Eggs	2007	81	Enteritidis	Yes	Complex	2005	26	Typhimurium	Yes
Pork	2006	55	Anatum	No	Complex	2006	161	Typhimurium	No
Pork	2007	31	Montevideo	No	Complex	2006	7	Typhimurium	No
Pork	2007	13	Infantis	No	Complex	2006	7	Typhimurium	No
Pork	2007	67	Newport	No	Complex	2007	16	Heidelberg	No
Poultry	2004	49	Newport	No	Complex	2007	46	Newport	No
Poultry	2004	21	Typhimurium	No	Complex	2007	33	Typhimurium	No
Poultry	2005	83	Enteritidis	No	Complex	2007	27	Enteritidis	No
Poultry	2006	22	Heidelberg	No	Complex	2007	87	Typhimurium	Yes
Poultry	2008	26	Saintpaul	Yes	Complex	2007	401	I4,[5],12:i:-	Yes
					Complex	2008	67	Muenchen	No
					Complex	2008	17	I4,[5],12:i:-	No
<u>Plants</u>									
Fruits-nuts	2005	157	Typhimurium	Yes	Complex	2008	101	Montevideo	No
Fruits-nuts	2006	715	Tennessee	Yes	Unclassifiable	2006	59	Oranienburg	No
Fruits-nuts	2006	41	Oranienburg	Yes	Unknown	2004	48	Agbeni	Yes
Fruits-nuts	2008	716	Typhimurium	Yes	Unknown	2004	66	Enteritidis	No
Fruits-nuts	2008	53	Litchfield	Yes	Unknown	2004	17	Typhimurium	No
Leafy	2004	97	Newport	Yes	Unknown	2004	4	Typhimurium	No
Leafy	2006	16	Javiana	No	Unknown	2004	10	Typhimurium	No
Leafy	2007	76	Typhimurium	Yes	Unknown	2005	95	Baildon	No
Sprout	2006	4	Braenderup	No	Unknown	2005	38	Newport	No
Sprout	2007	24	Montevideo	No	Unknown	2005	8	Typhimurium	No
Vine-stalk	2005	52	Newport	Yes	Unknown	2006	42	Enteritidis	No
Vine-stalk	2006	16	Berta	No	Unknown	2006	20	Typhimurium	No
Vine-stalk	2006	115	Newport	Yes	Unknown	2006	47	Heidelberg	No
Vine-stalk	2006	192	Typhimurium	Yes	Unknown	2006	5	Tallahassee	No
Vine-stalk	2008	1535	Saintpaul	Yes	Unknown	2006	9	Weltevreden	No
Vine-stalk	2008	61	Enteritidis	Yes	Unknown	2007	4	Newport	No
					Unknown	2007	7	Typhimurium	No
					Unknown	2007	6	Braenderup	No
<u>Aquatic animals</u>									
Finfish	2007	44	Paratyphi B Var. L(+) Tartrate+	Yes	Unknown	2008	8	Muenchen	No
					Unknown	2008	77	Typhimurium	Yes
					Unknown	2008	7	Poona	Yes
					Unknown	2008	6	Agona	Yes

* Outbreaks were considered to have no resistance detected if isolates were intermediate or susceptible to the antimicrobial agents tested by NARMS

Table 3. Number and percent of outbreaks caused by antimicrobial resistant non-typhoidal *Salmonella*, by agent and food commodity group* (N=18), 2004–2008

CLSI† Antimicrobial Class Antimicrobial Agent‡	Land animals (N=6)		Plants (N=2)		Complex or unclassifiable food (N=2)		Unknown Food (N=8)		Total (N=18)	
	n	(%)	n	(%)	n	(%)	n	(%)	n	(%)
	Aminoglycosides									
Amikacin	0	(0)	0	(0)	0	(0)	0	(0)	0	(0)
Gentamicin	1	(17)	1	(50)	0	(0)	1	(13)	3	(17)
Streptomycin	2	(34)	1	(50)	2	(100)	6	(75)	11	(61)
Kanamycin	1	(17)	0	0	0	(0)	1	(13)	2	(11)
β-lactam/β-lactamase inhibitor combinations										
Amoxicillin-clavulanic acid	3	(50)	1	(50)	2	(100)	5	(63)	11	(61)
Cephems										
Ceftriaxone	3	(50)	0	(0)	1	(50)	5	(63)	9	(50)
Ceftiofur	3	(50)	0	(0)	1	(50)	5	(63)	9	(50)
Cefoxitin	3	(50)	0	(0)	1	(50)	5	(63)	9	(50)
Penicillins										
Ampicillin	3	(50)	2	(100)	2	(100)	4	(50)	11	(61)
Quinolones										
Ciprofloxacin	0	(0)	0	(0)	0	(0)	0	(0)	0	(0)
Nalidixic acid	0	(0)	0	(0)	0	(0)	2	(25)	2	(11)
Folate pathway inhibitors										
Sulfamethoxazole/Sulfisoxazole§	5	(83)	2	(100)	2	(100)	4	(50)	13	(72)
Trimethoprim-sulfamethoxazole	0	(0)	0	(0)	1	(50)	0	(0)	1	(6)
Phenicol										
Chloramphenicol	2	(33)	1	(50)	2	(100)	4	(50)	9	(50)
Tetracyclines										
Tetracycline	5	(83)	2	(100)	2	(100)	6	(75)	15	(83)

* No outbreaks caused by resistant strains were attributed to aquatic animals

† CLSI: Clinical and Laboratory Standards Institute

‡ Antimicrobial agent categories are not mutually exclusive; outbreaks can be caused by strains resistant to multiple antimicrobial agents

§ Sulfamethoxazole was replaced by sulfisoxazole during 2004

Table 4. Antimicrobial resistance patterns of non-typhoidal *Salmonella* outbreak strains, by commodity group (N=103), 2004–2008

Resistance Pattern†	Simple food commodity*				Complex or unclassifiable				Overall (N=103)	
	Land animals (N=28)		Plants (N=18)		food (N=28)		Unknown Food (N=28)			
	n	(%)	n	(%)	n	(%)	n	(%)		
No resistance detected	22	(79)	16	(89)	26	(93)	20	(71)	85	(83)
Resistant to 1–4 agents	3	(11)	1	(6)	0	(0)	3	(11)	7	(7)
Resistant to ≥5 agents	3	(11)	1	(6)	2	(7)	5	(18)	11	(11)
At least ACSSuT‡	1	(4)	1	(6)	2	(7)	4	(14)	8	(8)
At least ACT/S§	0	(0)	0	(0)	1	(4)	0	(0)	1	(1)
At least ACSSuTAuCx¶	1	(4)	0	(0)	1	(4)	4	(14)	6	(6)

* No resistance was detected in one outbreak associated with an aquatic animal

† ACSSuT, ACT/S, and ACSSuTAuCx resistance patterns are not mutually exclusive; outbreaks can be categorized into multiple patterns

‡ ACSSuT: resistant to ampicillin, chloramphenicol, streptomycin, sulfamethoxazole/sulfisoxazole, tetracycline

§ ACT/S: resistant to ampicillin, chloramphenicol, trimethoprim-sulfamethoxazole

¶ ACSSuTAuCx: resistant to ACSSuT, amoxicillin-clavulanic acid, ceftriaxone

Appendix B – Criteria for Retesting of Isolates

Repeat testing of an isolate must be done when one or more of the following conditions occur:

- No growth on panel
- Growth in all wells
- Multiple skip patterns
- Apparent contamination in wells or isolate preparation
- Unlikely or discordant susceptibility results (Table 1)

If an isolate is retested, data for all antibiotics should be replaced with the new test results. *Categorical changes may require a third test (and may indicate a mixed culture).*

Uncommon test results (Table 2) may represent emerging resistance phenotypes. Retesting is encouraged.

Table 1. Retest criteria for unlikely or discordant resistance phenotypes

Organism(s)	Resistance phenotype	Comments
<i>Salmonella</i> and <i>E. coli</i>	nalidixic acid ^S (≤ 16) AND ciprofloxacin ^R (≥ 4)	The stepwise selection of mutations in the QRDR* does not support this phenotype
	ceftiofur ^R (≥ 8) AND ampicillin ^S (≤ 8)	The presence of an ESBL† or AmpC beta-lactamase should confer resistance to ampicillin.
	ceftiofur ^R (≥ 8) AND ceftriaxone ≤ 1	
	ampicillin ^S (≤ 8) AND amoxicillin-clavulanic acid ^R ($\geq 32/16$)	
	sulfisoxazole ^S (≤ 256) AND trimethoprim-sulfamethoxazole ^R ($\geq 4/76$)	
<i>Campylobacter</i>	erythromycin ^S (≤ 8) AND azithromycin ^R (≥ 8)	Erythromycin is class representative for 14- and 15-membered macrolides (azithromycin, clarithromycin, roxithromycin, and dirithromycin)
	erythromycin ^R (≥ 32) AND azithromycin ^S (≤ 2)	
	nalidixic acid ^S (≤ 16) AND ciprofloxacin ^R (≥ 4)	In <i>Campylobacter</i> , one mutation is sufficient to confer resistance to both nalidixic acid and ciprofloxacin
	nalidixic acid ^R (≥ 64) AND ciprofloxacin ^S (≤ 1)	

* quinolone resistance-determining regions

† extended-spectrum beta-lactamase

Table 2. Uncommon resistance phenotypes for which retesting is encouraged

Organism(s)	Resistance phenotype
<i>Salmonella</i> and <i>E. coli</i>	Pan-resistance
	Resistance to amikacin (≥ 64), ceftriaxone and/or ceftiofur MIC ≥ 2 AND ciprofloxacin ≥ 0.125 and/or nalidixic acid ≥ 32
<i>Campylobacter</i>	Pan-resistance
	Resistance to gentamicin (≥ 8)
	Not susceptible to florfenicol (≥ 8)